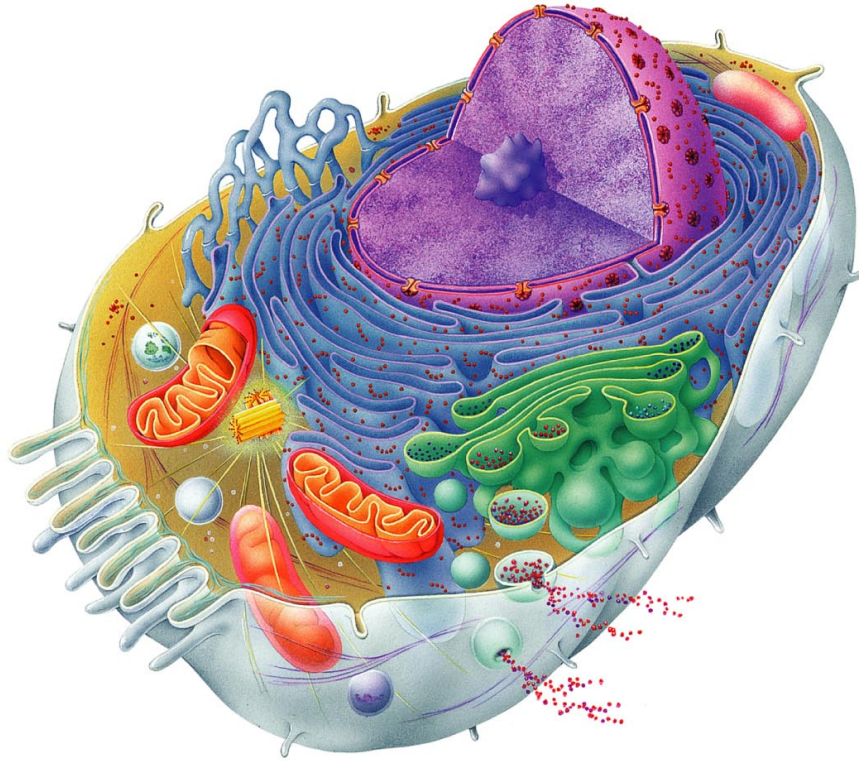
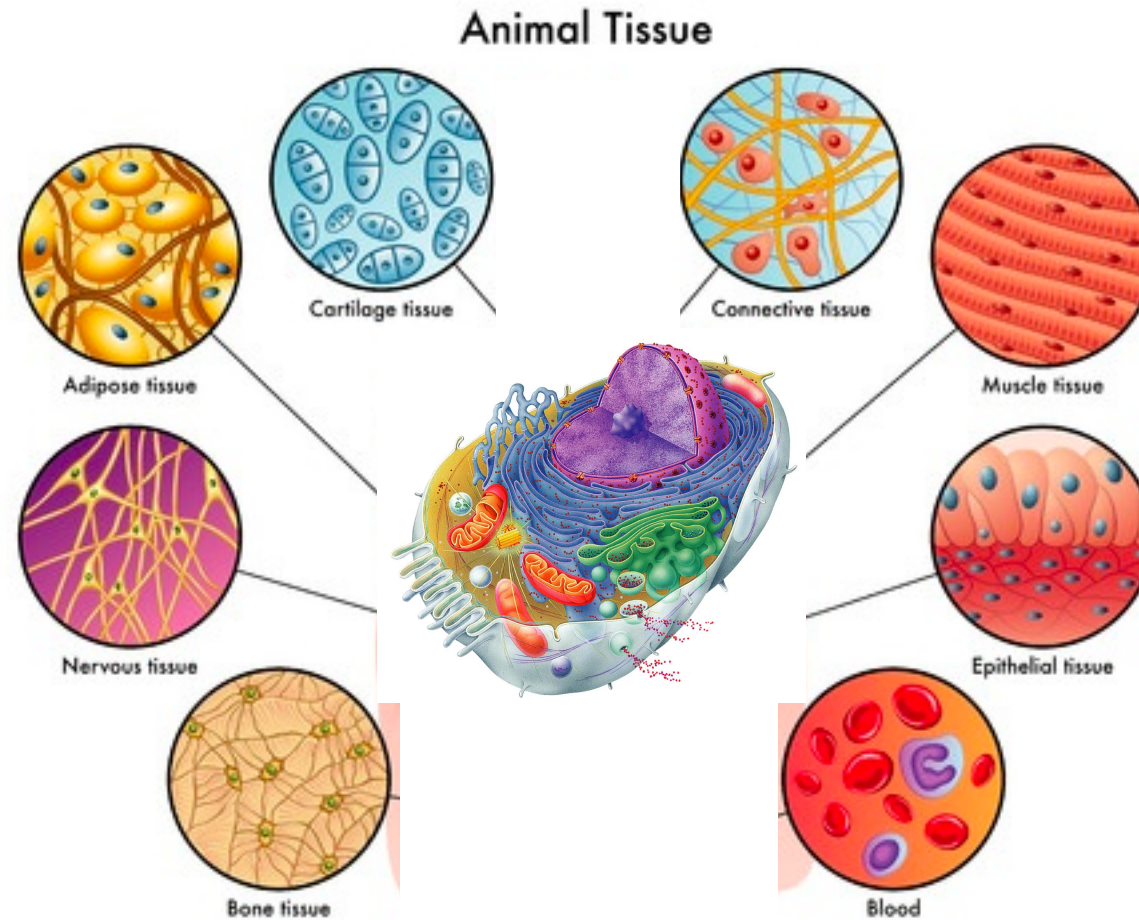


Cells - the autonomous unit of life

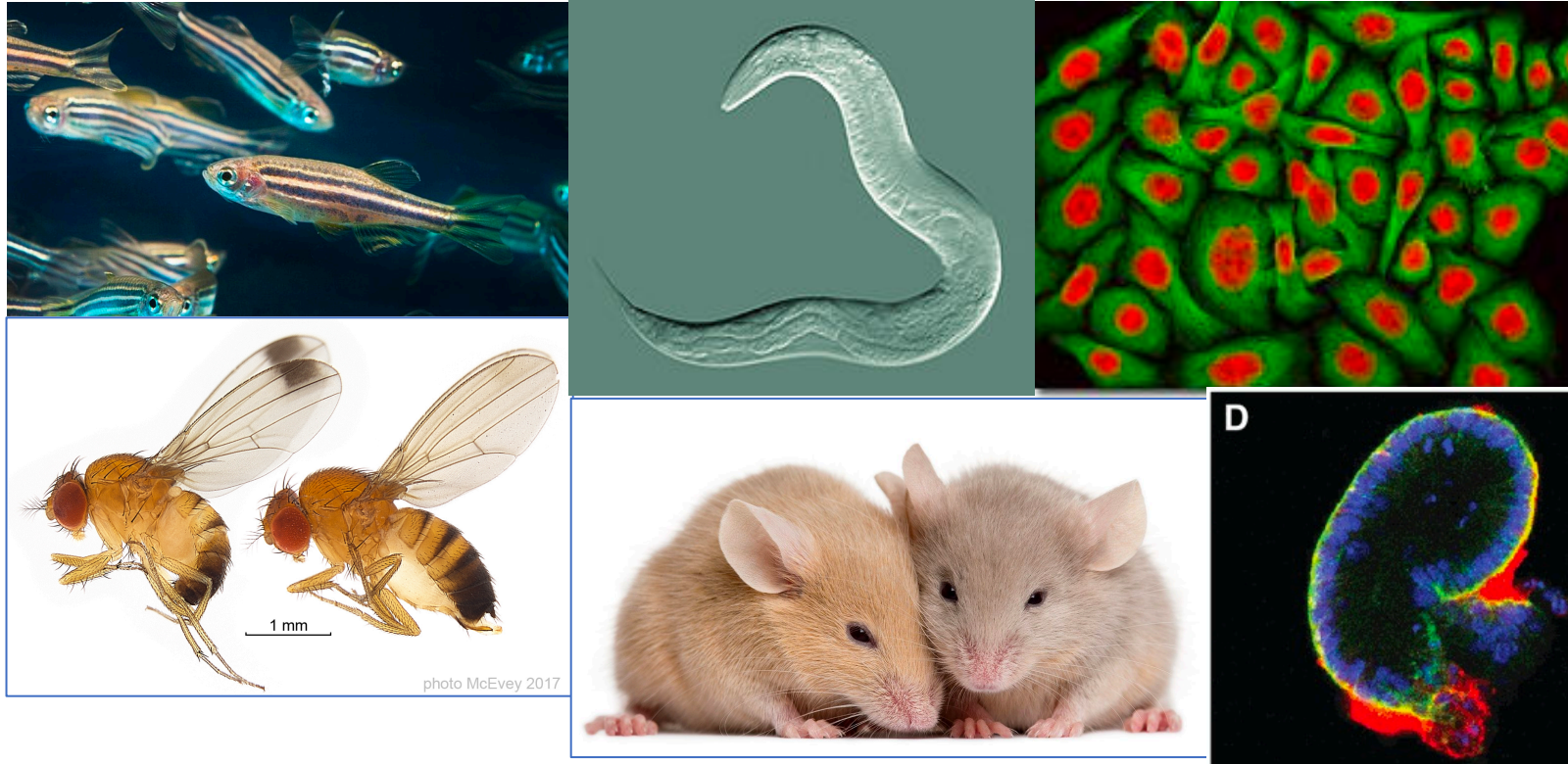


The mission - understanding how cells function

Cells come in different flavors



Model systems in cell biology

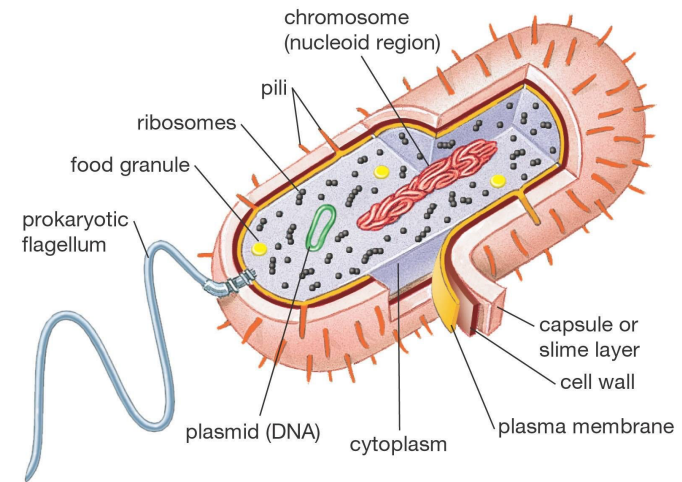


Advantages:

- 1. Easy to grow.*
- 2. Powerful genetics.*
- 3. Suitable for experimental techniques.*

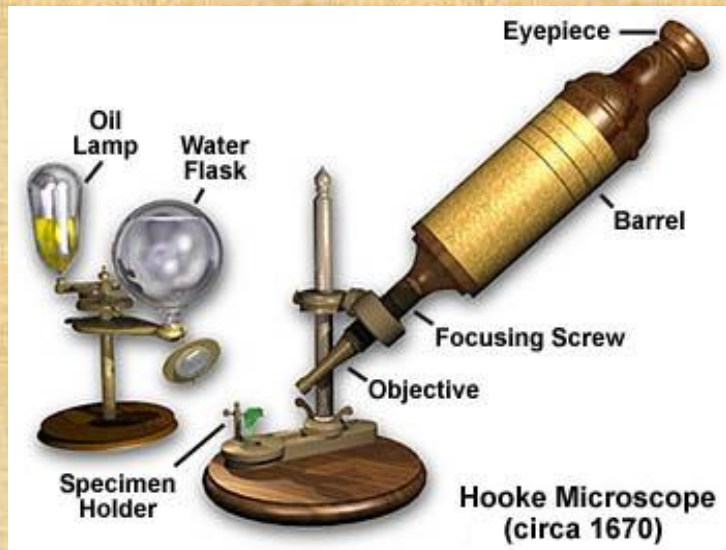
Documentation of cellular processes

1. All **three dimensions** (x, y, z) should be documented simultaneously.
2. Image acquisition **speed** has to be greater than the speed of the biological process.
3. Spatial **resolution** of the microscope should match the scale of the object examined.
4. It must be possible to **track individual components**.
5. Process should be imaged under **physiological conditions**.

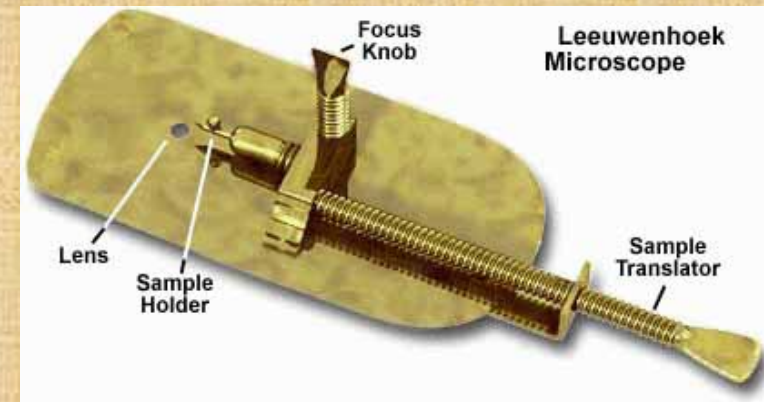
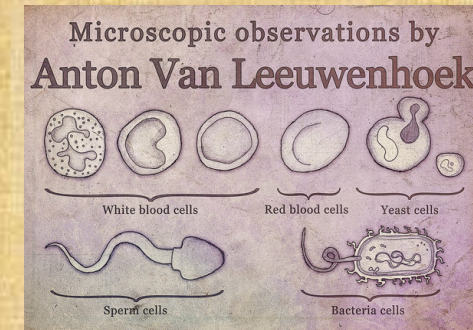


Cells and microscopes

Robert Hooke

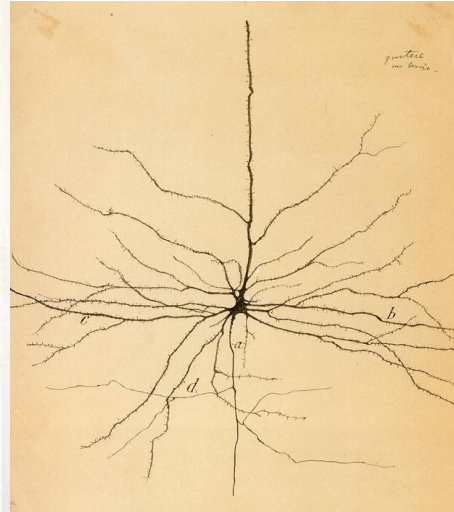
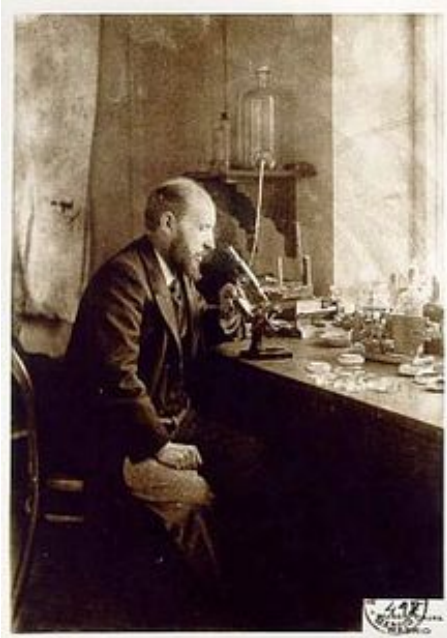


Anton Van Leeuwenhoek



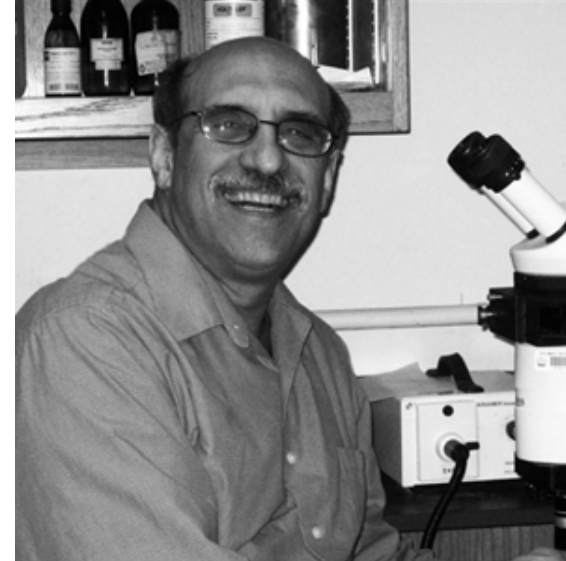
Cells and microscopes

Ramon y cajal



1890 – Describing the nervous system using specific staining (golgi staining).

Martín Chalfie



1995 – GFP in *C. elegans*

The Royal Swedish Academy of Sciences has decided to award the Nobel Prize in Chemistry for 2008 jointly to:

Osamu Shimomura, Martin Chalfie, and
Roger Y. Tsien,

"for the discovery and development of the green fluorescent protein, GFP".

The discovery and development of the green fluorescent protein (GFP) have radically changed the scientific agenda. Improved variants of GFP and GFP-like proteins in synergy with high-resolution microscopes, computational techniques and powerful theoretical approaches are currently fuelling a scientific revolution focused on quantitative analyses of complex biological systems. **The gradual appearance of a world of hitherto unseen structures and dynamic principles is now impacting virtually all aspects of biological, medical and pharmaceutical research.** (presentation speech, 2008)

Fluorescence super resolution microscopy



Nobel Prize 2014, Chemistry



Eric Betzig



William Moerner

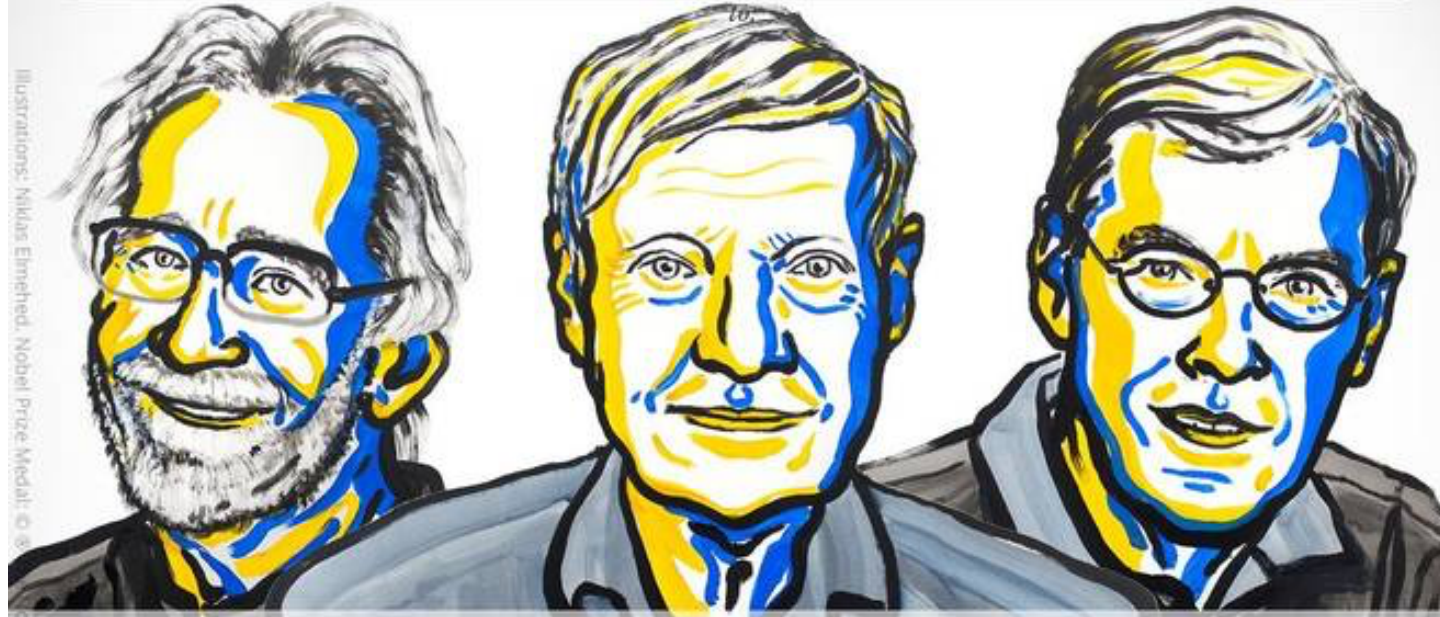


Stefan Hell

"for the development of super-resolved fluorescence microscopy".

The Royal Swedish Academy of Sciences has decided to award the

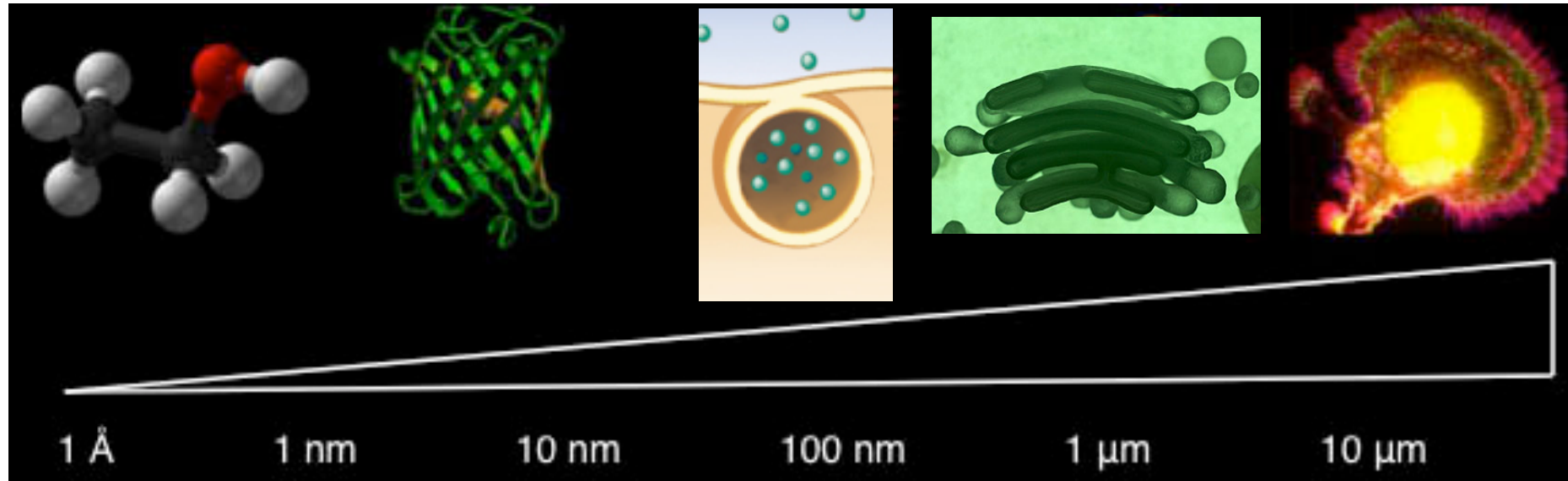
2017 NOBEL PRIZE IN CHEMISTRY



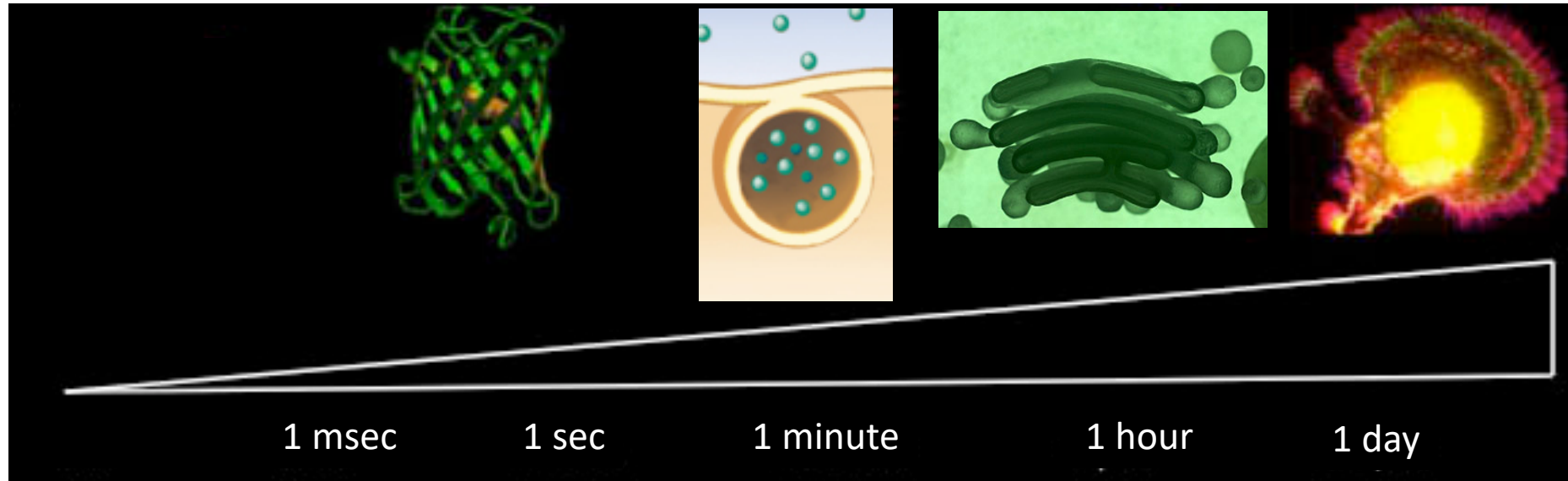
Jacques Dubochet
Joachim Frank
Richard Henderson

"for developing cryo-electron microscopy for the high-resolution structure determination of biomolecules in solution"

The cellular ruler - size



The cellular ruler - time



Diffusion of cytosolic proteins: msec

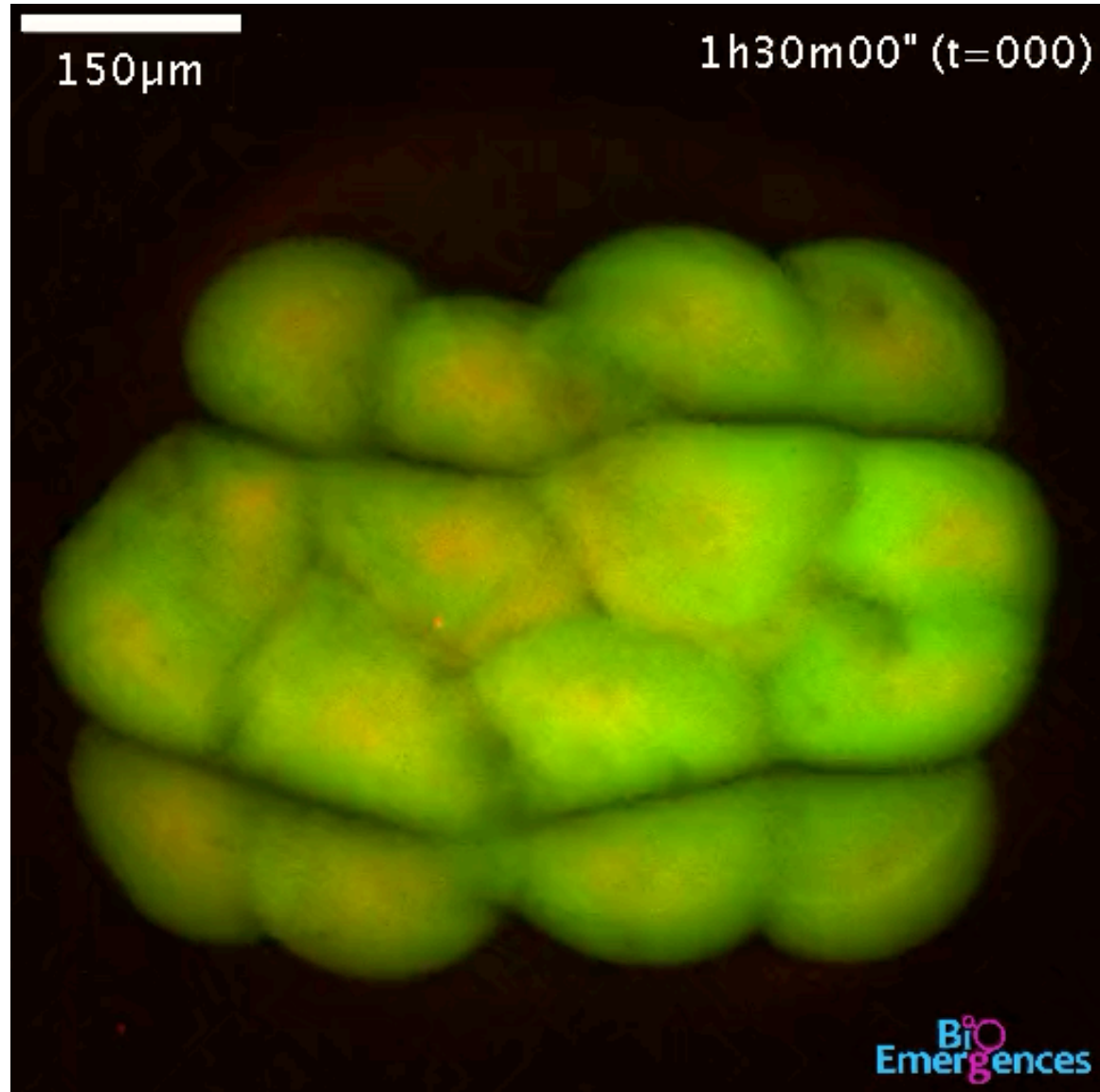
Diffusion of TM proteins: sec

Endocytosis: minute

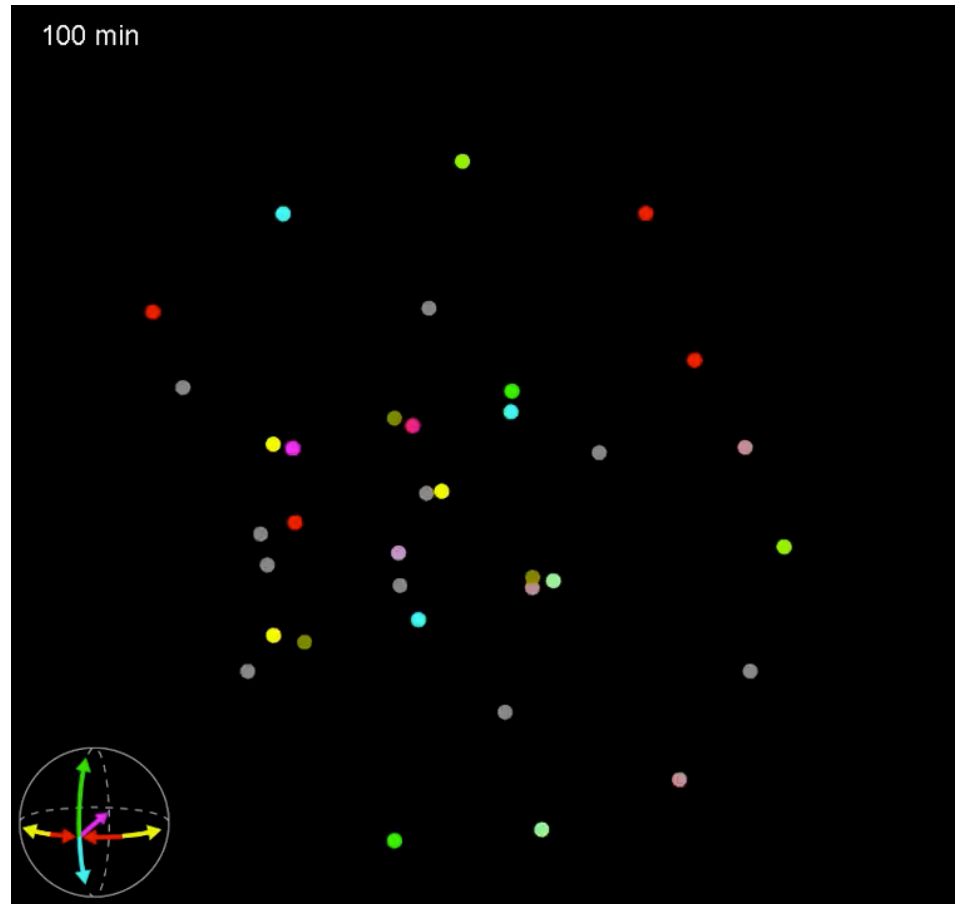
Trafficking in the secretory pathway Golgi-PM: hour

Cell cycle: day

Fish embryos

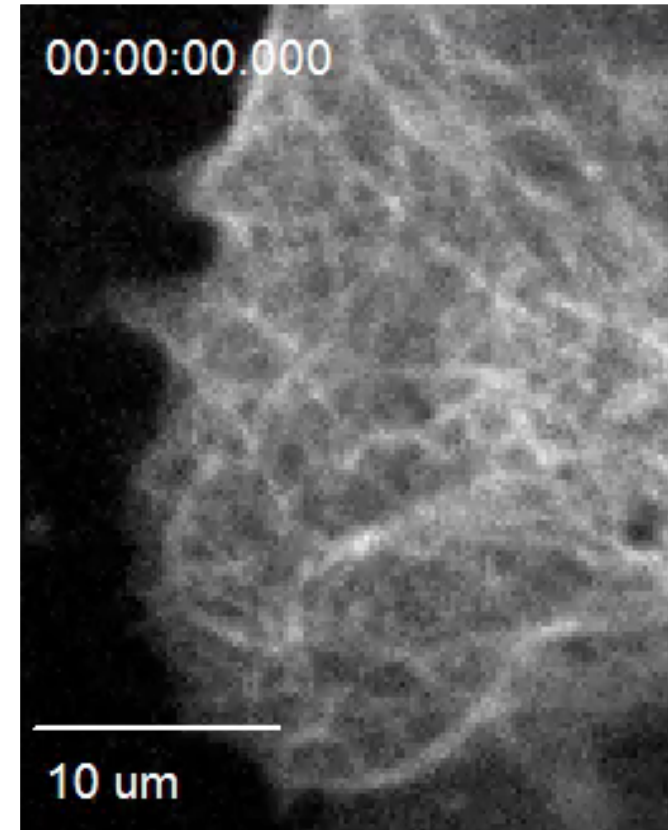
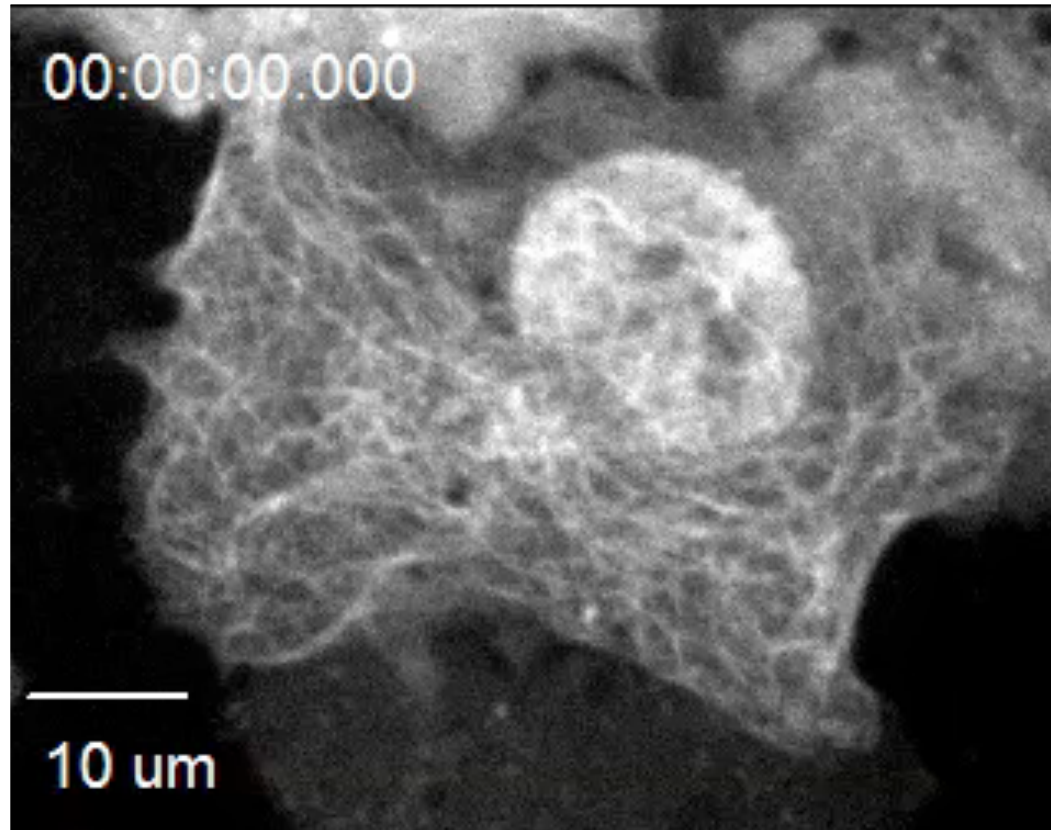


Fish embryos



Zebrafish development, cell tracking, janelia farms

Intracellular dynamics - microtubules



Intracellular dynamics

Part I

Tubular ER generation by hitchhiking on a moving mitochondrion

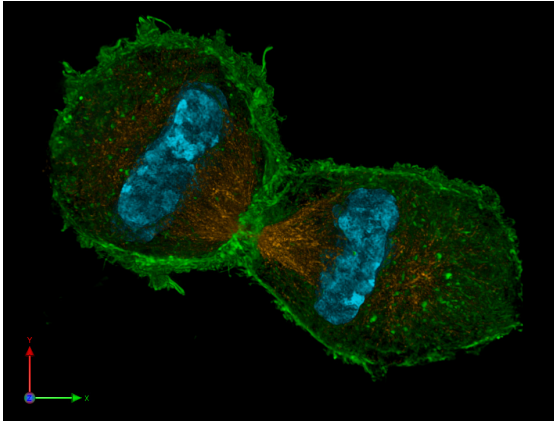
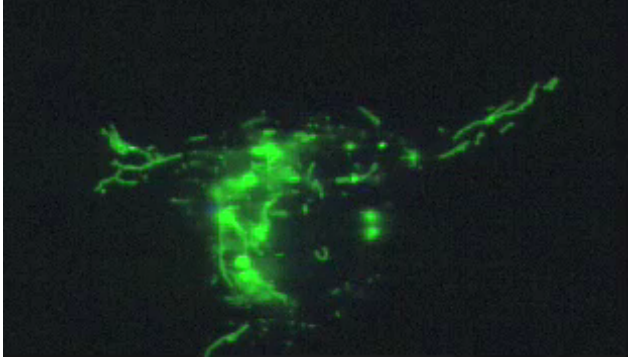
Example 1

97-nm resolution and 266 frames/s over thousands of time points

Cell, 2018

Imaging the structure of a cell

Fluorescence microscopy

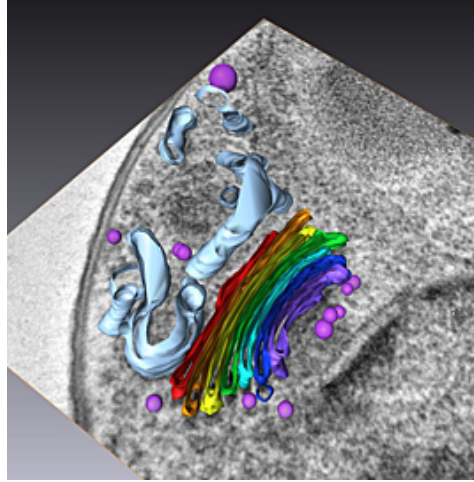


Structure and dynamics of selected protein complexes in cells in 3D.

Limitations: results depend on the fluorescent probe.

Resolution: up to 30 nm.

Electron microscopy

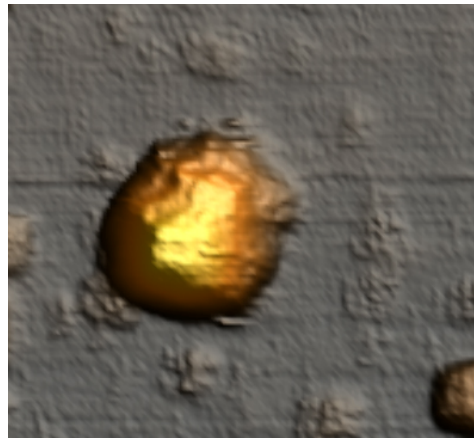


3D reconstruction of a cell and of large protein complexes.

Limitations: Difficult to label specific proteins, no dynamics.

Resolution: 0.1 nm in cells

AFM

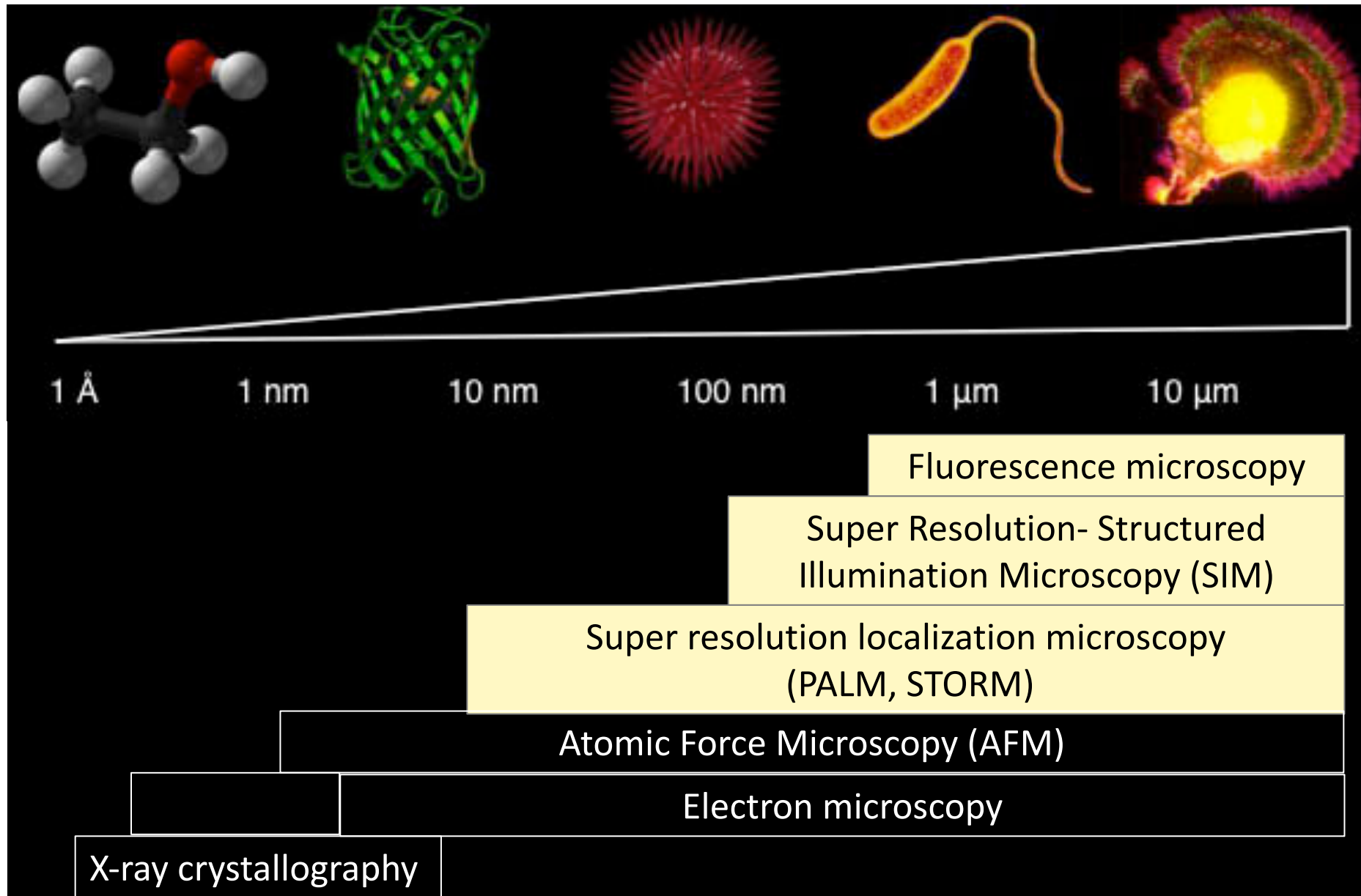


Mechanical properties of cells and proteins in living systems.

Limitations: limited to cell surface.

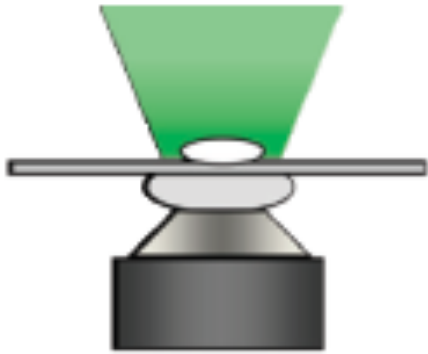
Resolution: sub nm

Bridging the resolution gap

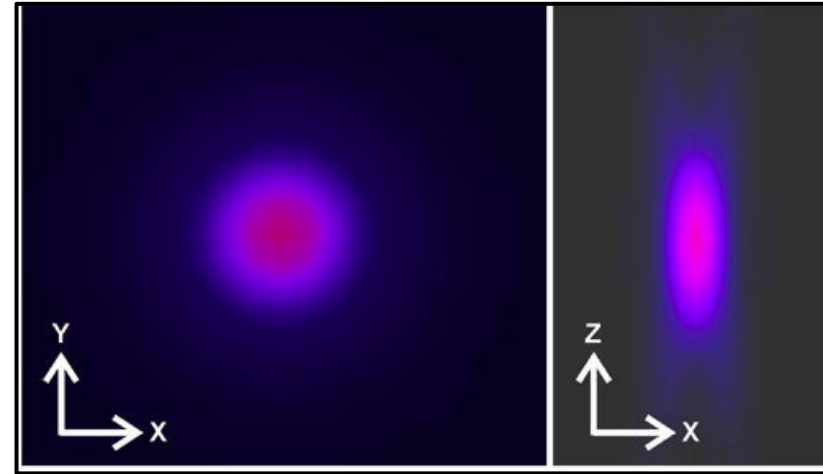




PSF (point spread function)



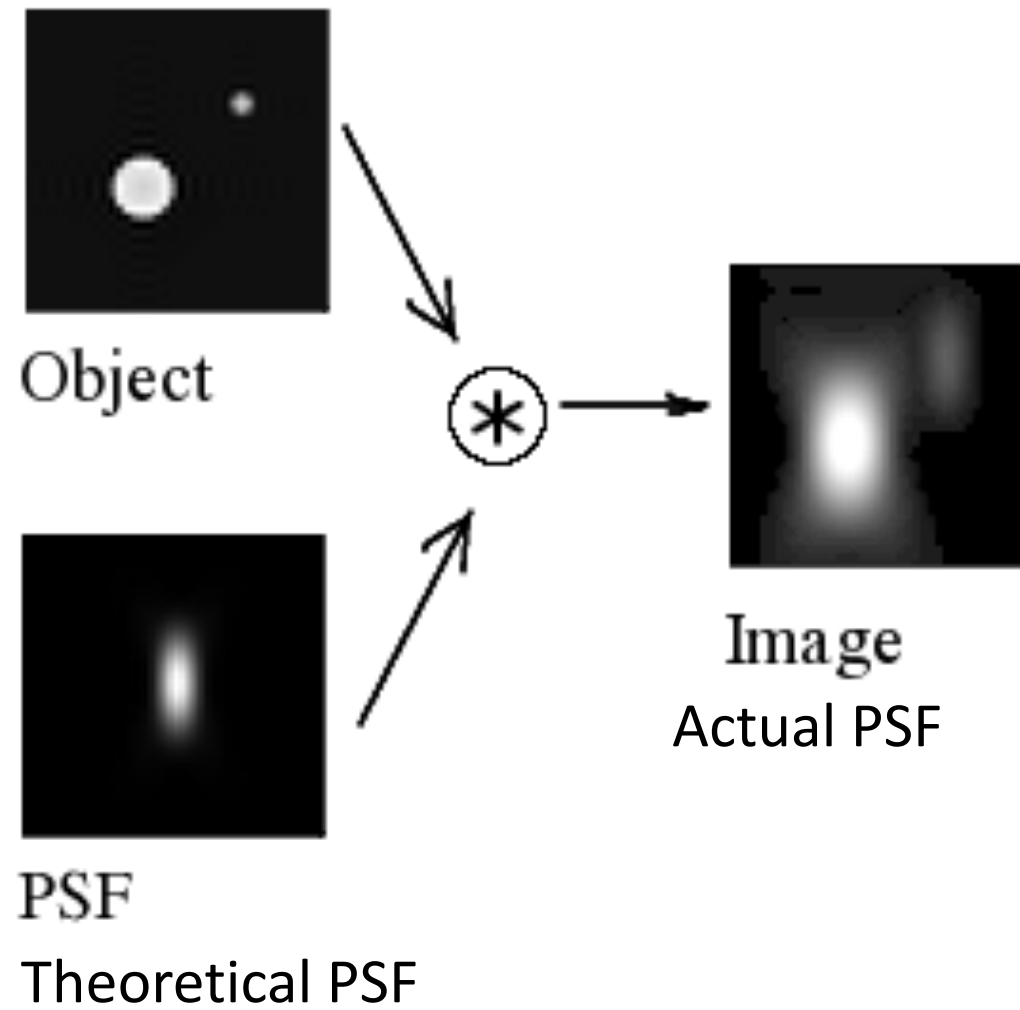
Wide Field



The objective lens does not focus the emitted light to an infinitely small point in the image plane. Light waves converge and interfere at the focal point.

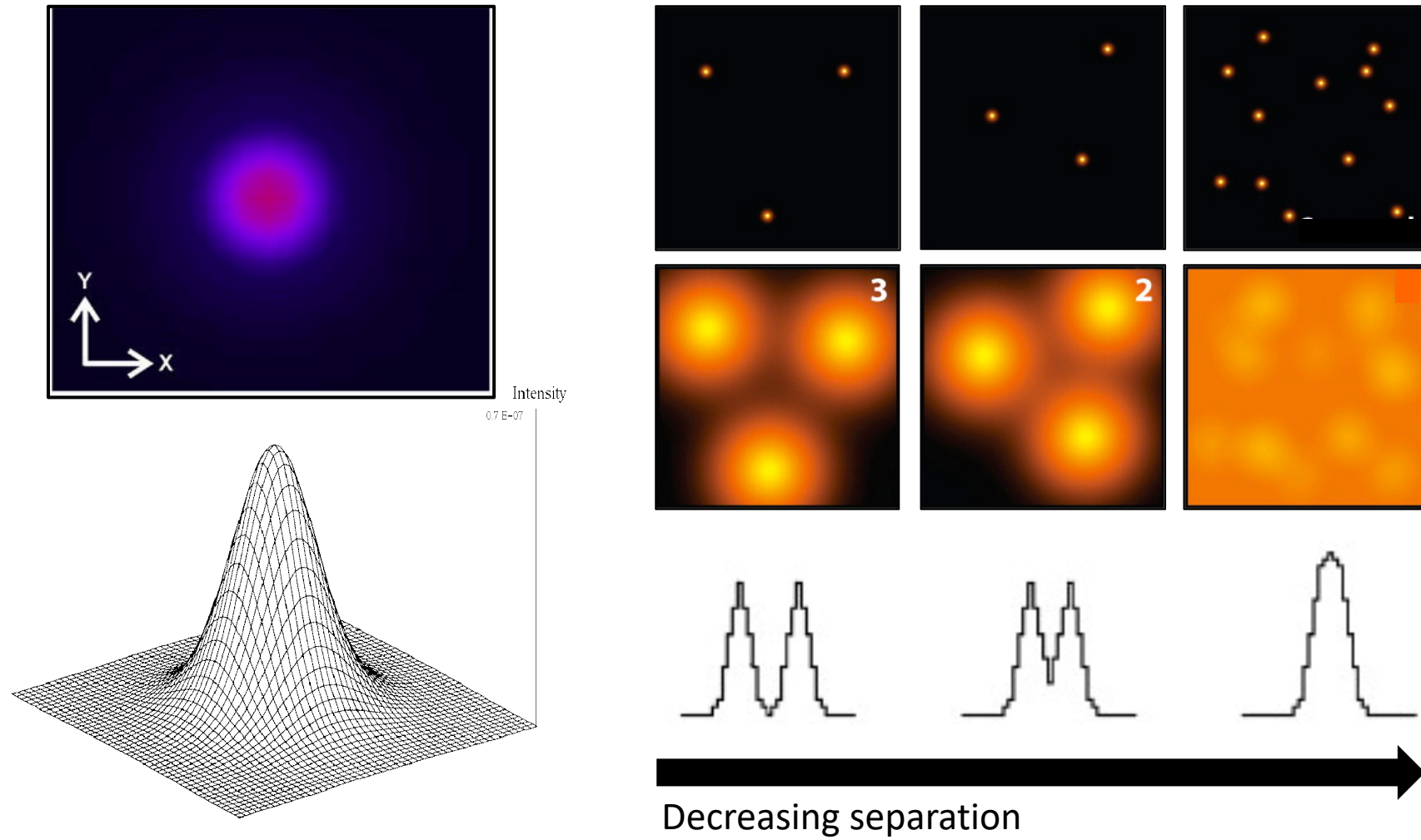
PSF depends on the NA of the objective.
The higher the NA the smaller the PSF.

PSF



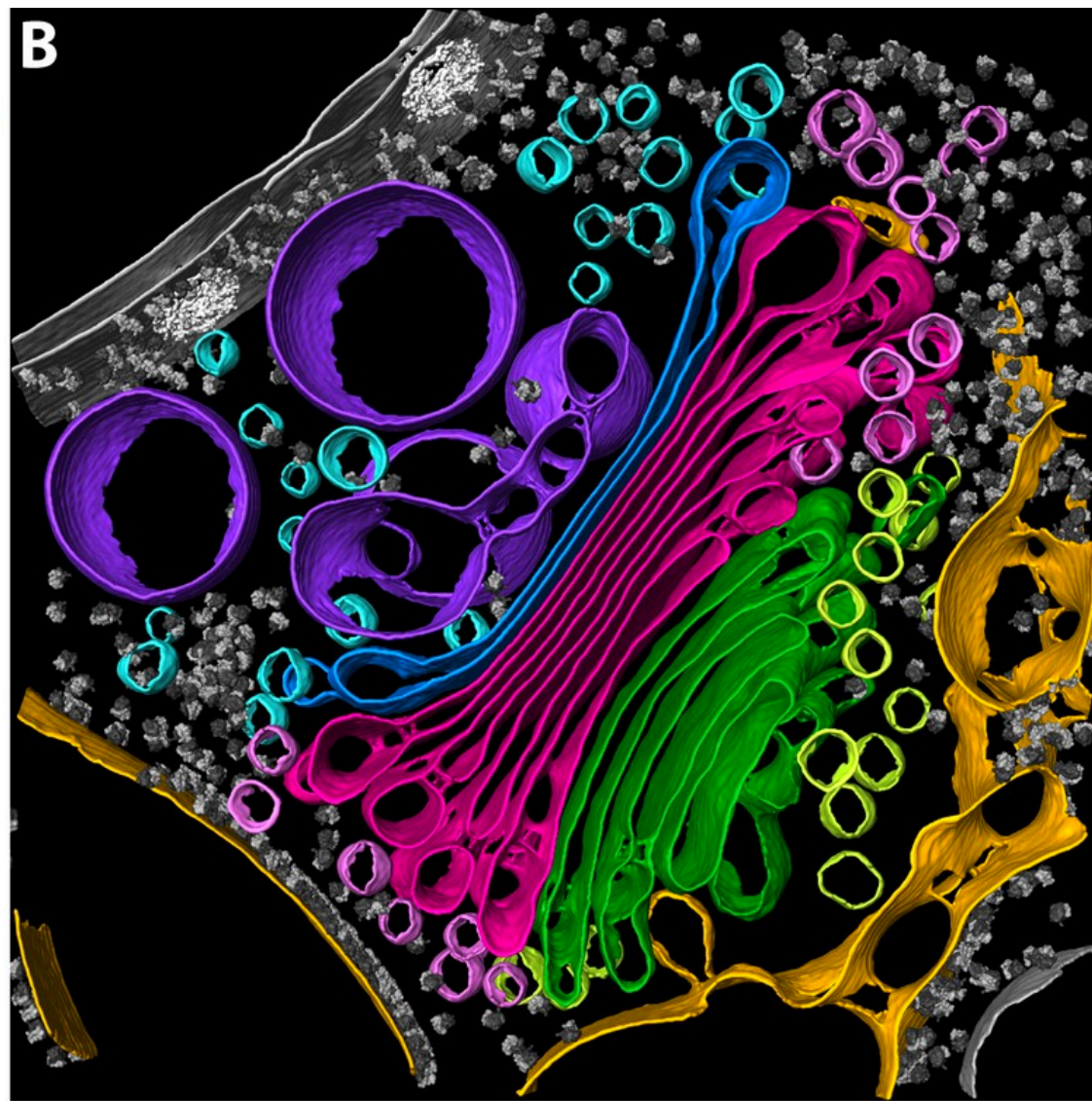
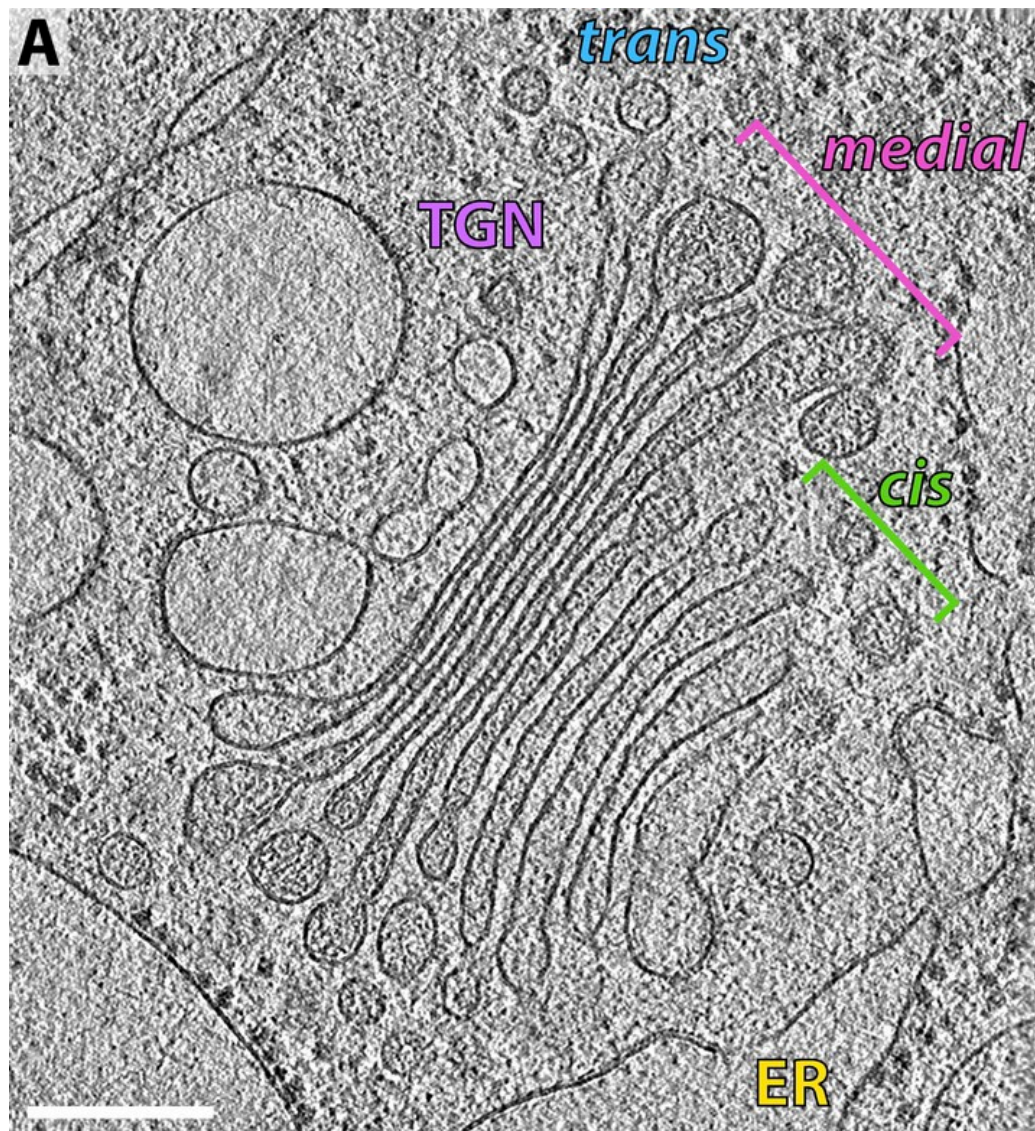
It is important to measure the actual PSF for each objective

Resolution



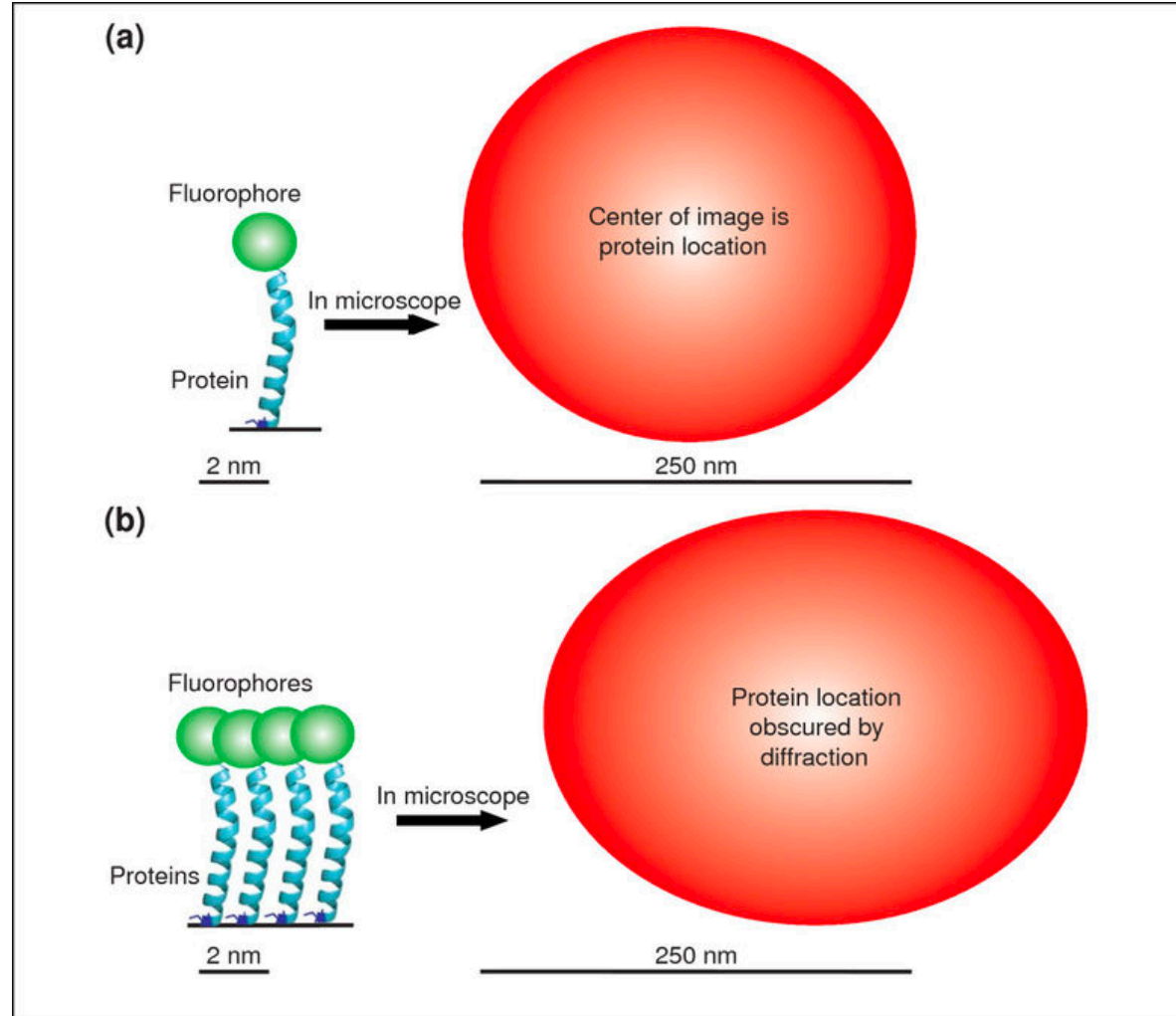
$$\text{Resolution} = \lambda / 2NA$$



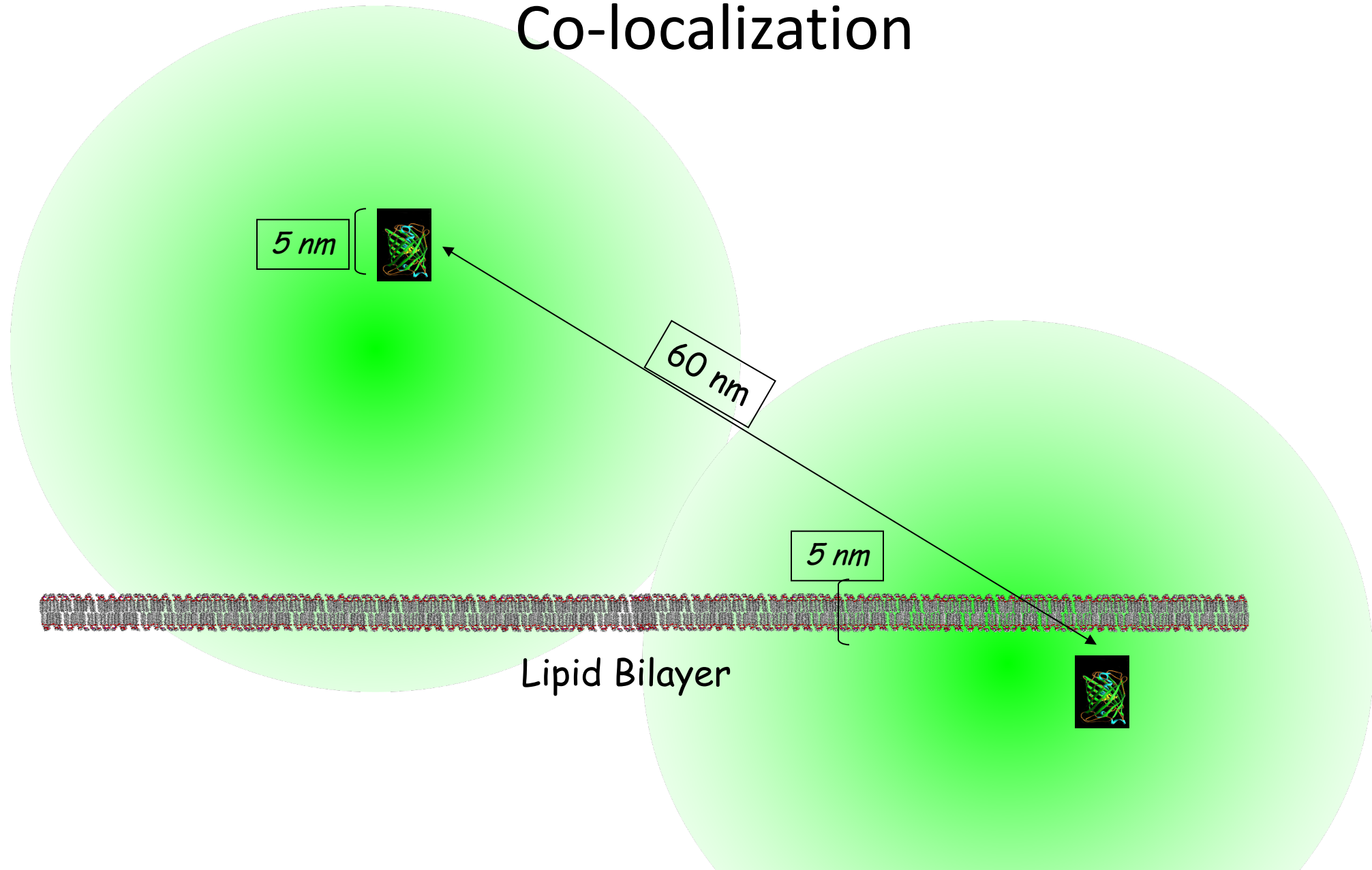


Issues associated with limited resolution:

Quantitation

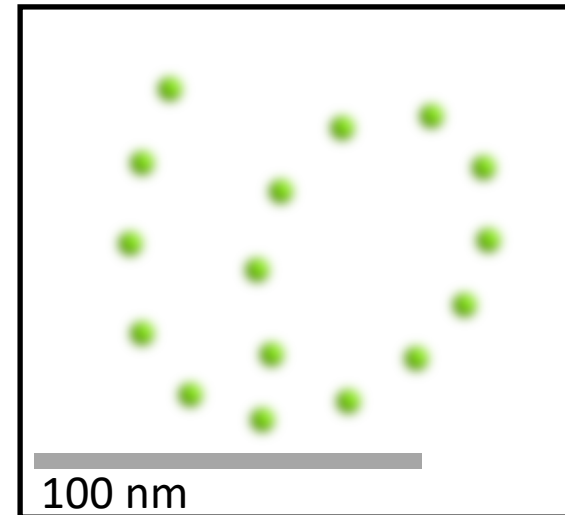
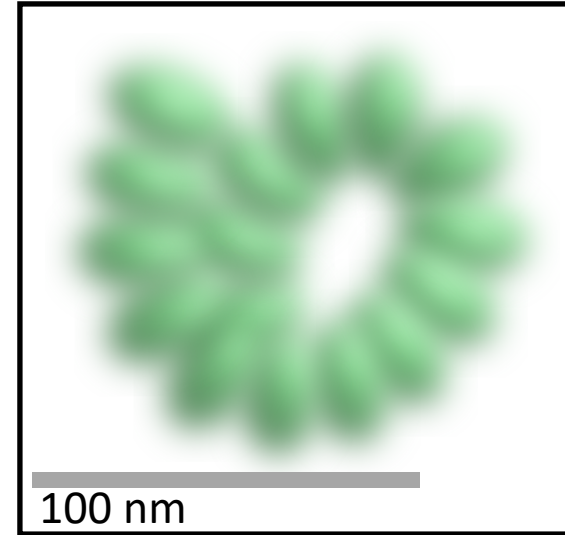
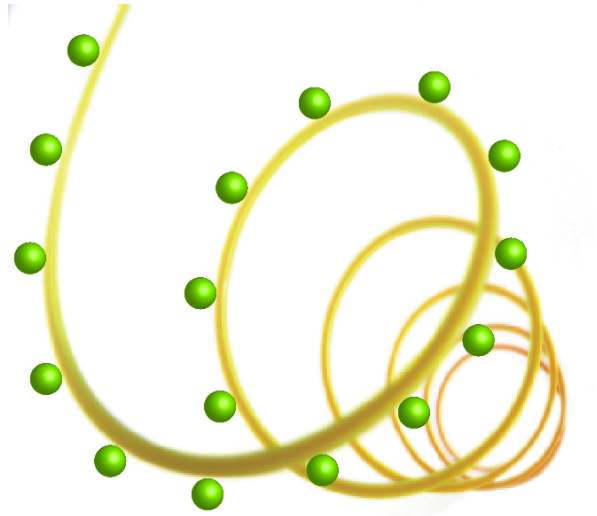


Issues associated with limited resolution: Co-localization



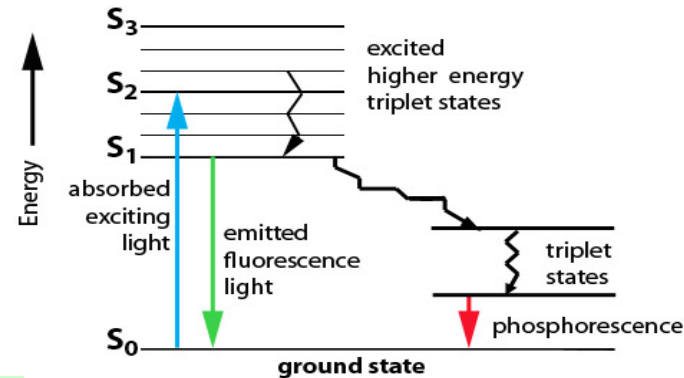
Issues associated with limited resolution:

Resolving a structure

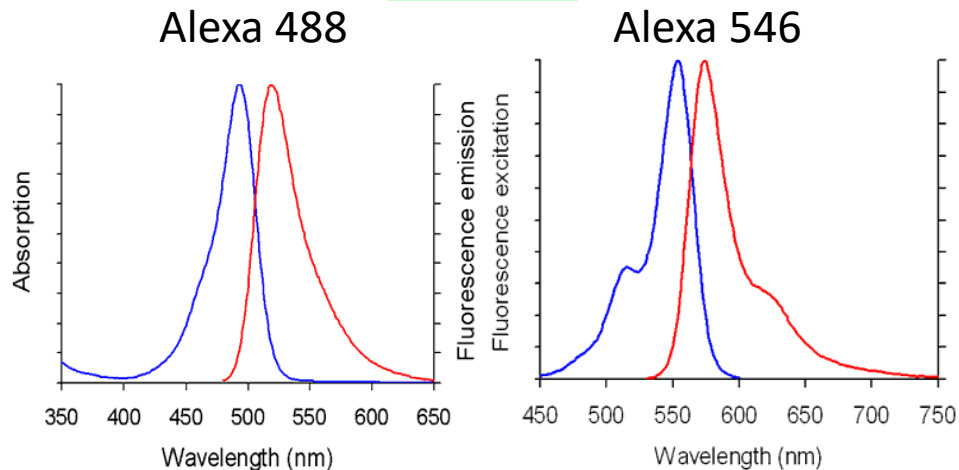


Fluorescence Microscopy: How does it work?

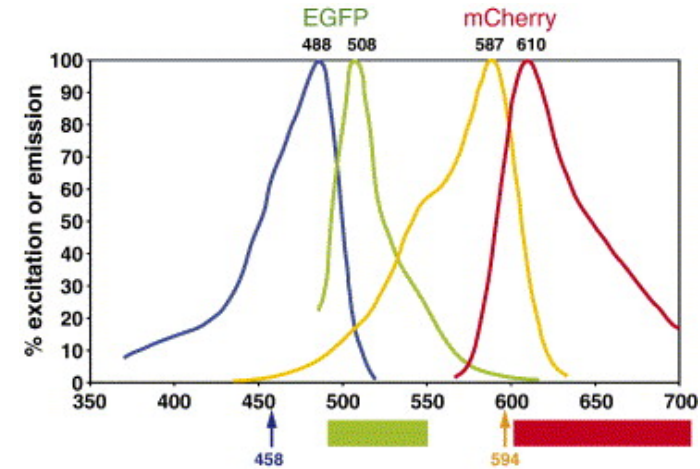
The high energy absorbed in the excited state is being released as fluorescent light.



Fl- dyes

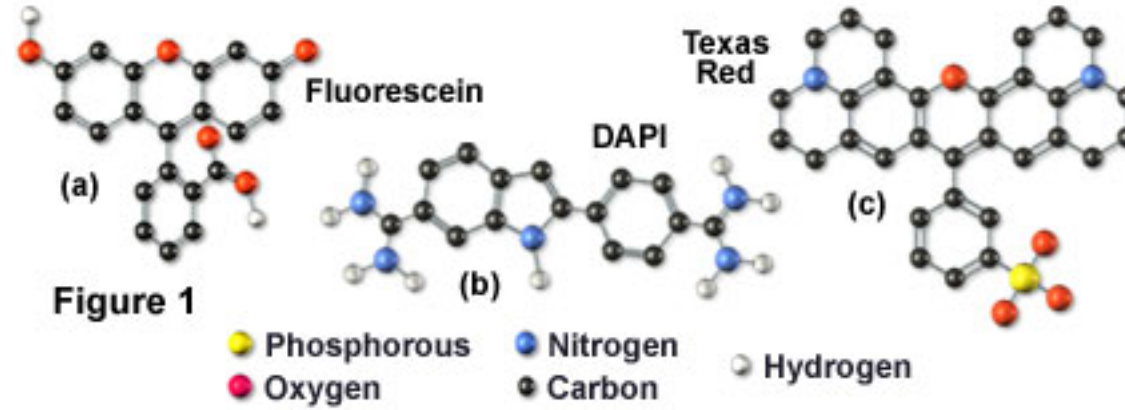


Fl- Proteins

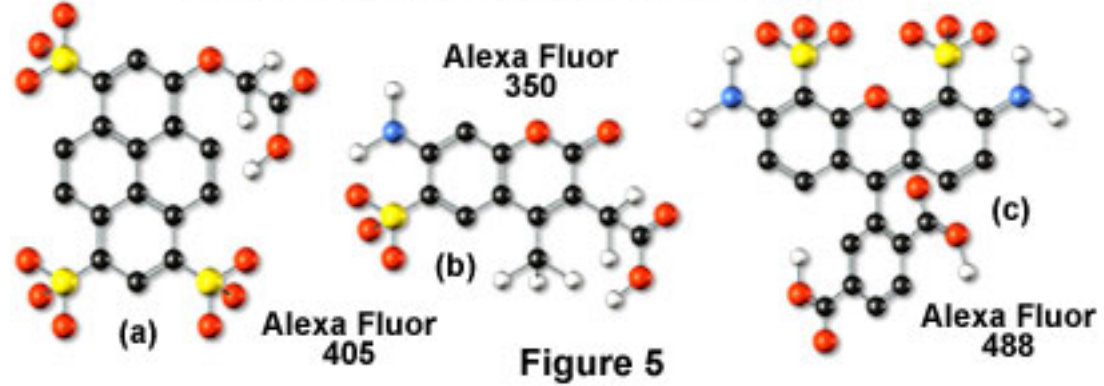


Fluorophores

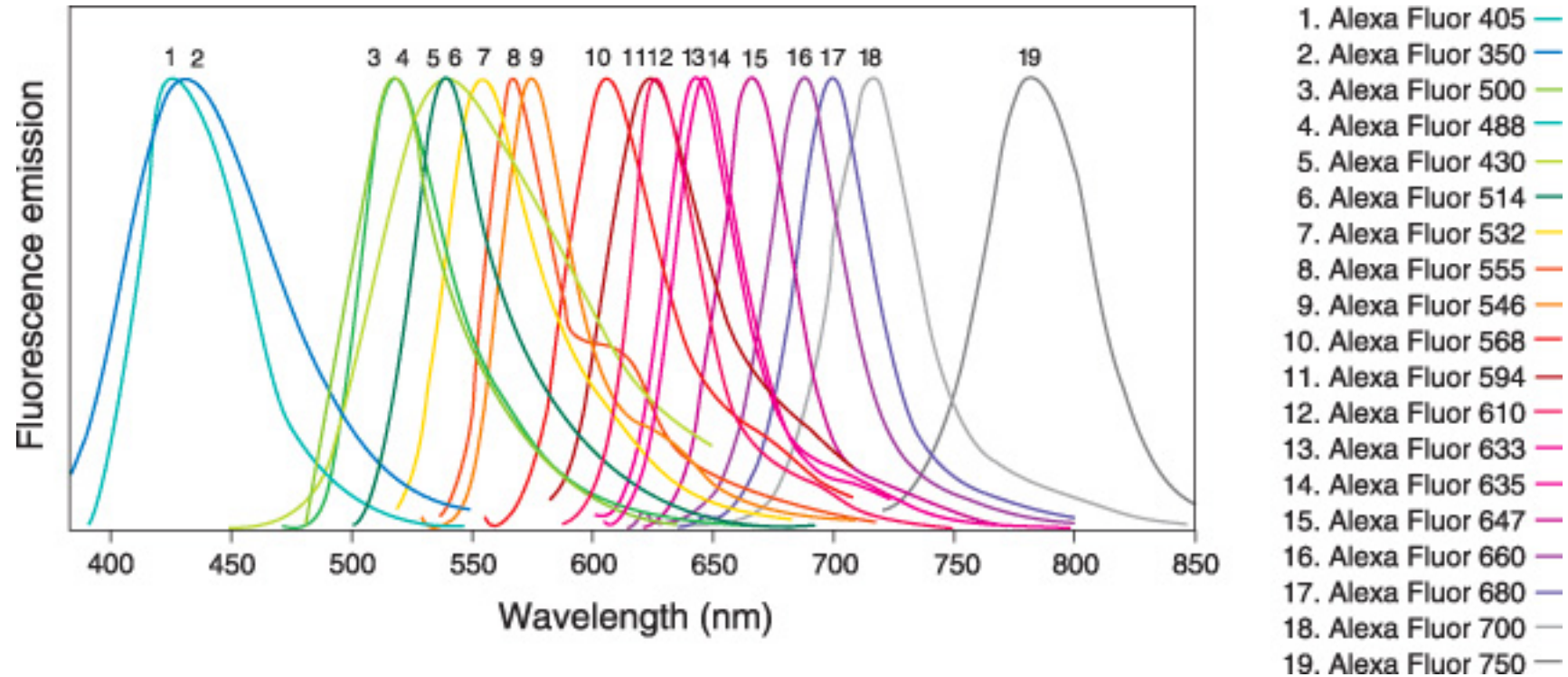
Common Fluorophores in Widefield and Confocal Microscopy



Alexa Fluor Synthetic Fluorochromes

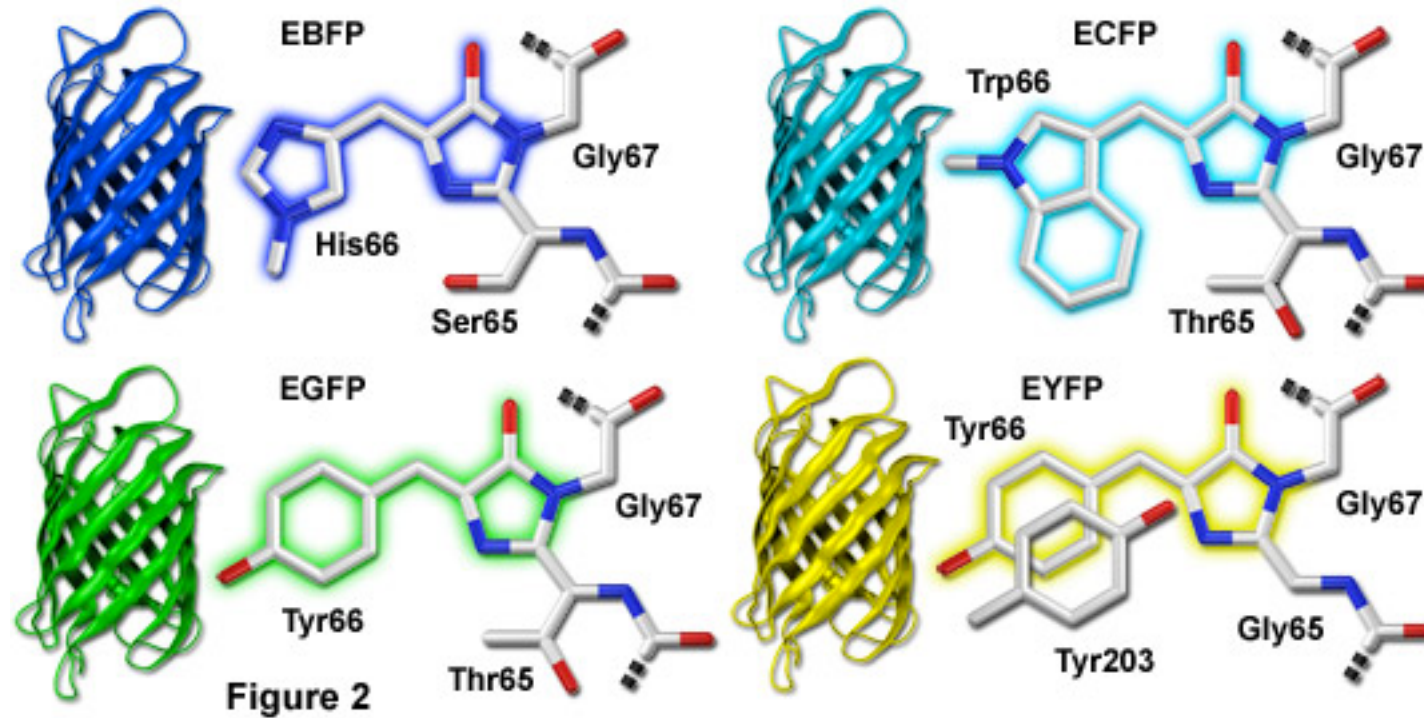


Fluorophores

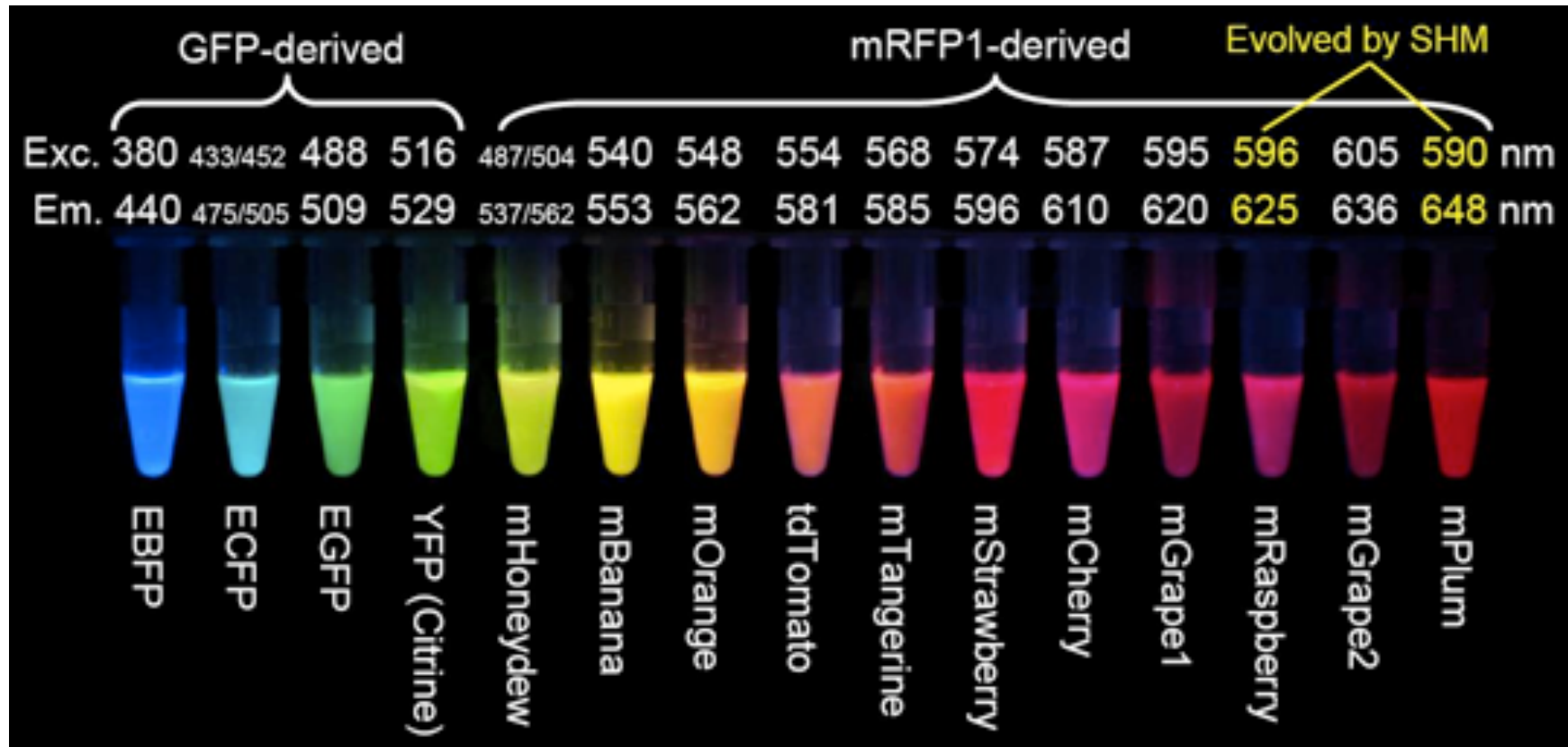


Fluorescent proteins

Chromophore Structural Motifs of Green Fluorescent Protein Variants



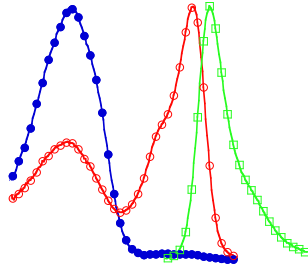
Fluorescent proteins



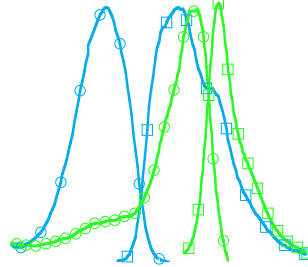
Photoactivatable Fluorescent Proteins

Irreversible

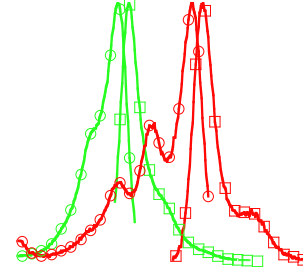
PAGFP



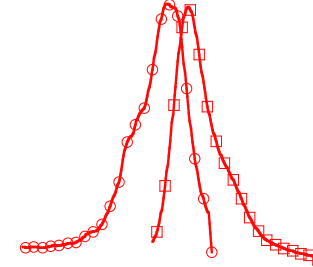
PS-CFP



Kaede
KikGR
EosFP
Dendra

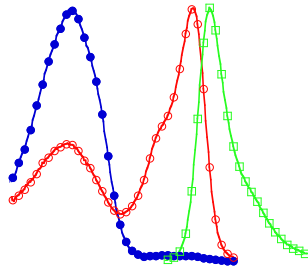


PAmRFP

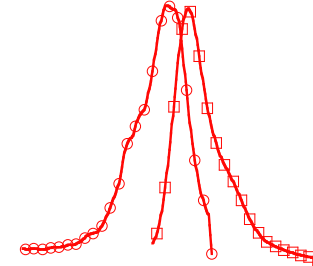


Reversible

Dronpa
rsFastlime
Padron

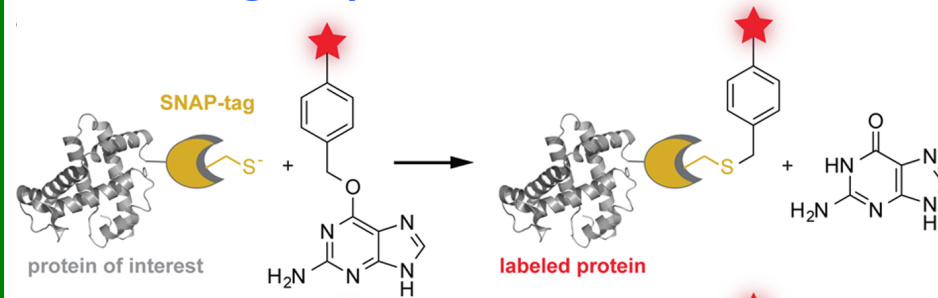


KFP1
rsCherry
rsCherryRev



Labeling proteins in live cells with Fl-dyes

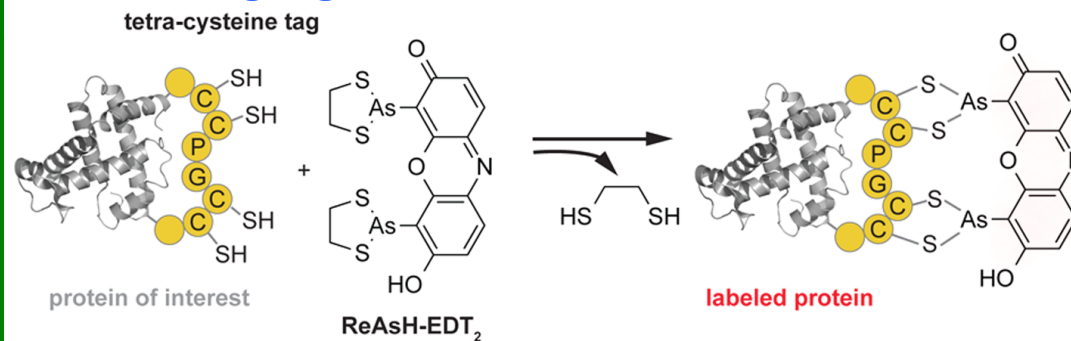
Self Labeling Enzymes



Disadvantages:

- tag is large: 20 - 34 kDa

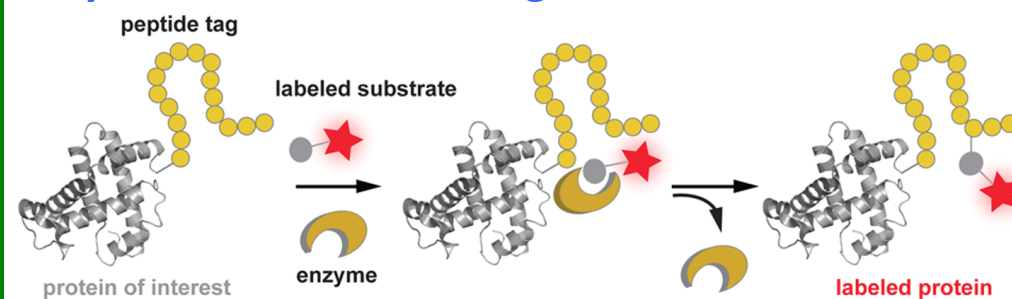
Self Labeling tags



Disadvantages:

- cell toxicity
- non specific binding.

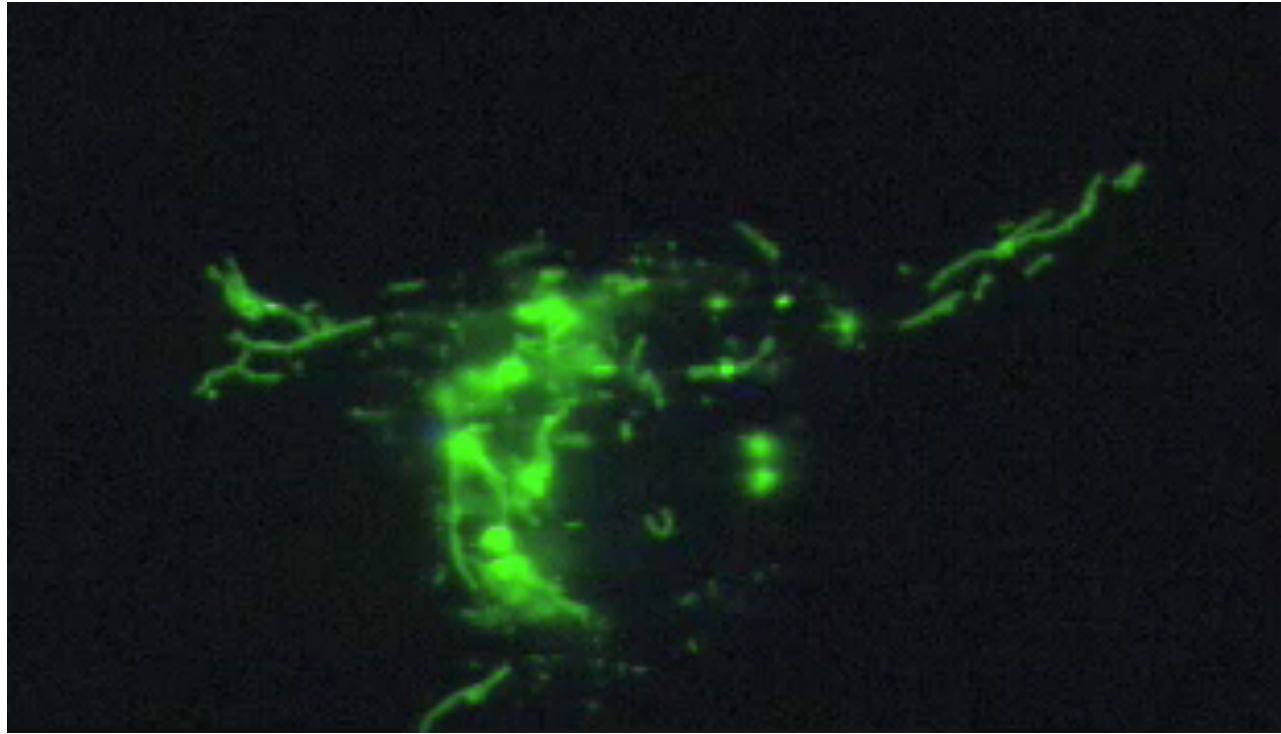
Enzyme Mediated Labeling



Disadvantages:

- cell permeability
- non specific enzymatic activity

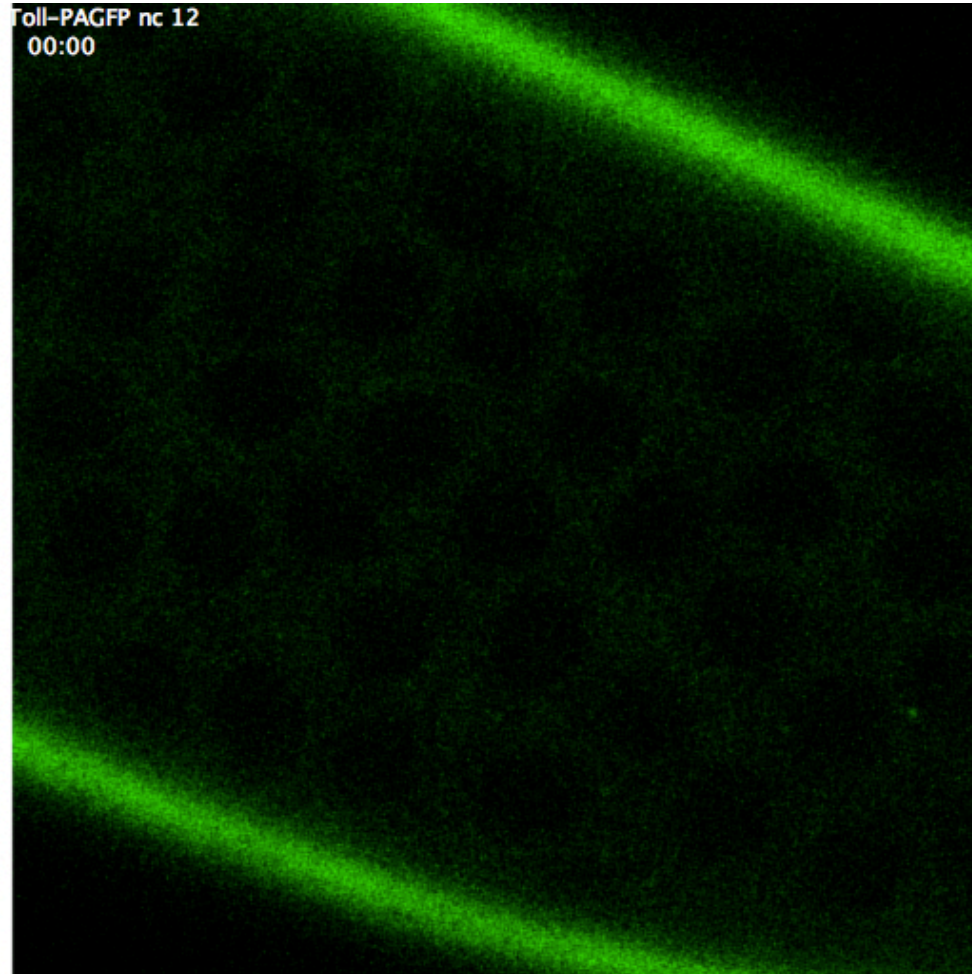
Photoconversion – getting 2 in 1



Using a targeted laser to *repetitively photoconvert mEos from green to red in the mitochondrial network.*

We can assess the degree of connectivity while tracking very dynamic cellular structures.

Photoactivation and photoconversion in *Drosophila* embryos



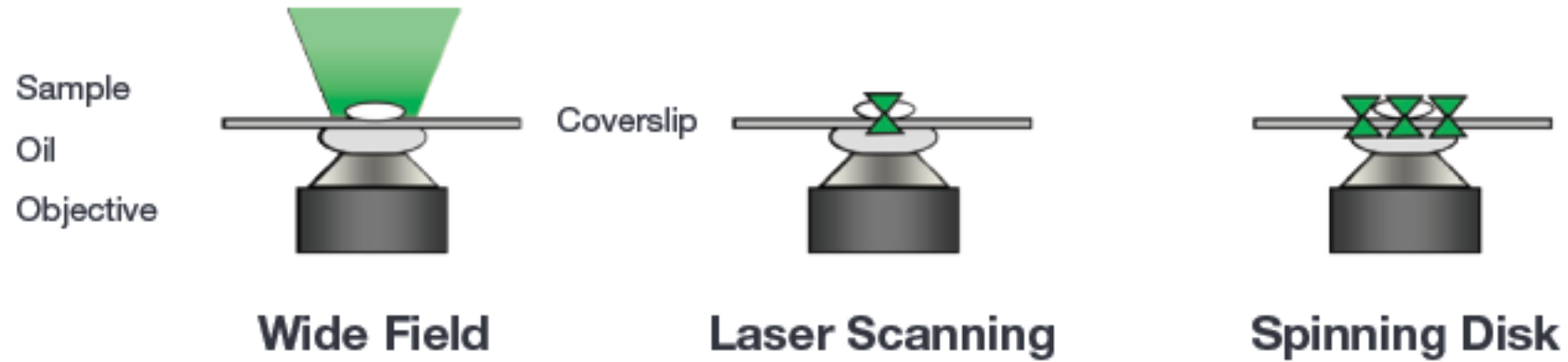
Comparison of available labeling techniques for live cell imaging

	Fl-proteins	techniques using Fl-dyes
Brightness	+	+++
stability	+	+++
Applicability to SR techniques	+	+++
Size of tag	+	+
Flexibility in choosing the tagging site	+	+
SNR	++	++
Over expression artifacts	++	++

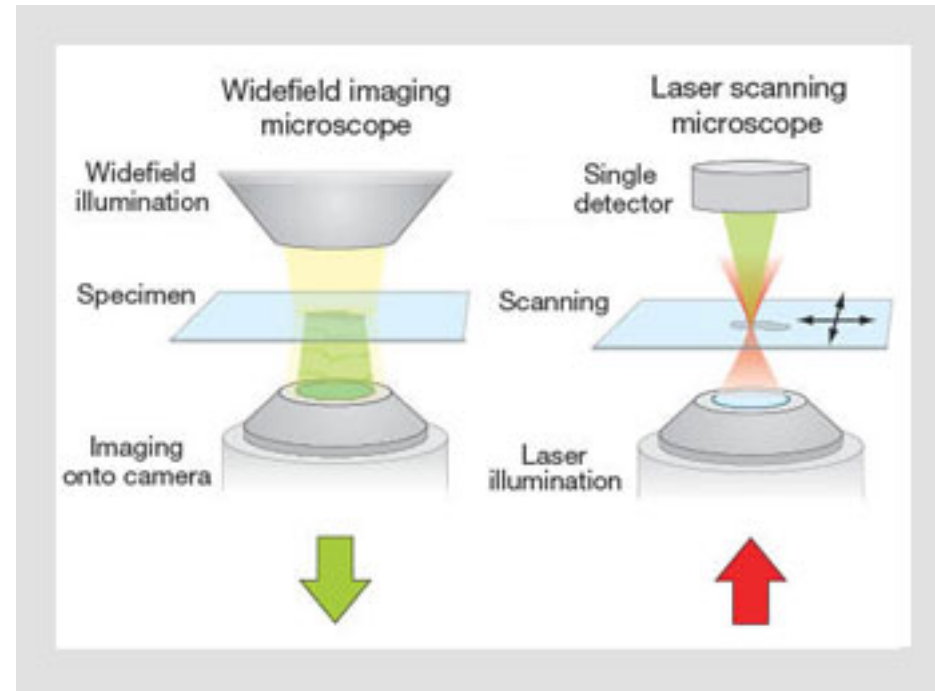
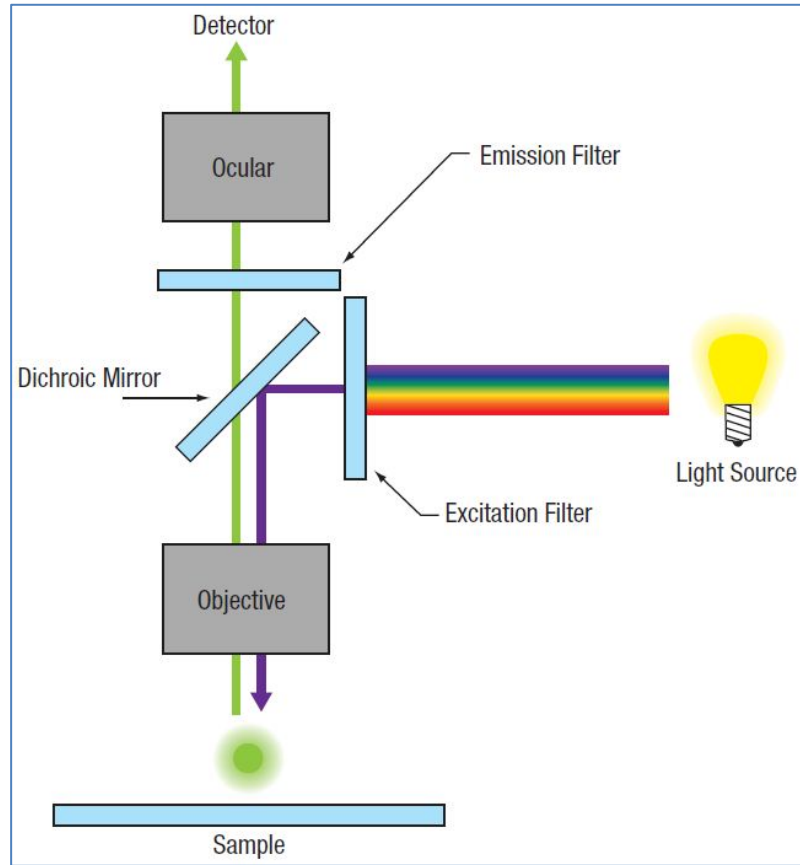
Live cell imaging - considerations

- Cells are sensitive to laser illumination in the blue-green range – laser exposure should be minimized. This means that imaging parameters should be compromised in order to maintain cells in physiological conditions.
- Temporal resolution should be determined based on the process studied.
- Temp. and CO₂ levels - incubation system. Good for cells but introduce heat to the optical system.

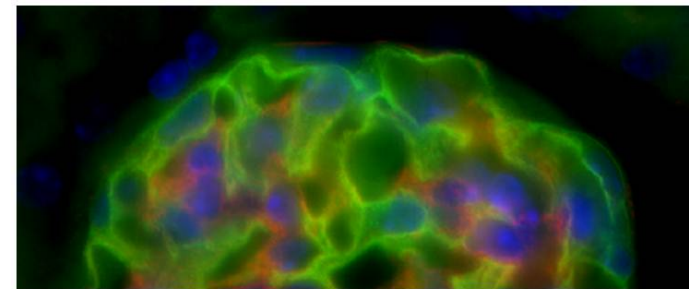
Method comparison



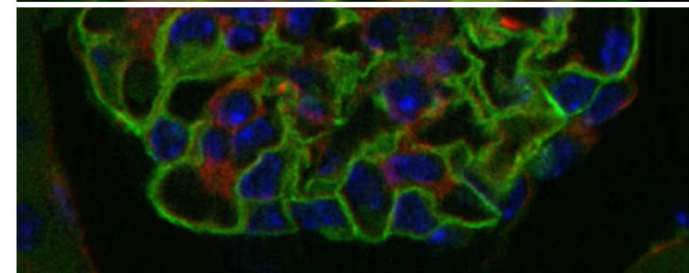
Widefield Microscope



Widefield

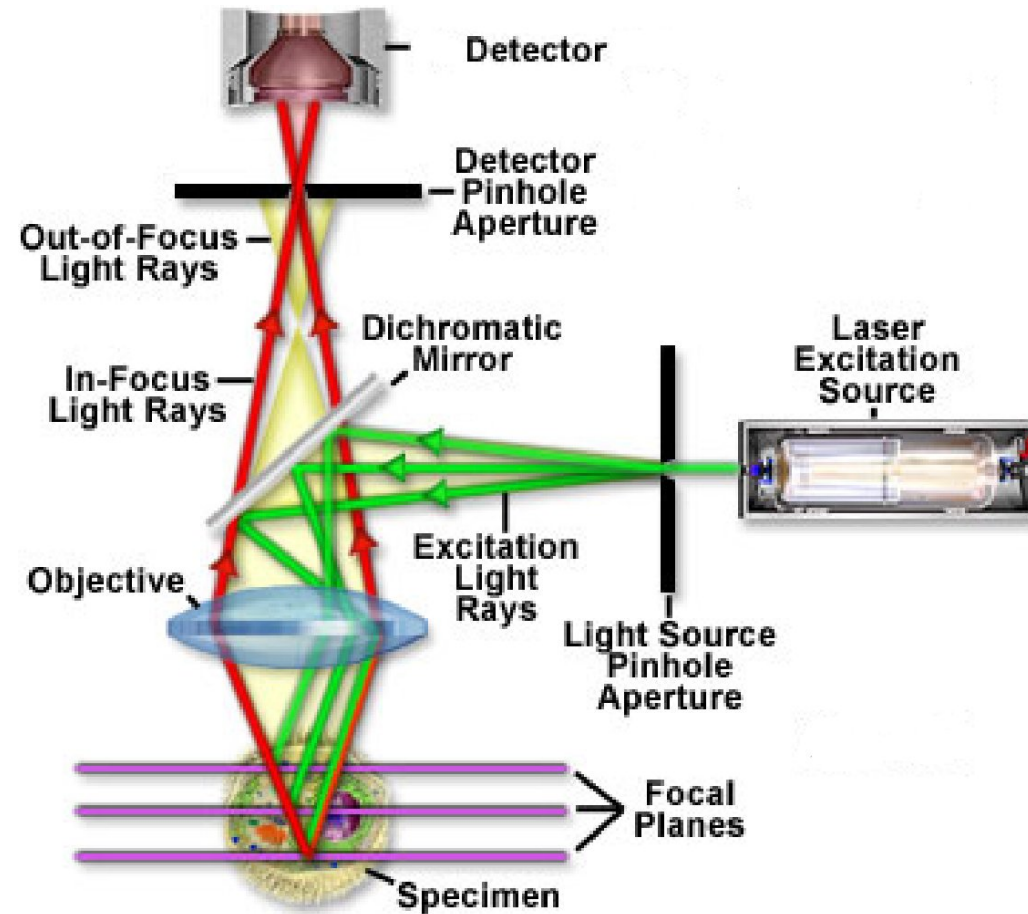


Confocal

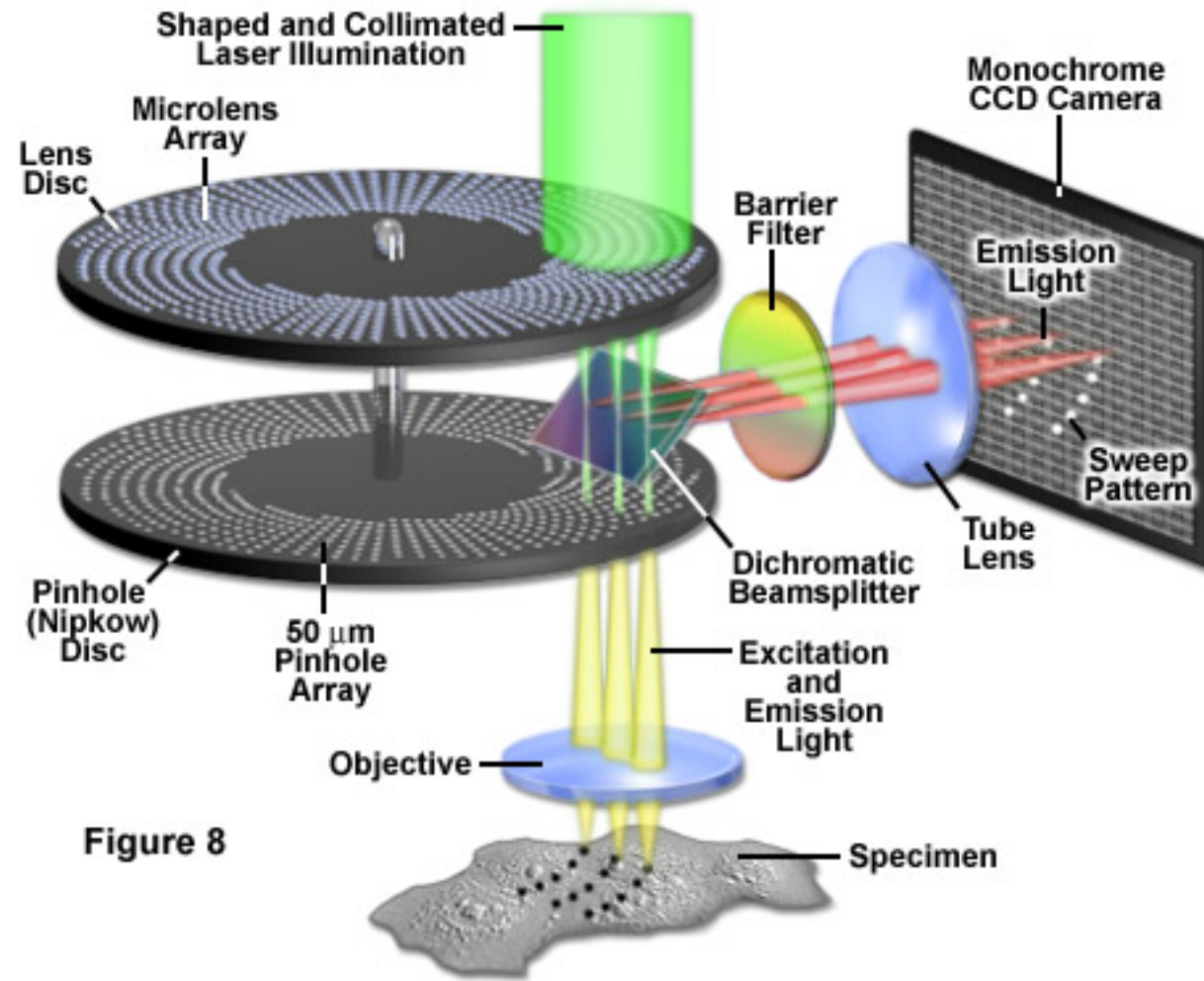


Delivering light energy to a region of interest in a cell

Targeted laser illumination typically uses the same technology as scanning laser confocal microscopy.

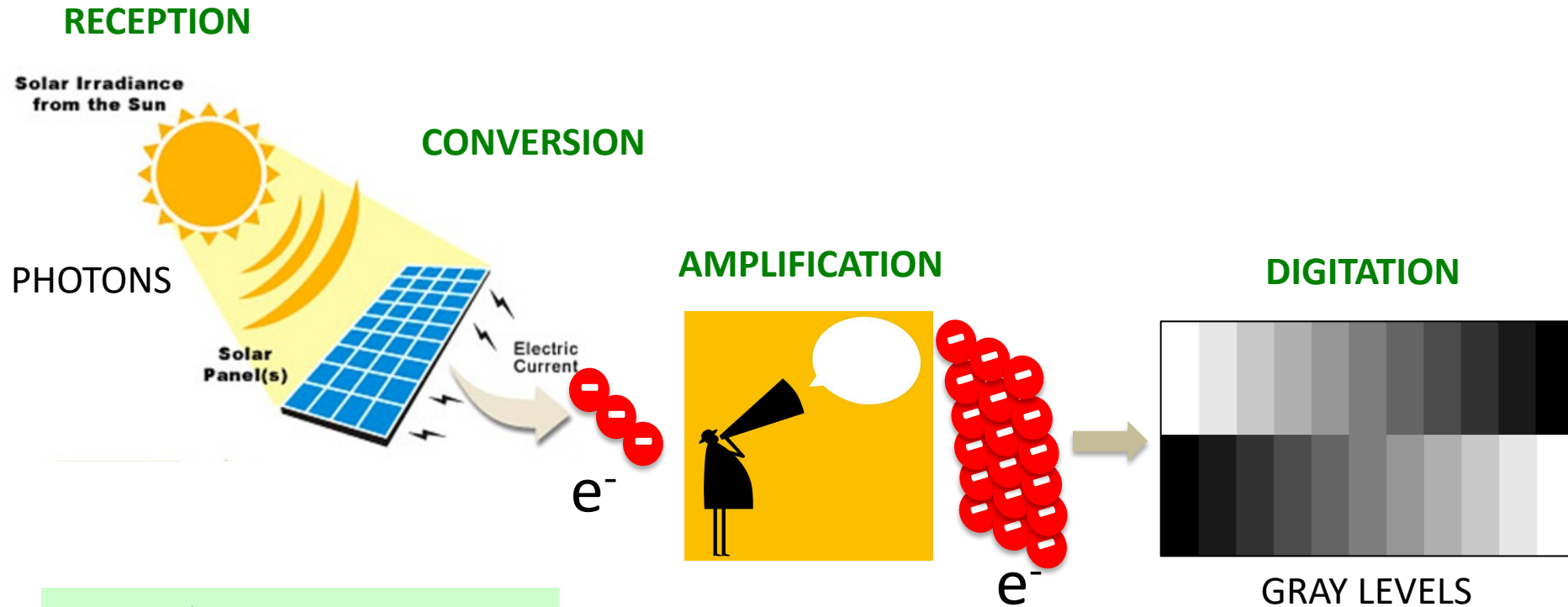


Spinning disk



Detectors

Detect photons -> convert to electrons -> amplify signal -> Convert to digital signal



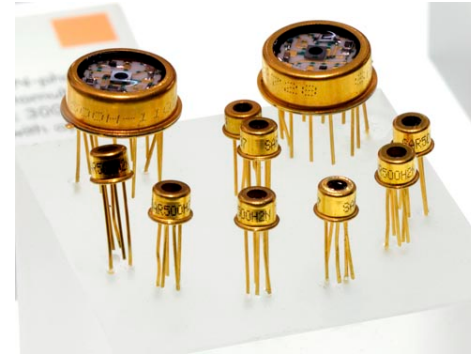
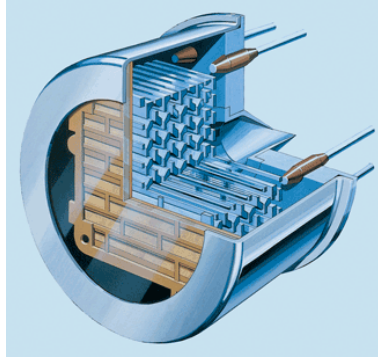
Demands:

- Speed
- Wide range of wavelength
- Good dynamic range
- High resolution
- Low noise (high SNR)

Types of detectors

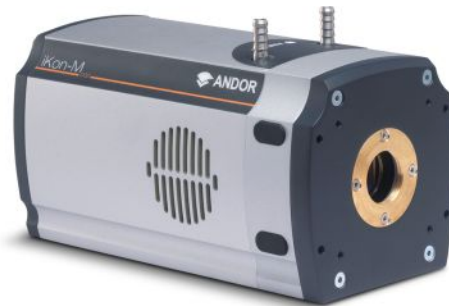
Spot Detectors:

- PMT
- APD

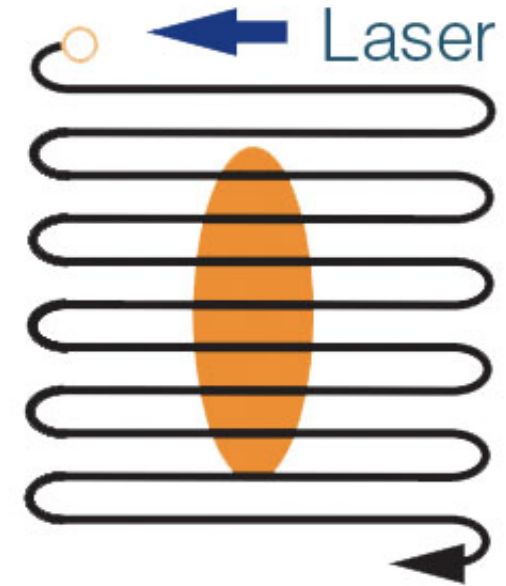
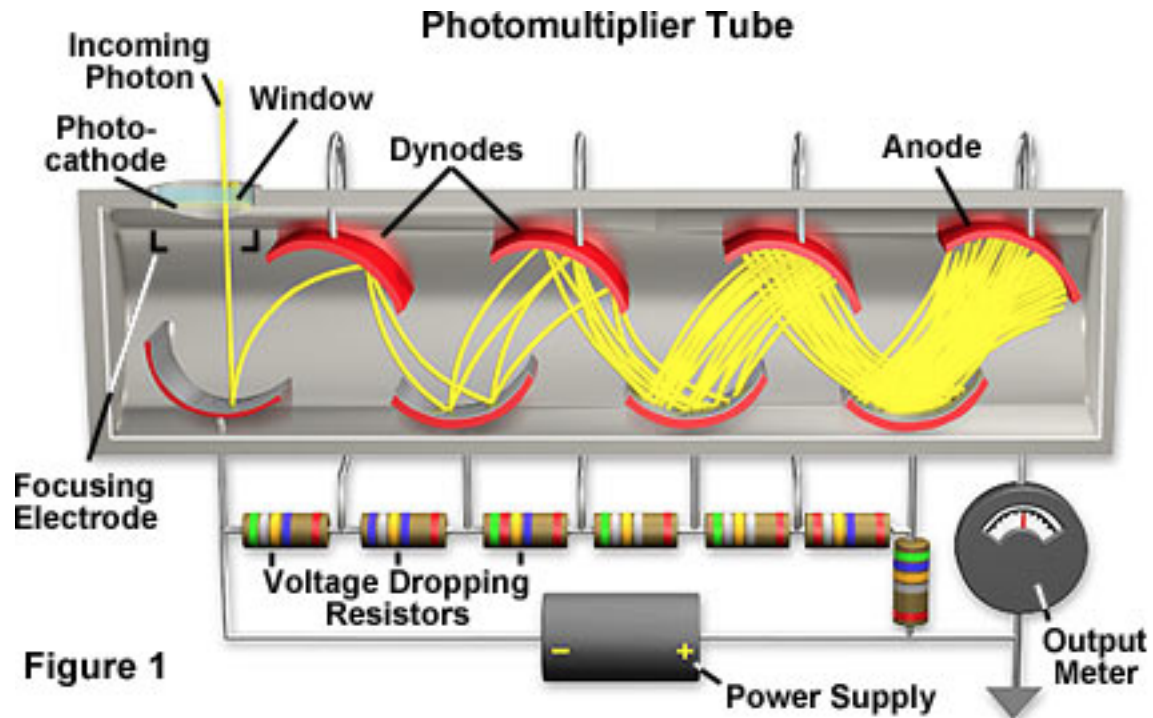


Area Detectors:

- CCD
- EMCCD
- sCMOS



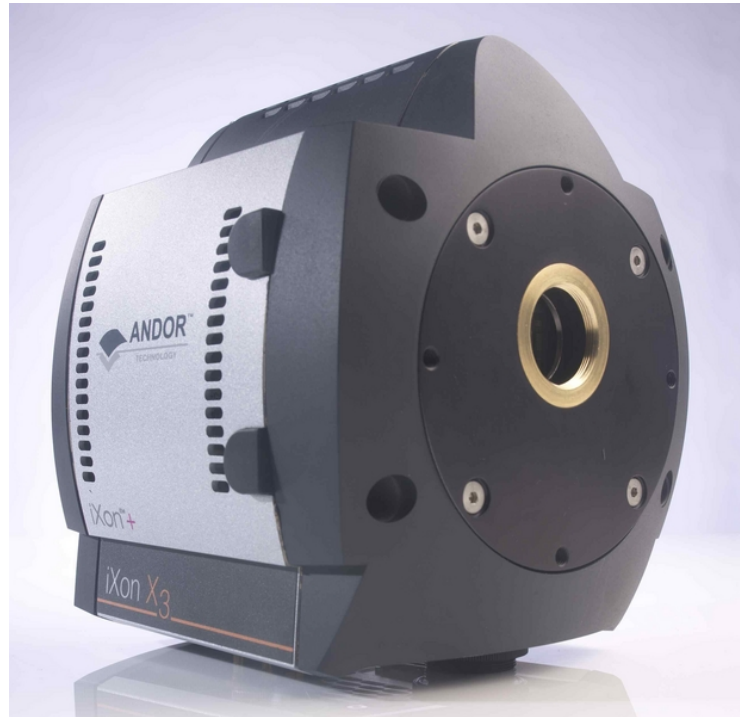
Detection: Photomultiplier (PMT)



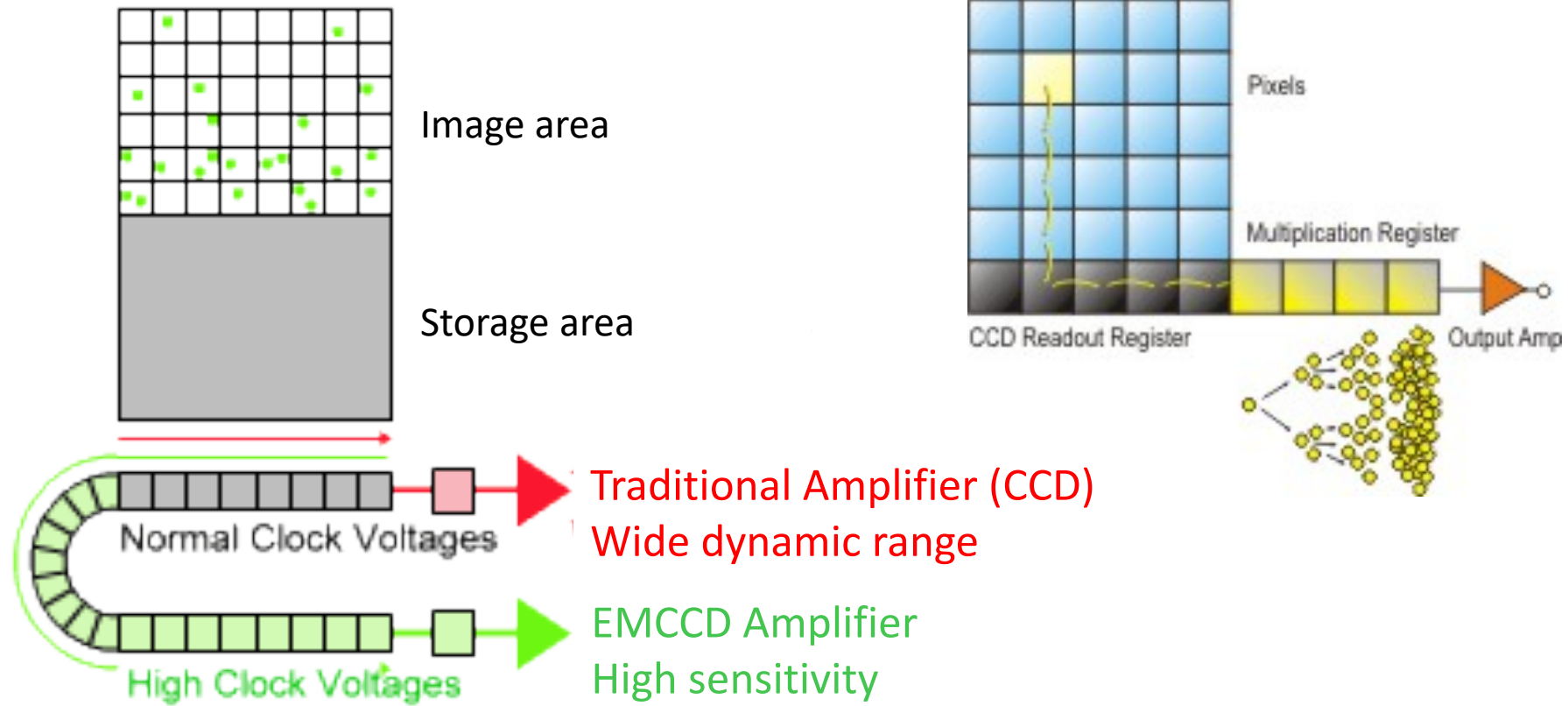
Laser scanning
confocal

The Detector – EMCCD camera

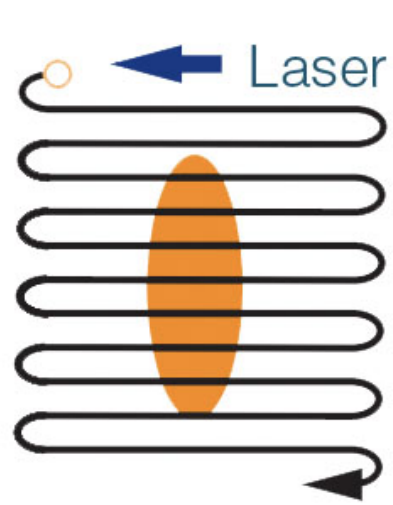
- * The whole field of view is imaged at the same time
- Camera is monochromatic – black and white – all colored images are artificial
- Pixel size is different in different cameras.



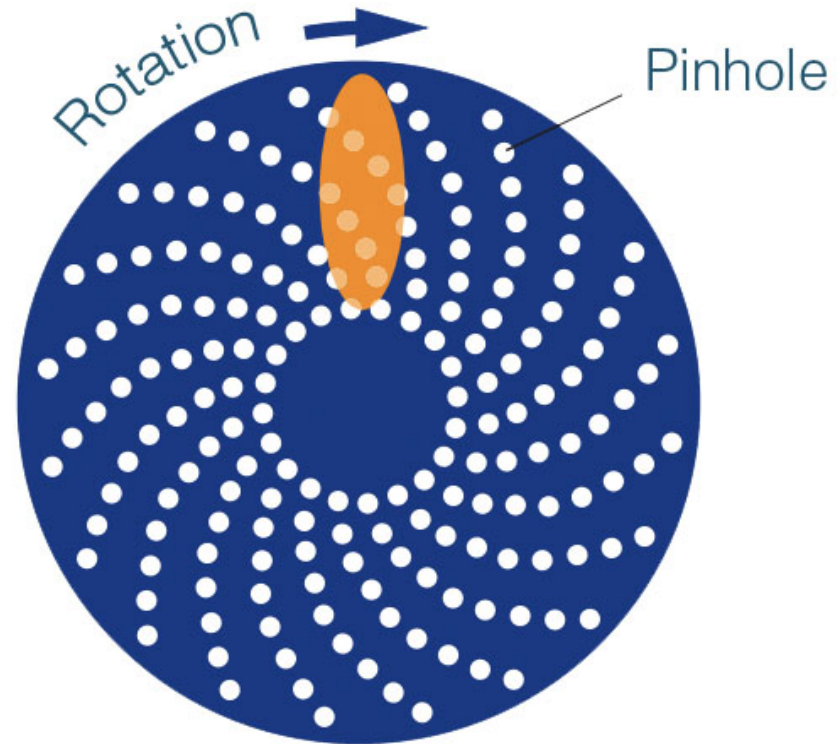
How does it work?



Detection confocal / Spinning Disk



Laser scanning
confocal



Nipkow disk
confocal

Spinning Disk: Faster acquisition + faster detection –
Ideal for live cell imaging

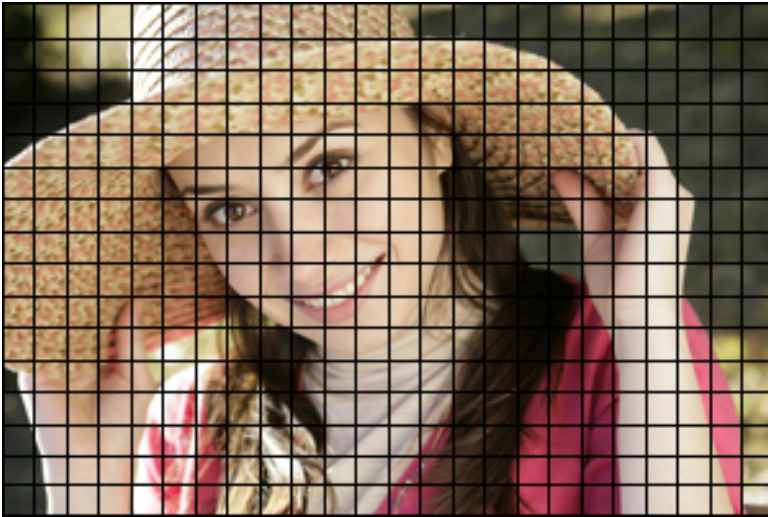
Area detectors - cameras

Pixel - the smallest component, sensor unit

Large Pixel:

Lower resolution

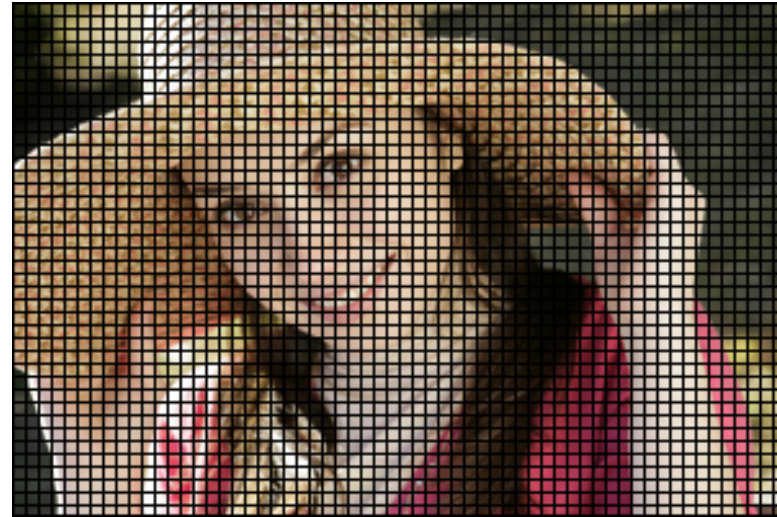
Higher sensitivity



Small Pixel:

Higher resolution

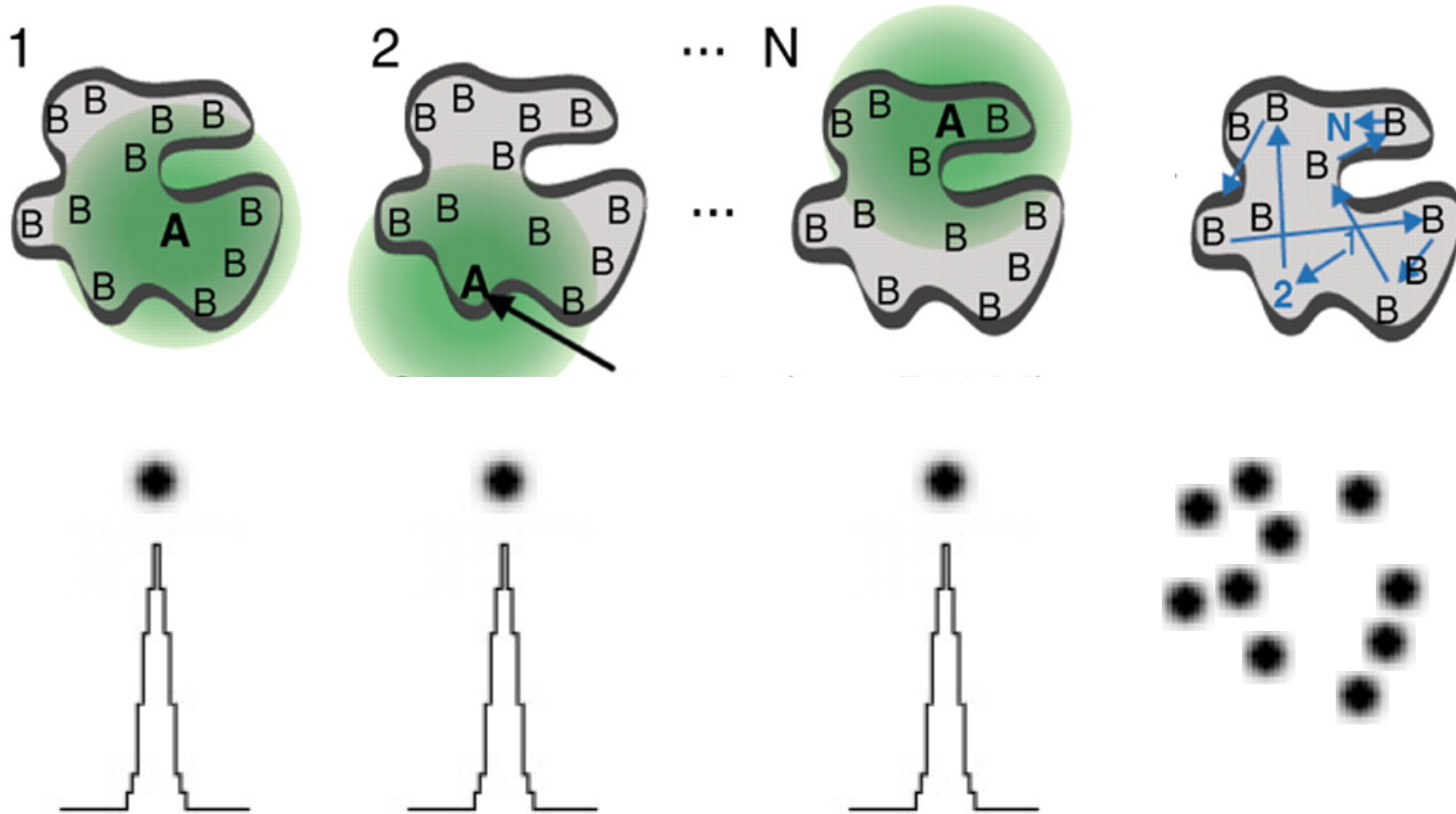
Lower sensitivity



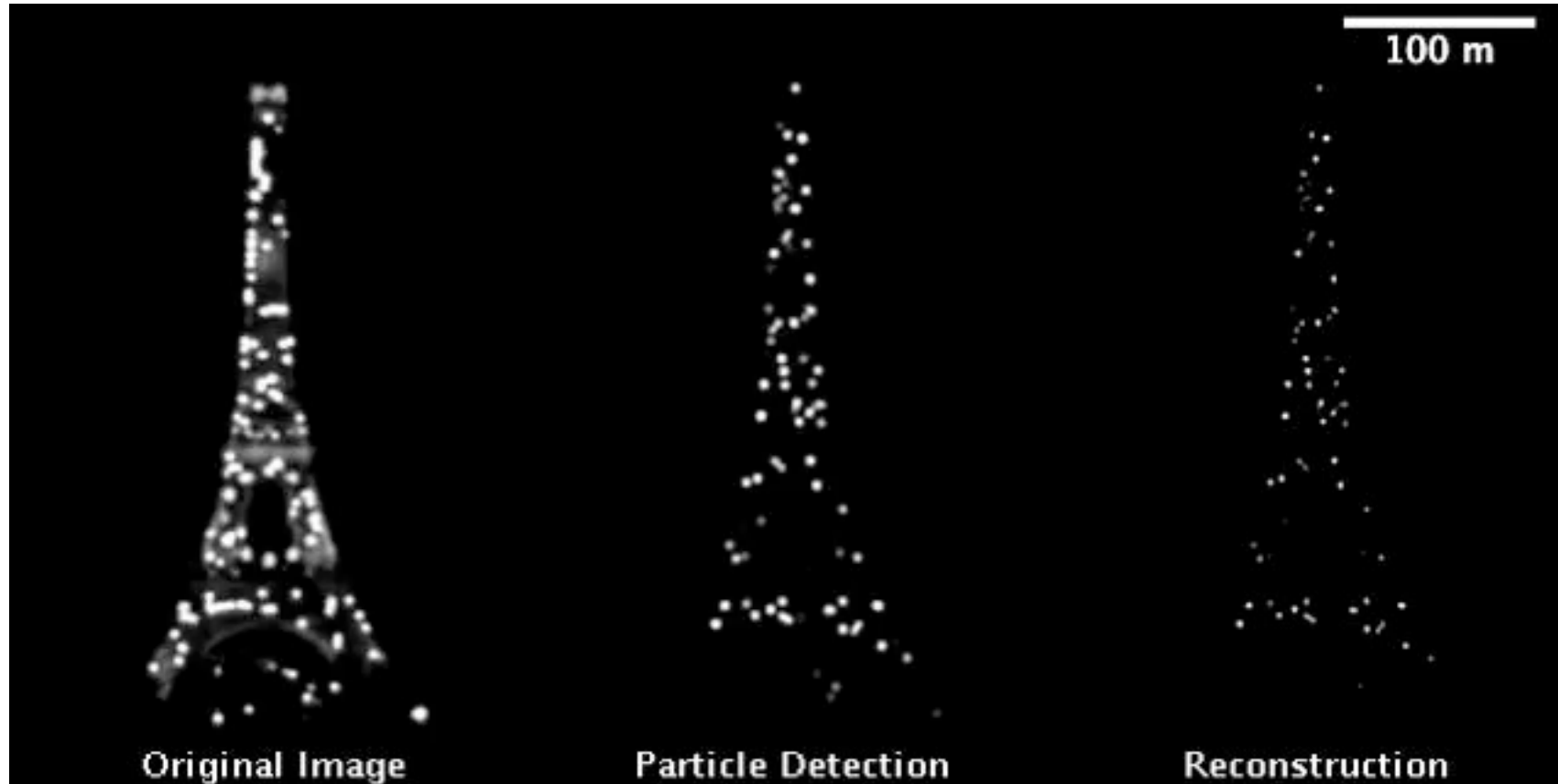
Detector comparison

	PMT	APD	CCD	EM-CCD	sCMOS
Quantum Efficiency	40%	80%	95%	95%	70%
Speed	Slow (pixel by pixel)	Slow (pixel by pixel)	Fast area scan (56 fps)	Fast area scan (26 fps)	Fastest (100fps)
Pixel size			Small (μm)	Normally Larger than CCD	Smallest
noise	low			Added noise factors. Low readout noise <1e	Low 1.6e
Sensitivity	High	High	Lower than EMCCD	High	High but small pixels
Saturation		Easy to saturate	Depends on pixel size	Depends on pixel size	Smaller pixel size lower FWC
Detector lifetime		Limited		Limited - detector aging	

Super resolution microscopy - Localization Microscopy



Super resolution microscopy - Localization Microscopy



Visualizing ESCRTs in STORM (30 nm resolution)

Tubulin, IST1

