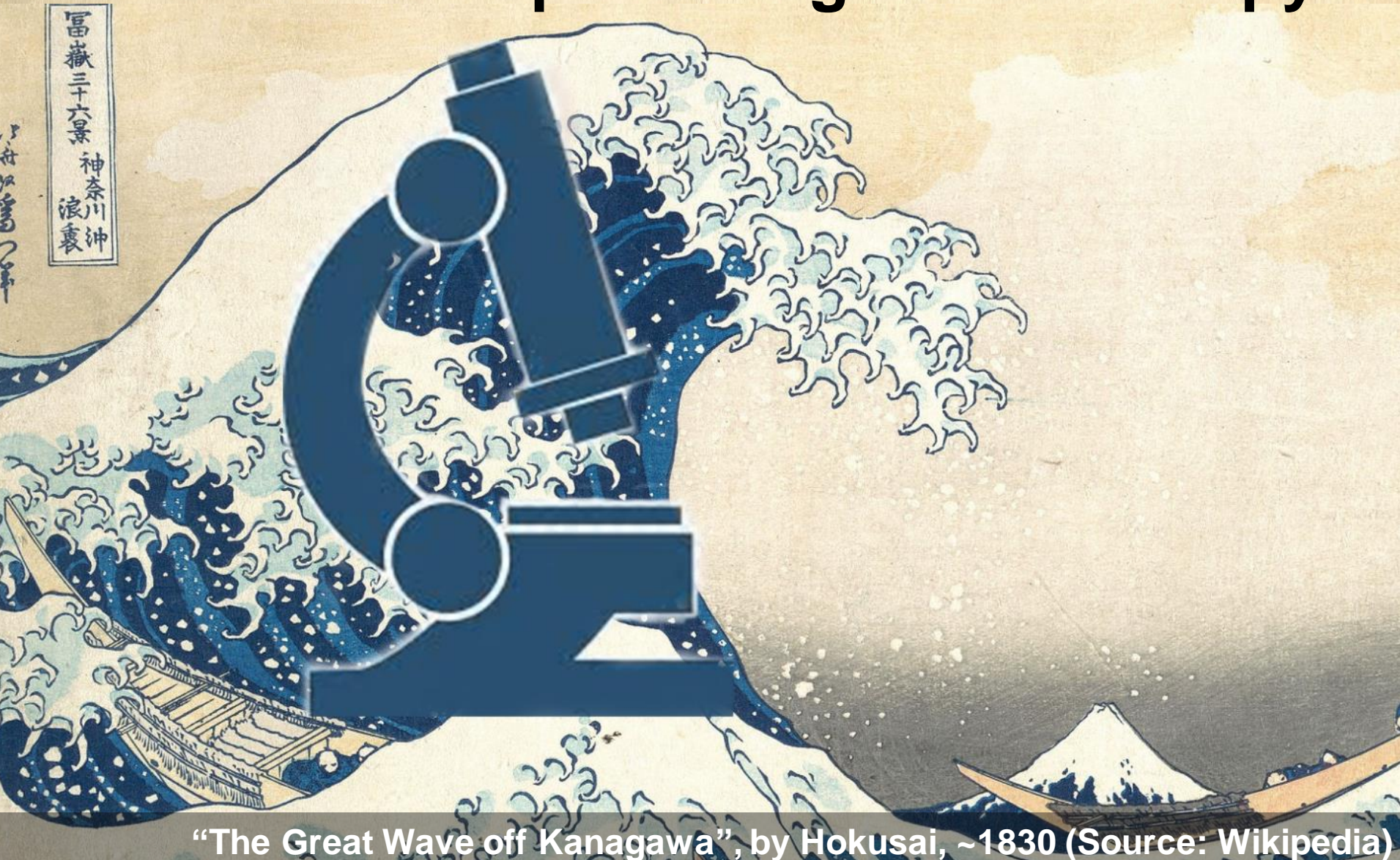


# Data science in cell imaging

## Lecture 5: deep learning in microscopy





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PPTX slides available [here](#)





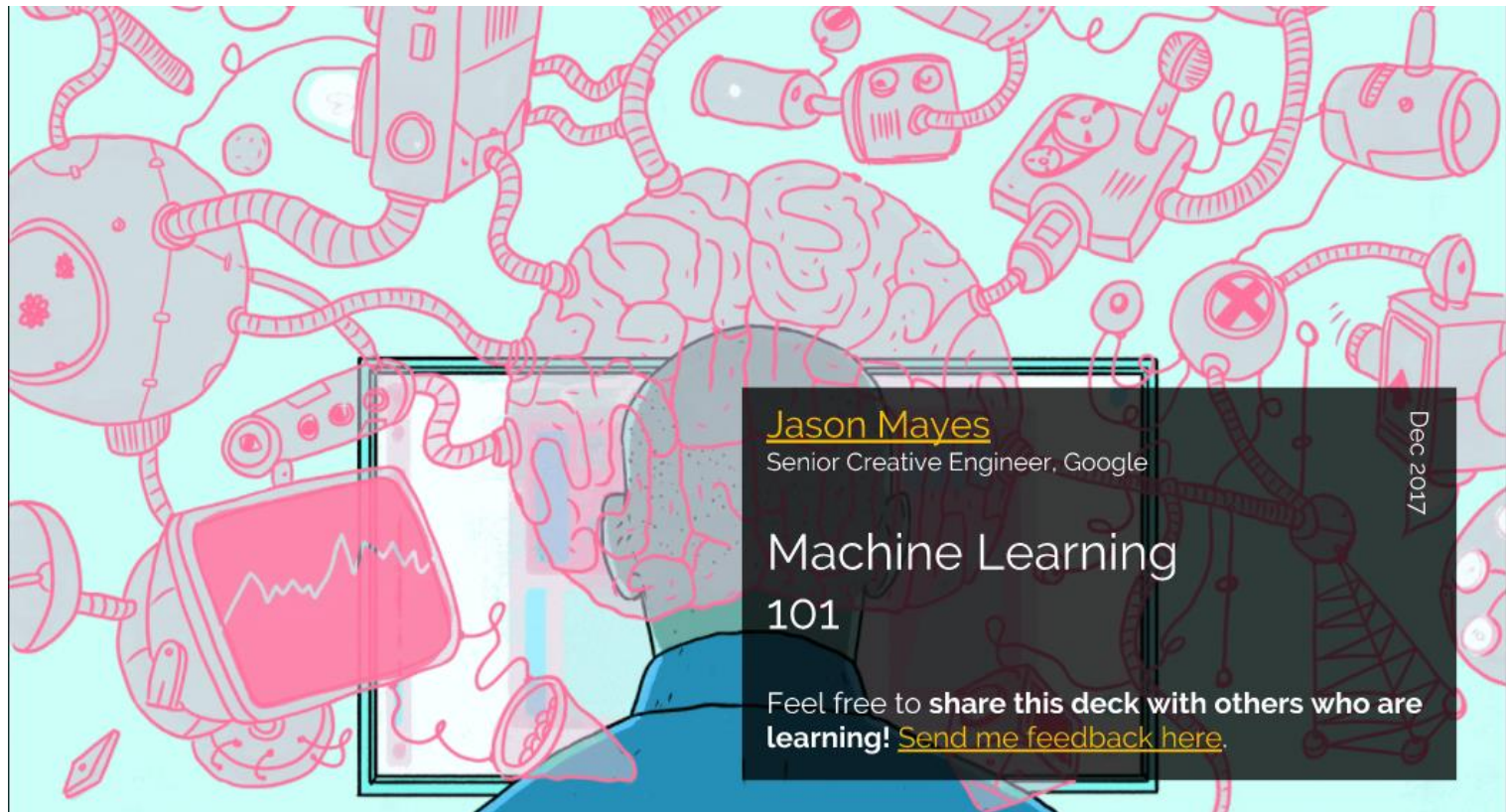
# Today and next week

- (Discussing potential course projects)
- Enhancing cell image quality with deep learning
- Generative models for cell structure with deep learning
- Classifying cell state with deep learning



# Machine learning 101

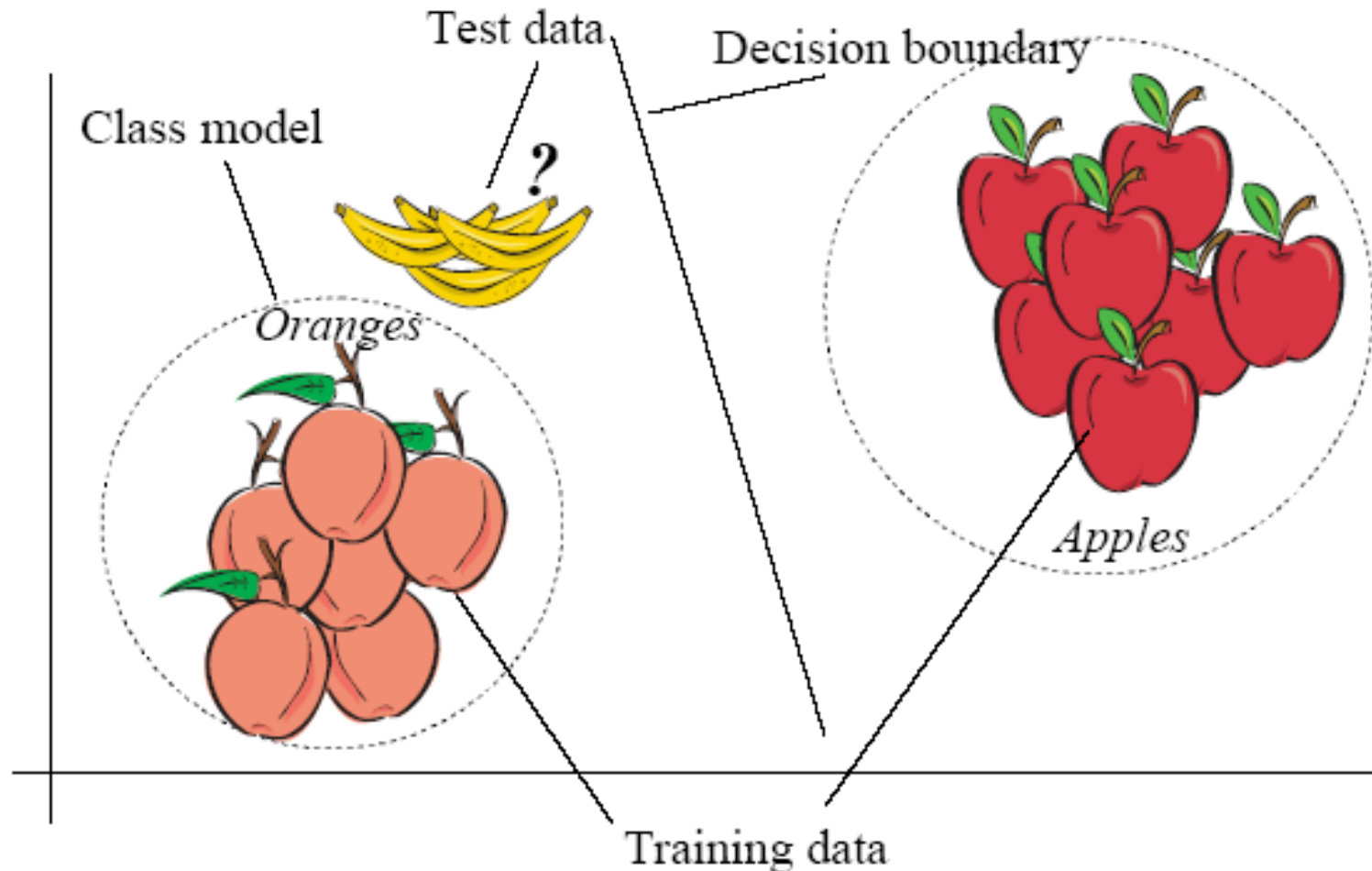
Recommended deck for non-computational students





# Supervised machine learning

Data → feature extraction → training →  
discriminative model



Images by MIT OCW.

Credit: B. Heisele, Y. Ivanov, T. Poggio



# Unsupervised machine learning

Data → feature extraction → clustering

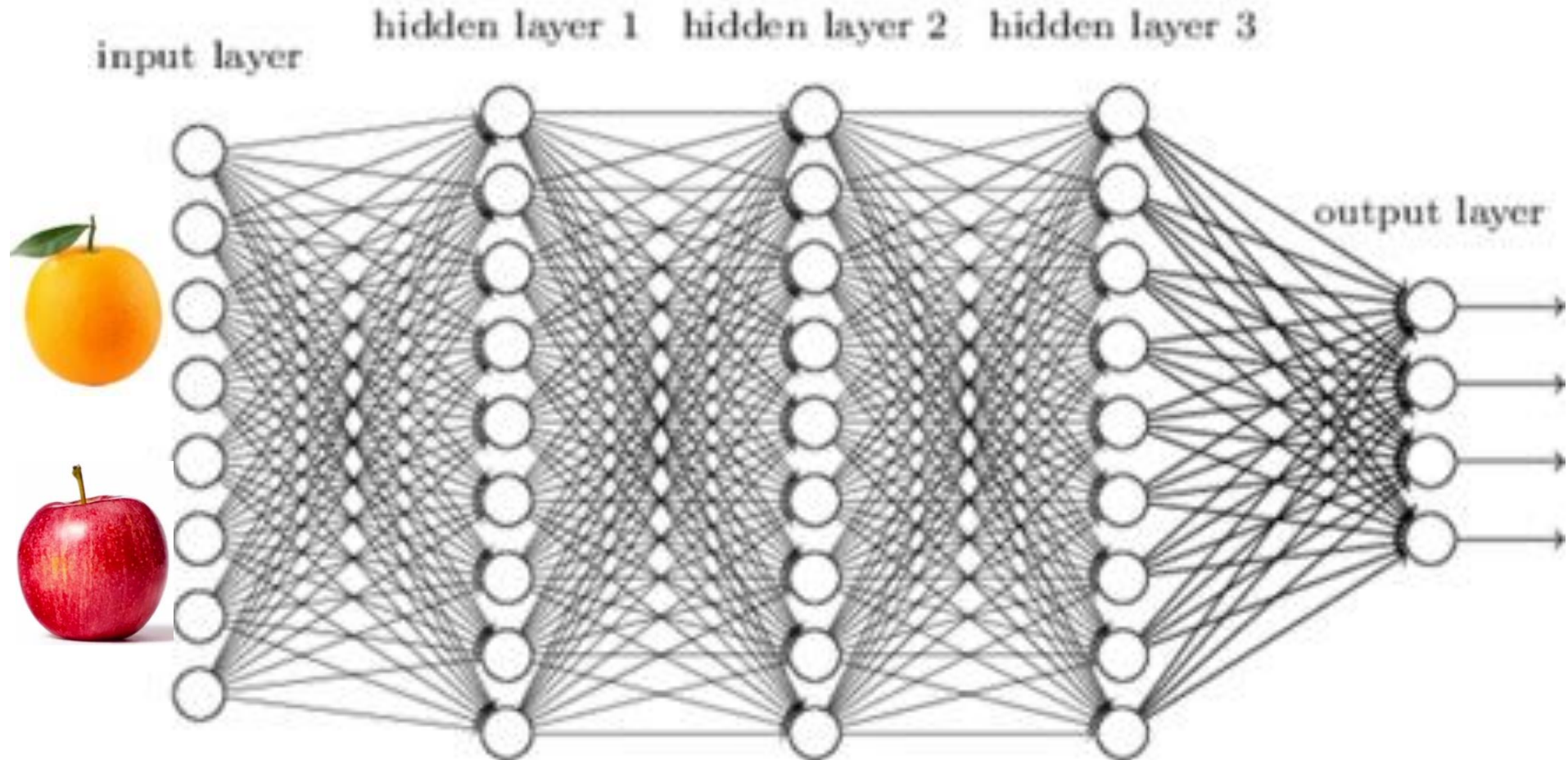


Source: <https://bit.ly/2IJN0NT>



# Deep learning

A powerful integration of automated feature extraction and model training



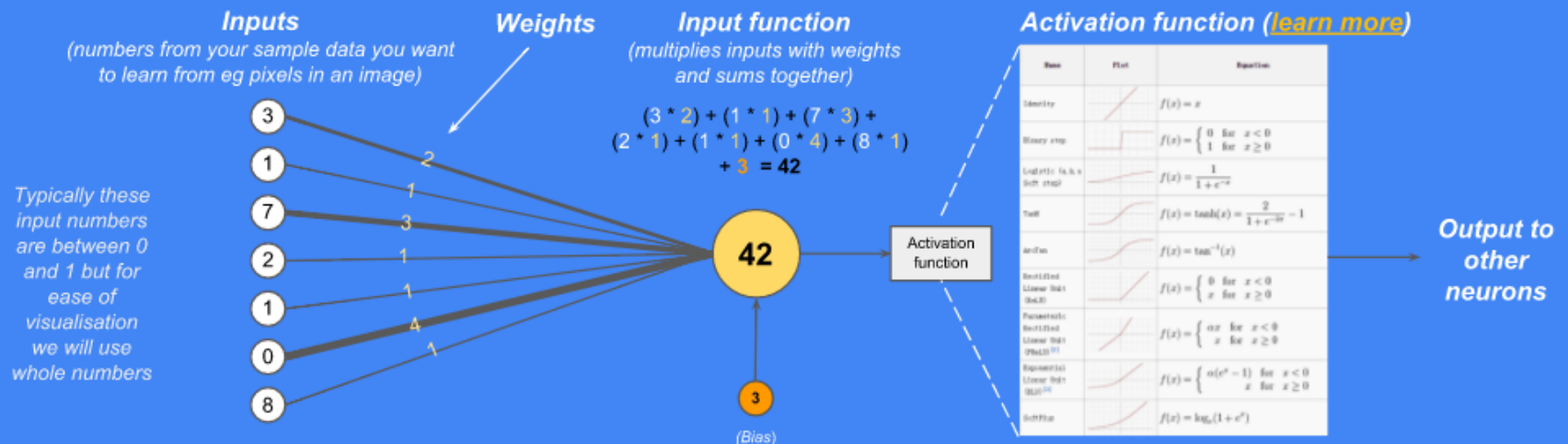


How does it works?



# Artificial neuron (perceptron)

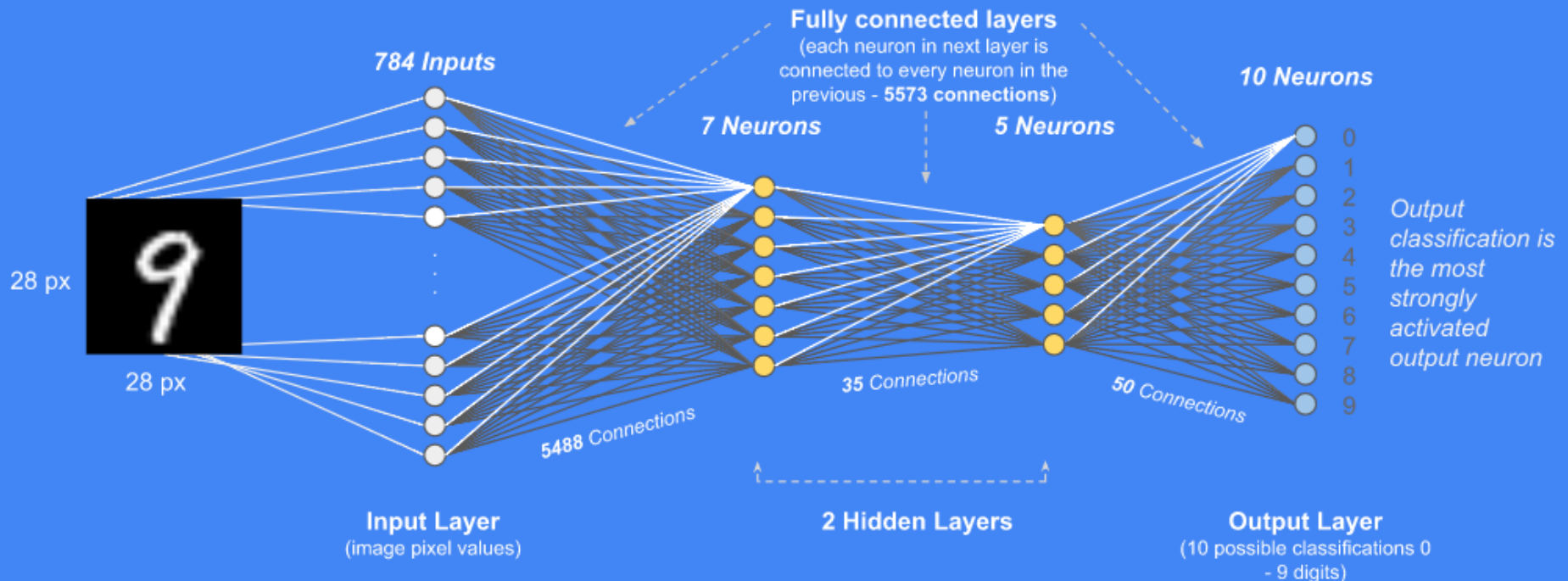
A neuron simply has a **bunch of weighted inputs** (take the input number and multiply by the weight) **that are summed together**. A bias is then added to this total. The weights and bias are determined when we train the system. If final result is greater than a threshold, it activates, providing an output. Strength of output depends on the activation function chosen. The output is then fed into other neurons and the process repeats.





# Deep neural network

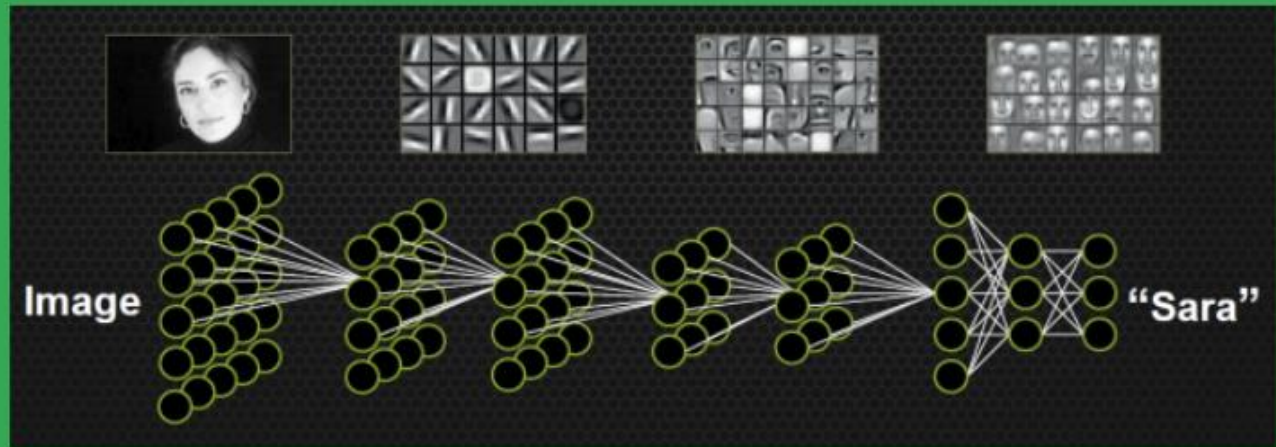
This is a **multi layered perceptron** (or deep neural network) - one of the oldest forms of “neural nets” - conceptually goes back to the 60s! Each layer is **fully connected** to the next and **data flows forwards only**:





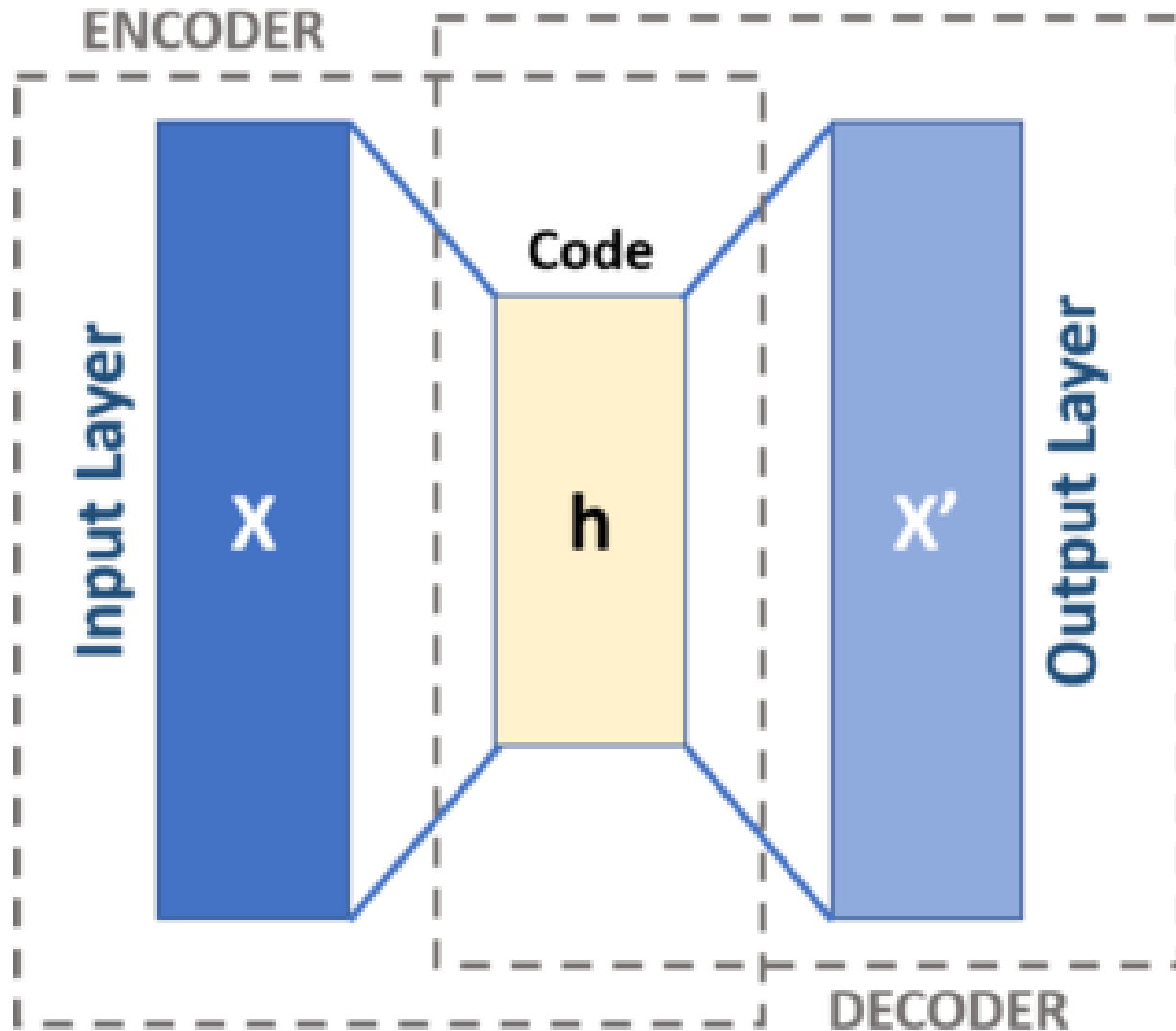
# Deep neural network

A Deep Neural Network (DNN) simply consists of many “hidden layers” between the input and the output. Each layer can learn from the one before it from which higher level learning can take place. These hidden layers typically are of lower dimensionality so they can generalise better and not overfit to the input data. These middle layers in the system can learn features of features. For example bunches of “edges” can lead to “face parts” which lead to “faces” that the system can then recognize.





# Autoencoder: unsupervised data encoding





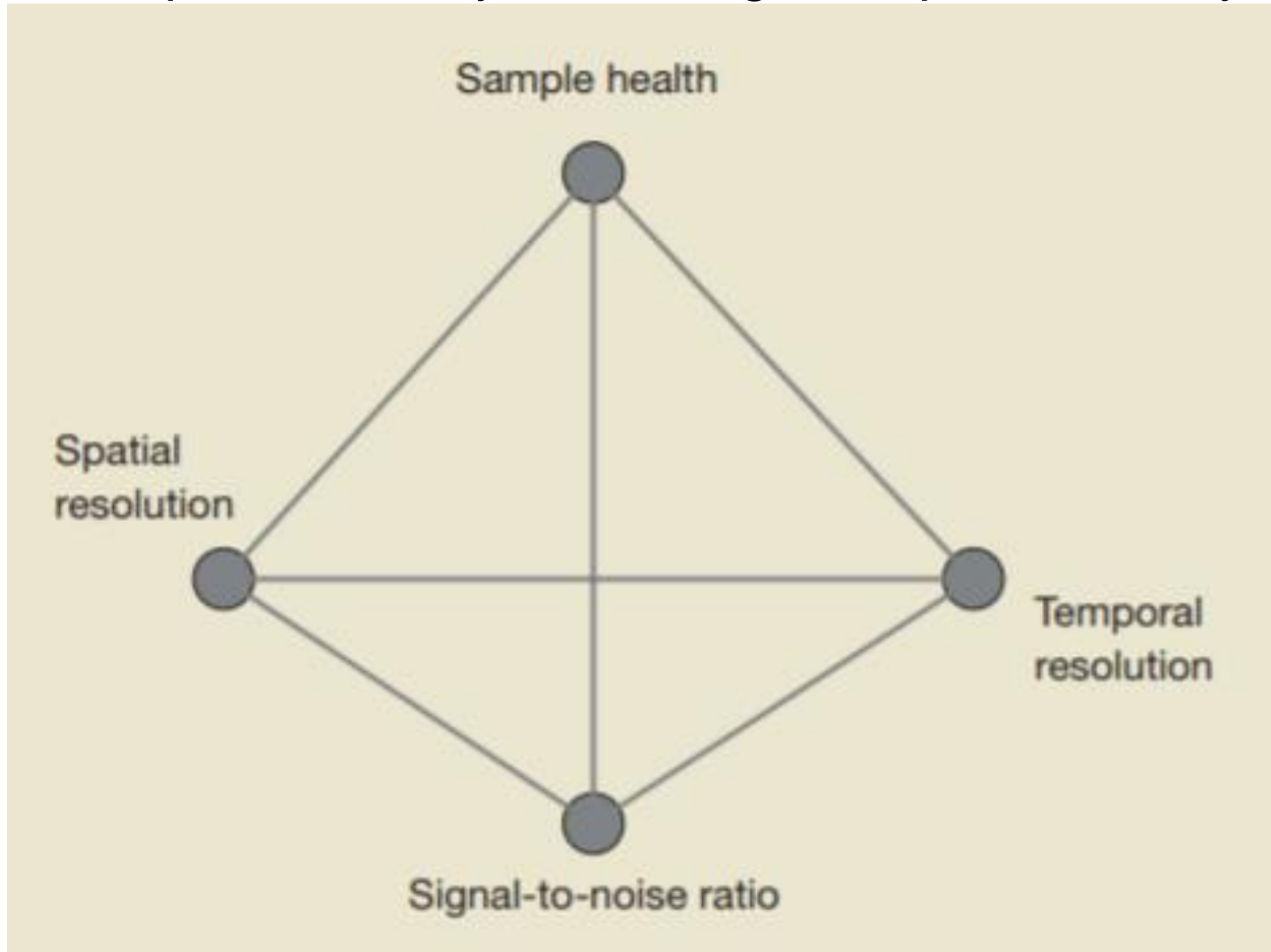
# Many applications for ML in cell biology!

Today I'll review a few specific examples on deep learning for (1) image restoration; (2) prediction of (un-labeled) protein localization patterns



# Imaging tradeoffs

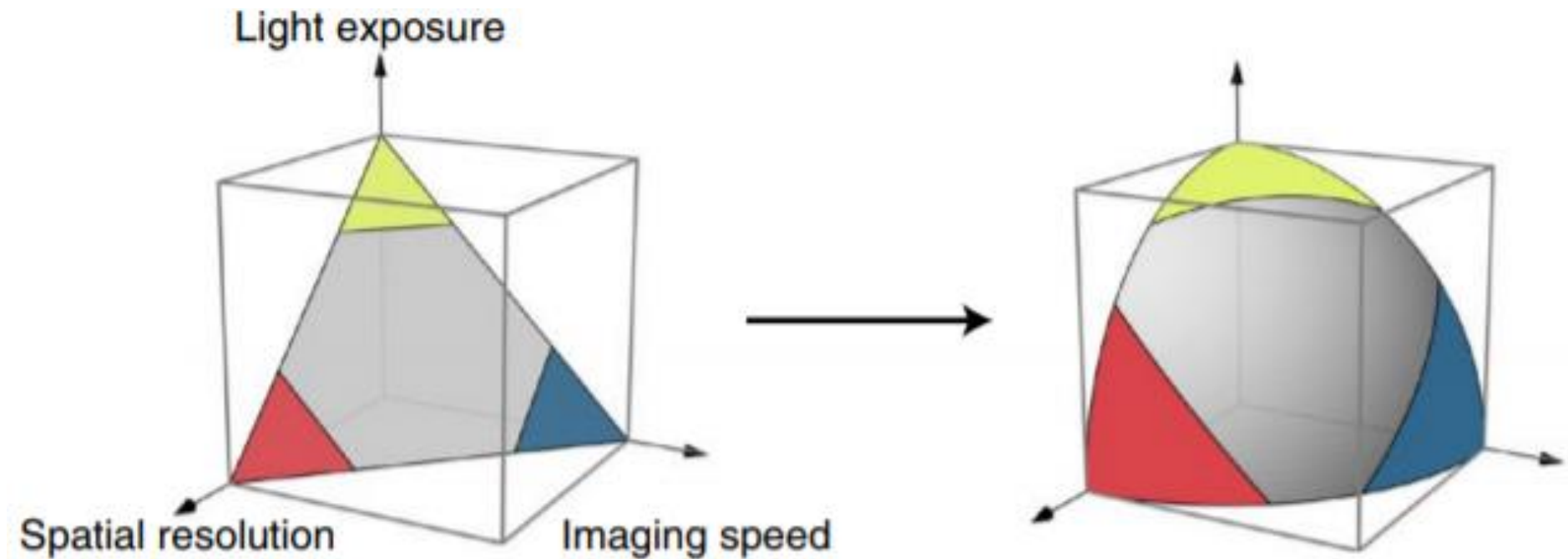
Spatial/temporal resolution, experiment time, imaging depth, fluorophore density, bleaching, and photo-toxicity



Hadar Aharoni (IDF) from Scherf et al. (2015)



# Content aware image restoration (CARE)

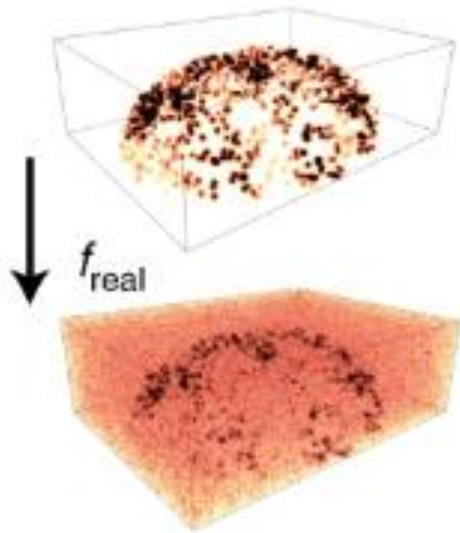


- Traditional methods use general assumptions to perform the restoration
- CARE leverages the available knowledge about the specific experimental task / setup

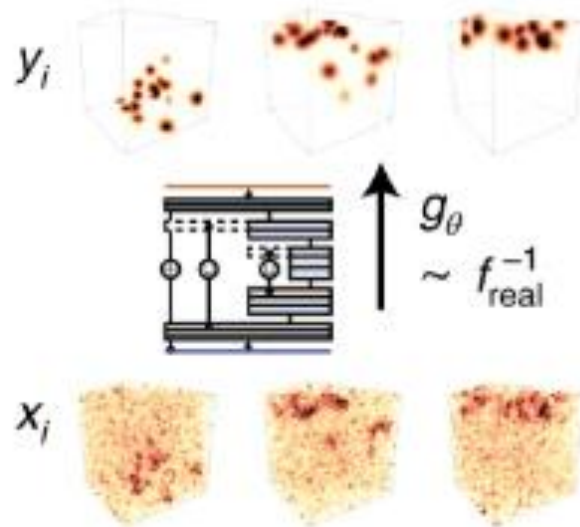


# Content aware image restoration (CARE)

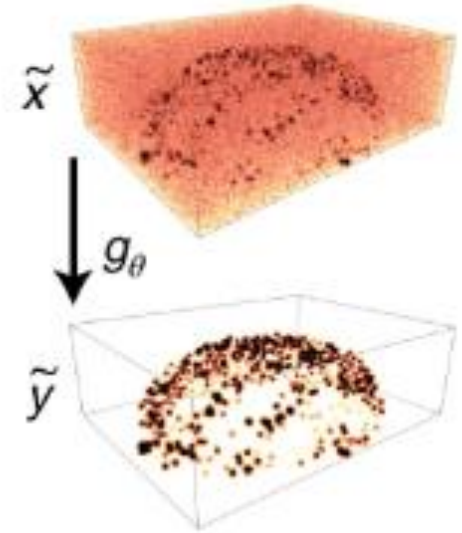
Training data generation



Training



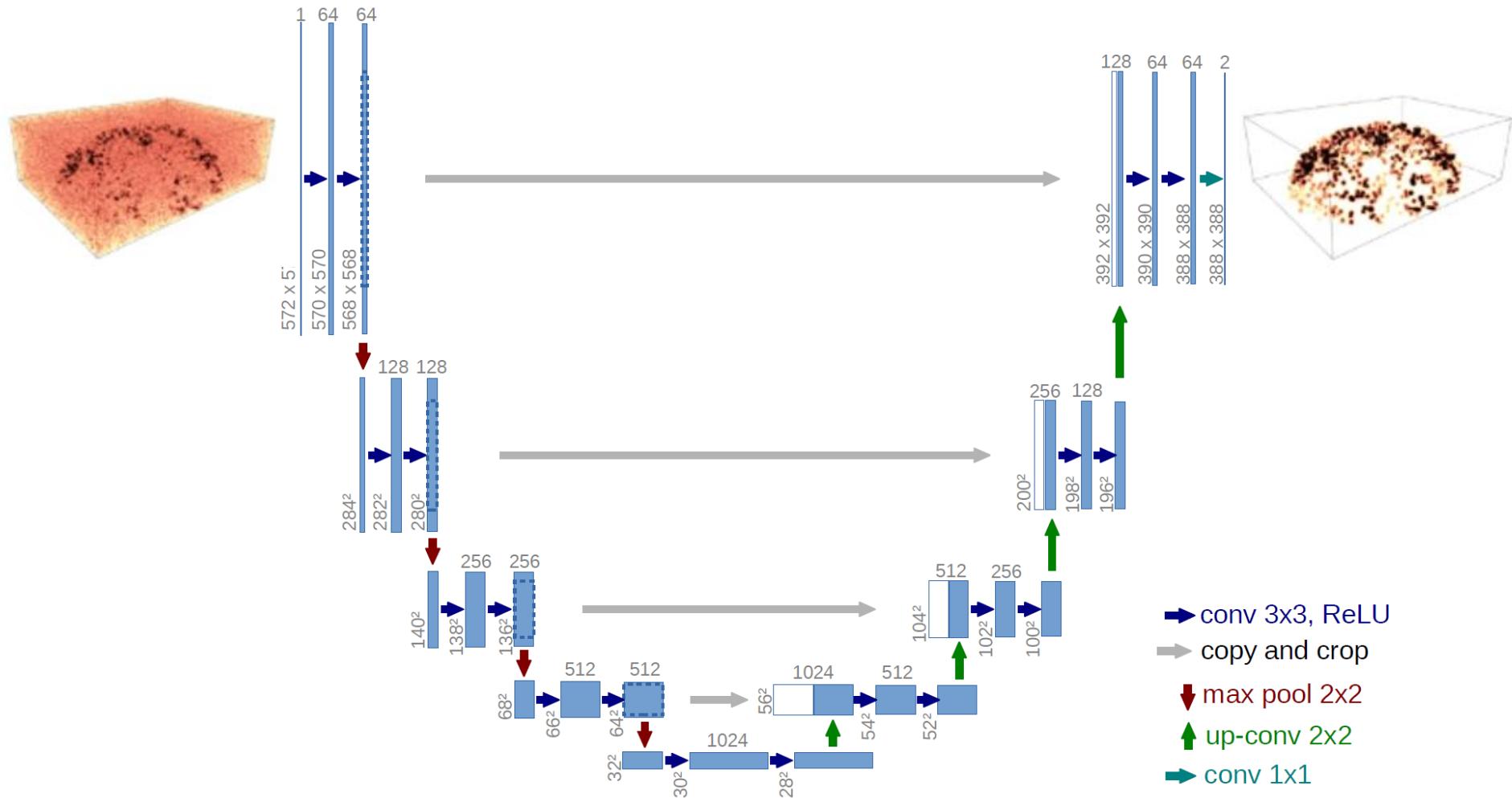
Application



- Traditional methods use general assumptions to perform the restoration
- CARE leverages the available knowledge about the specific experimental task / setup

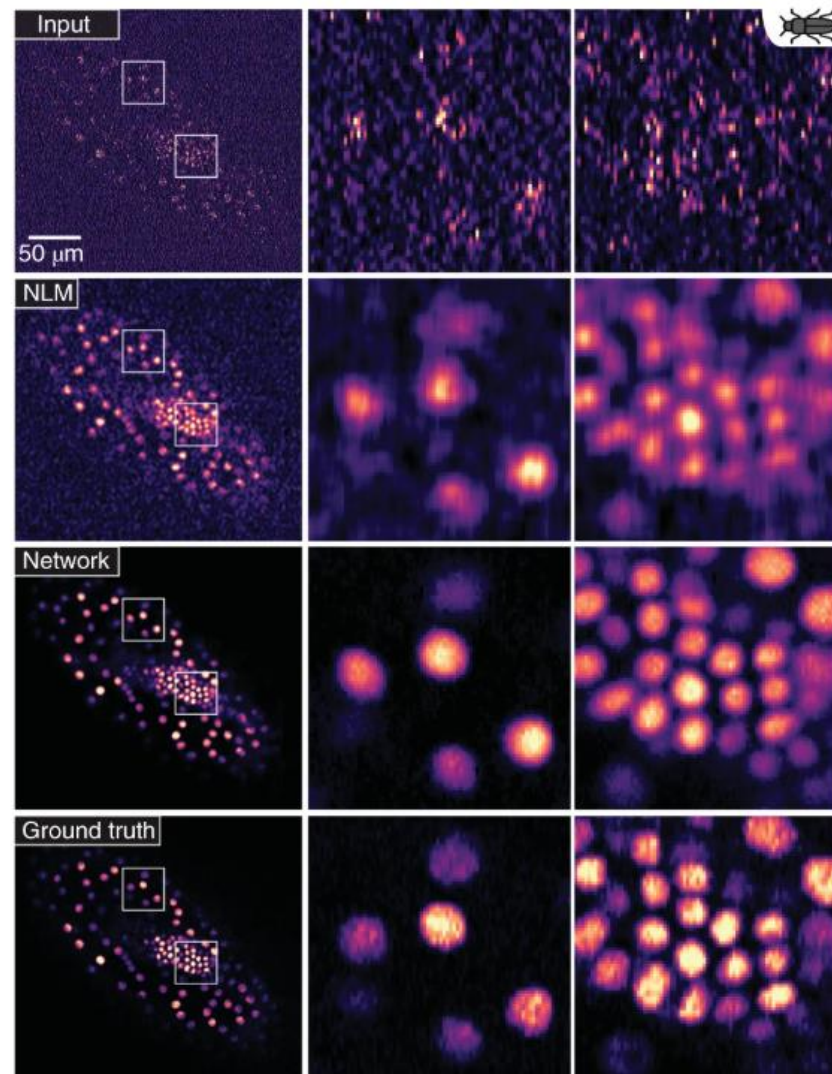
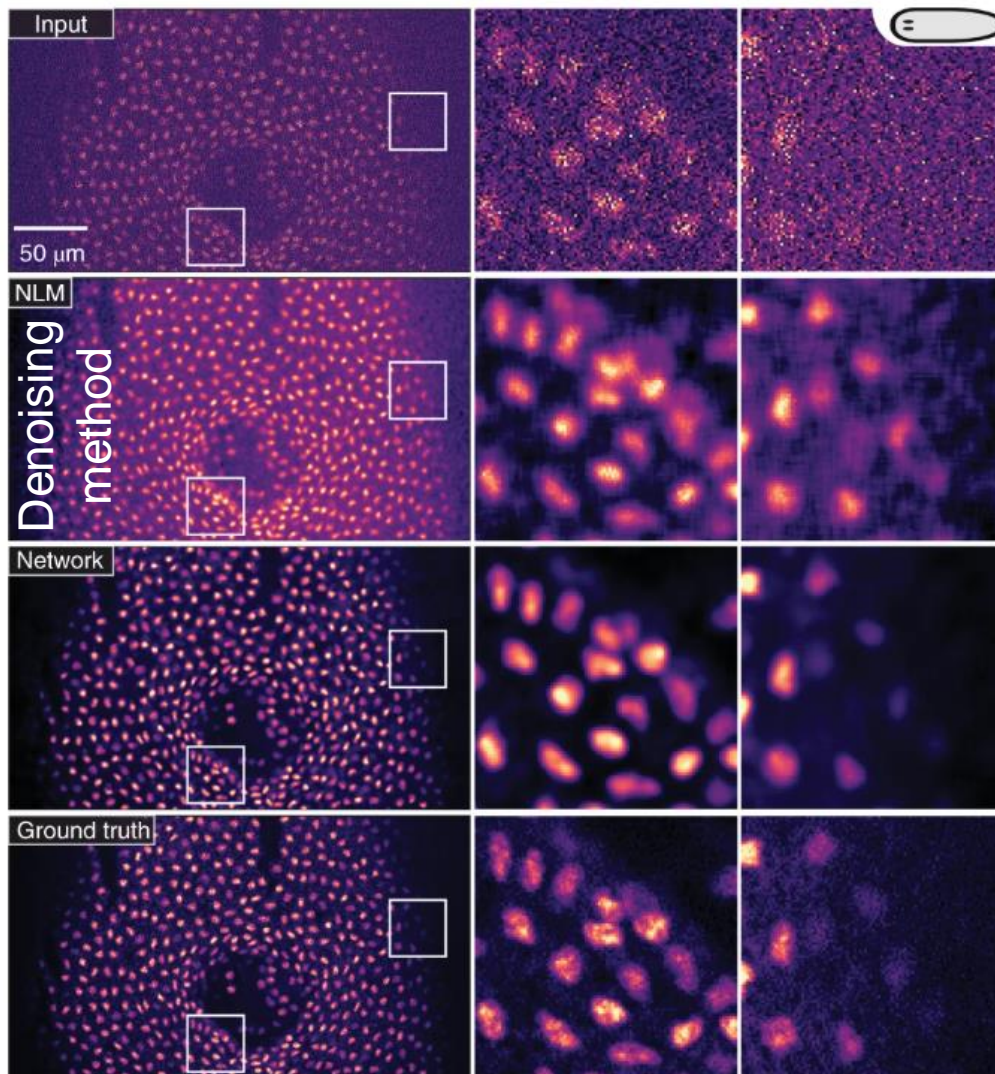


# The machinery: U-Net

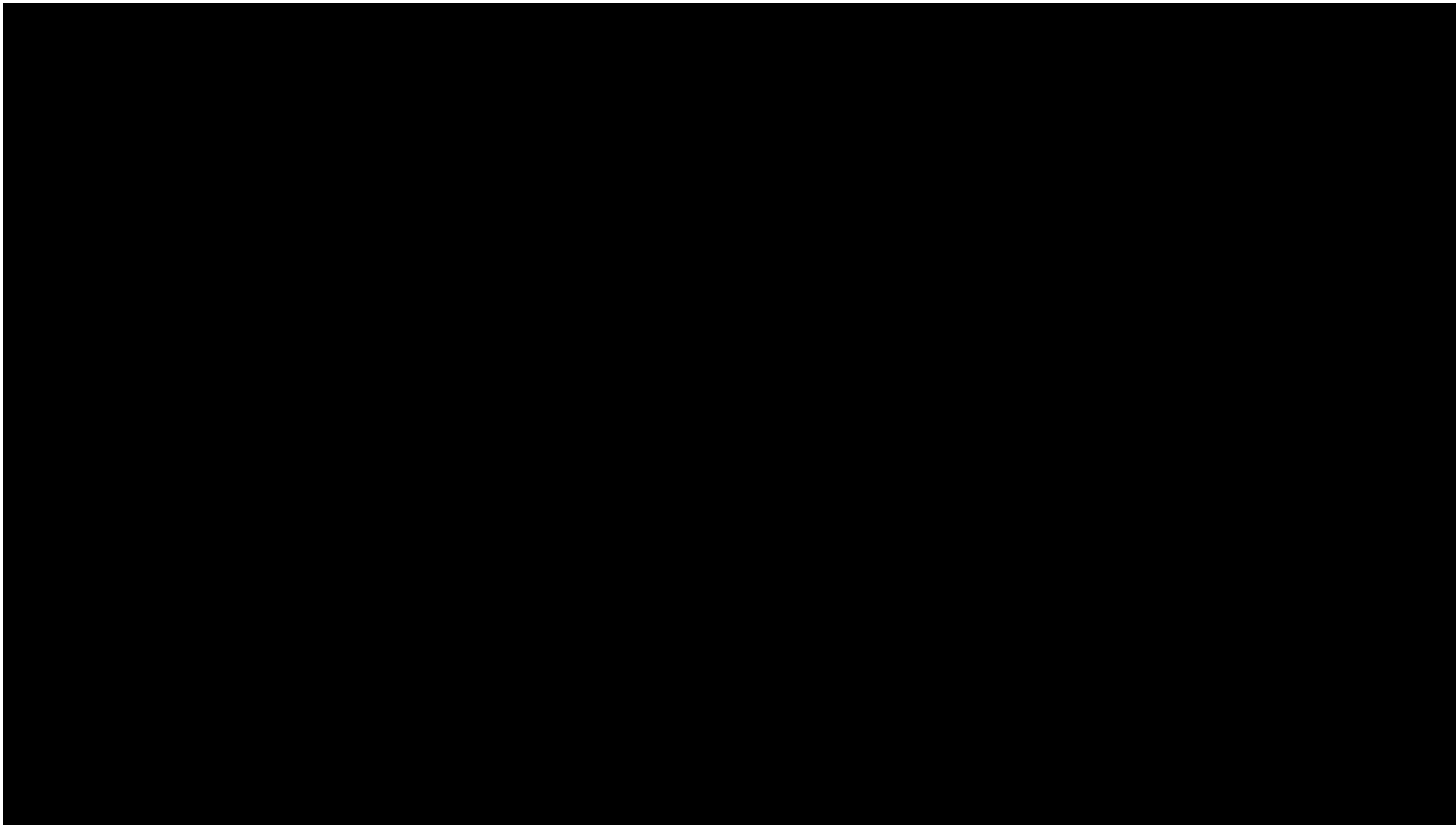




# Results

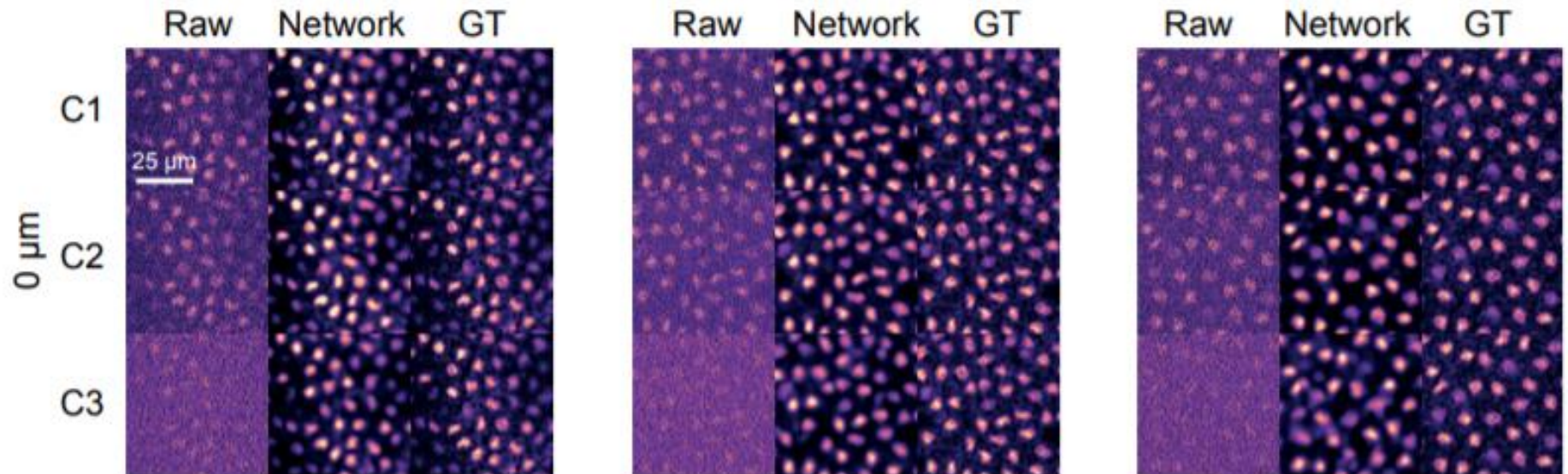
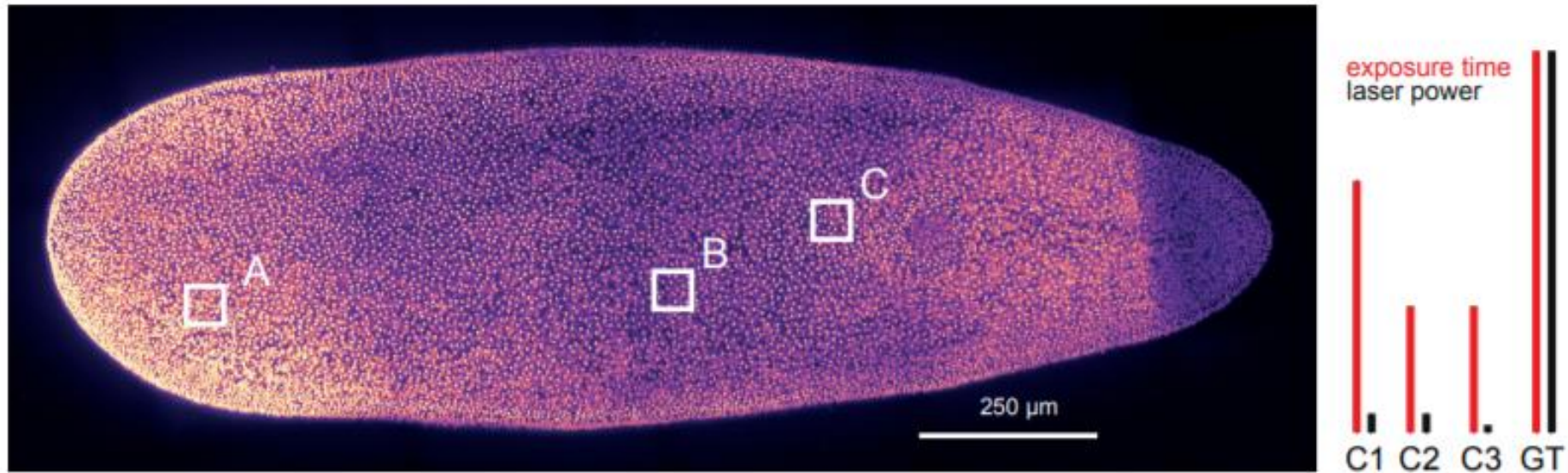






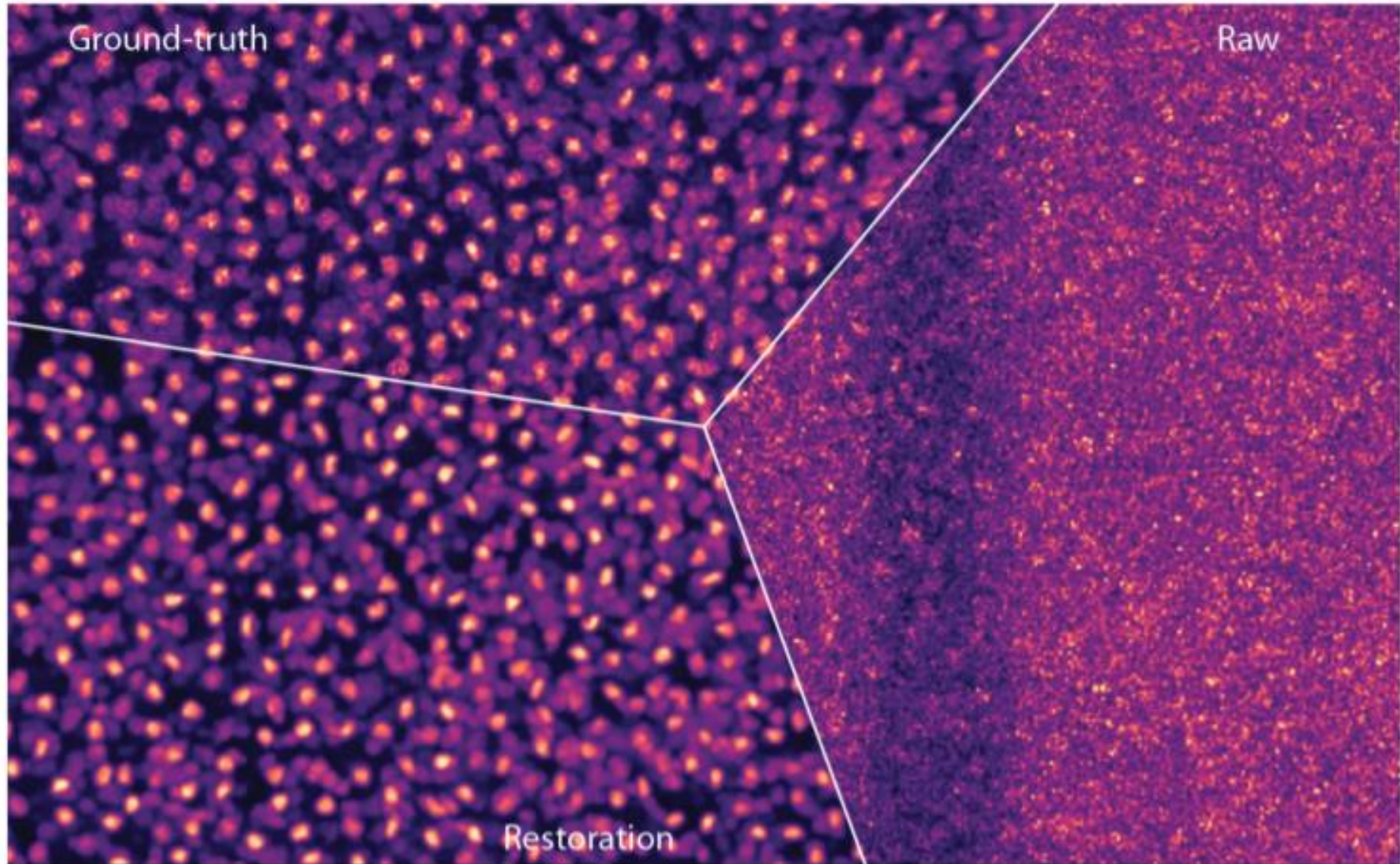


# Restoration of low-SNR images



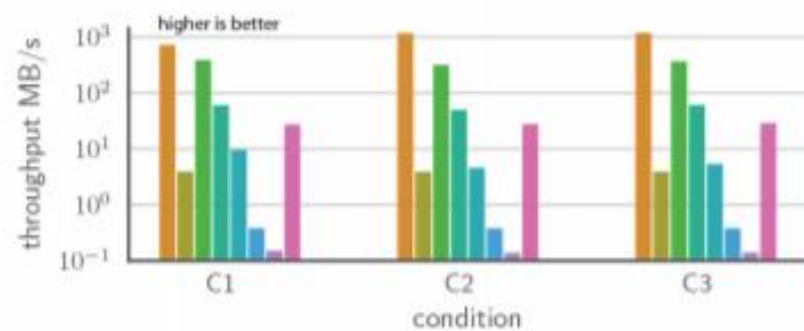
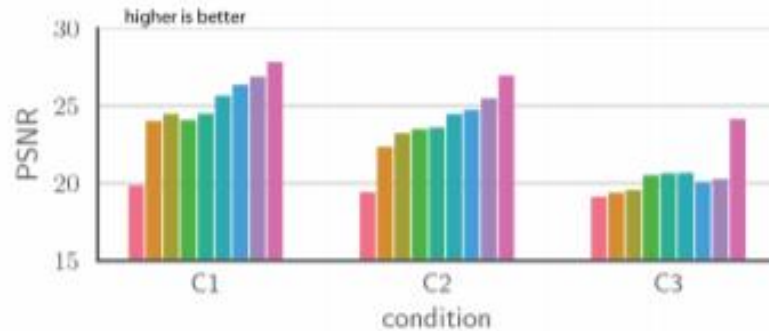
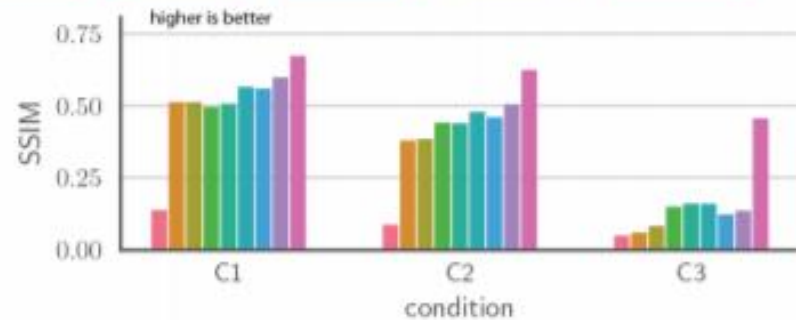
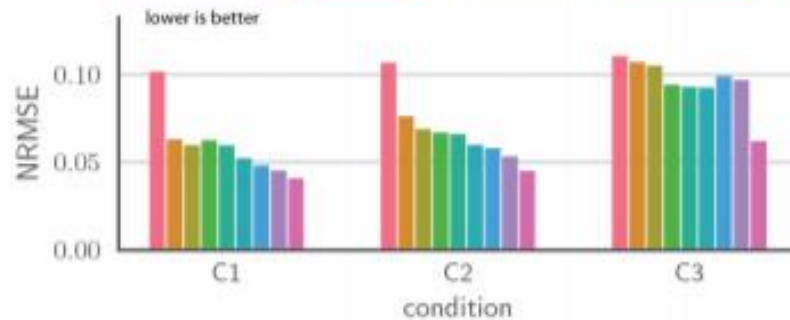
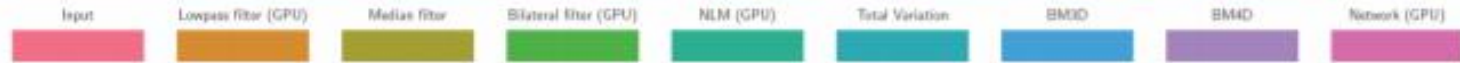
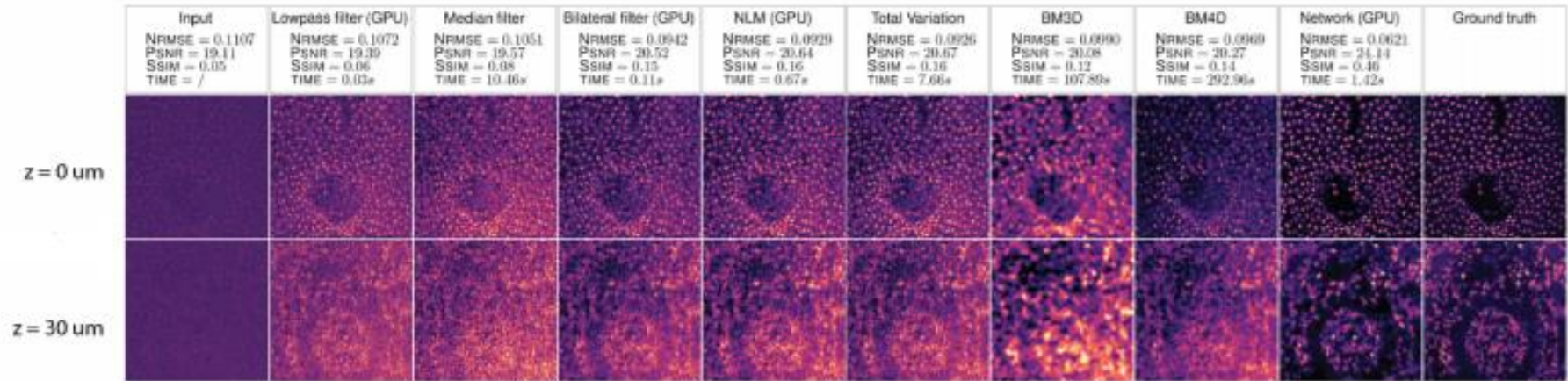


# Restoration of low-SNR images





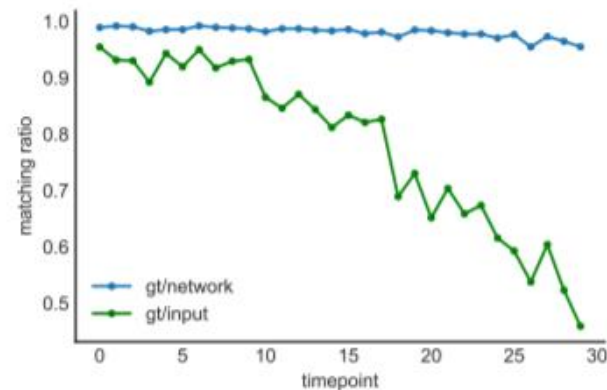
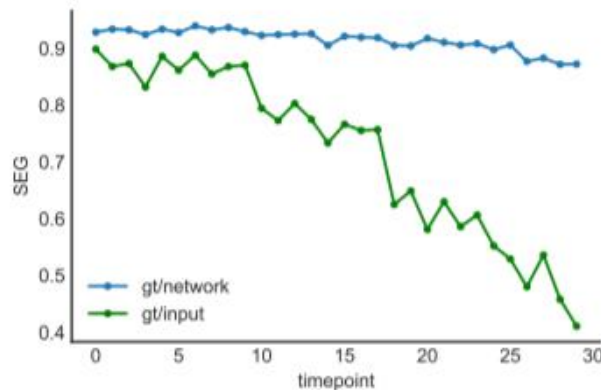
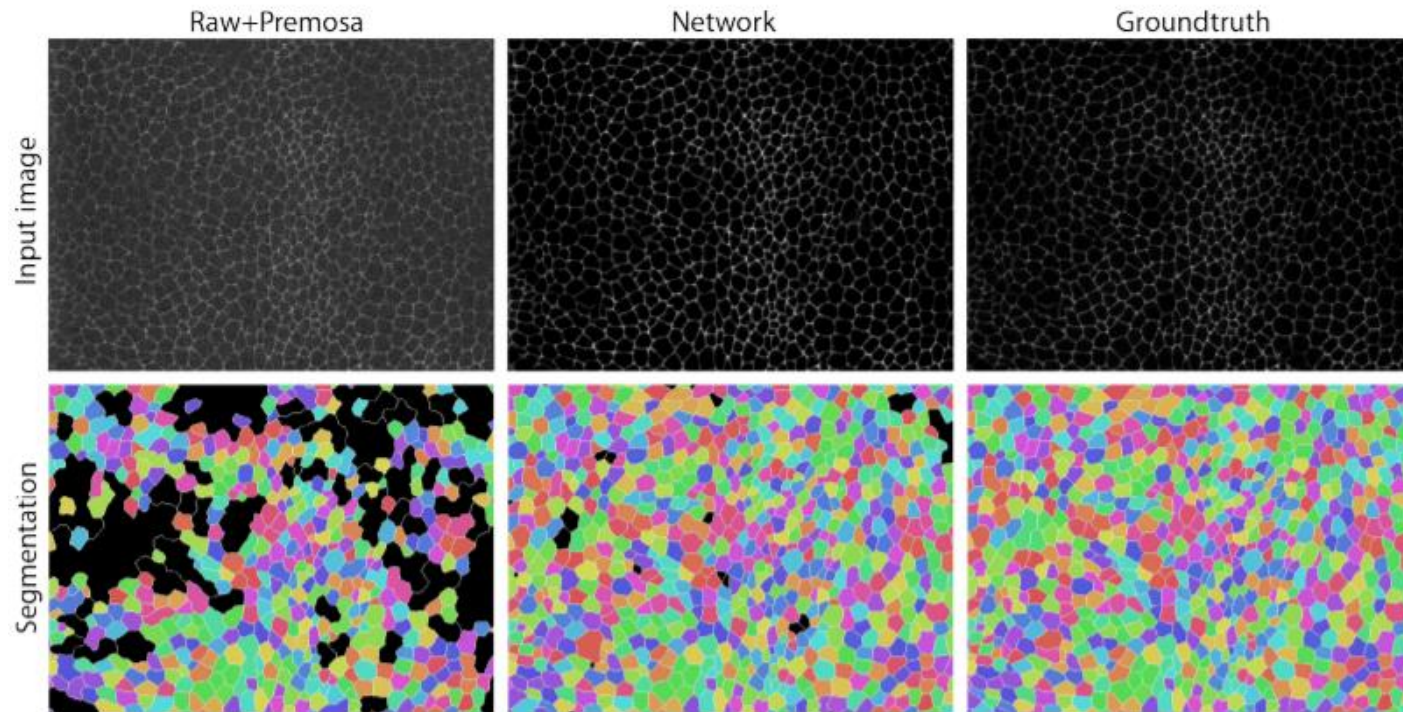
# Comparison: different denoising methods





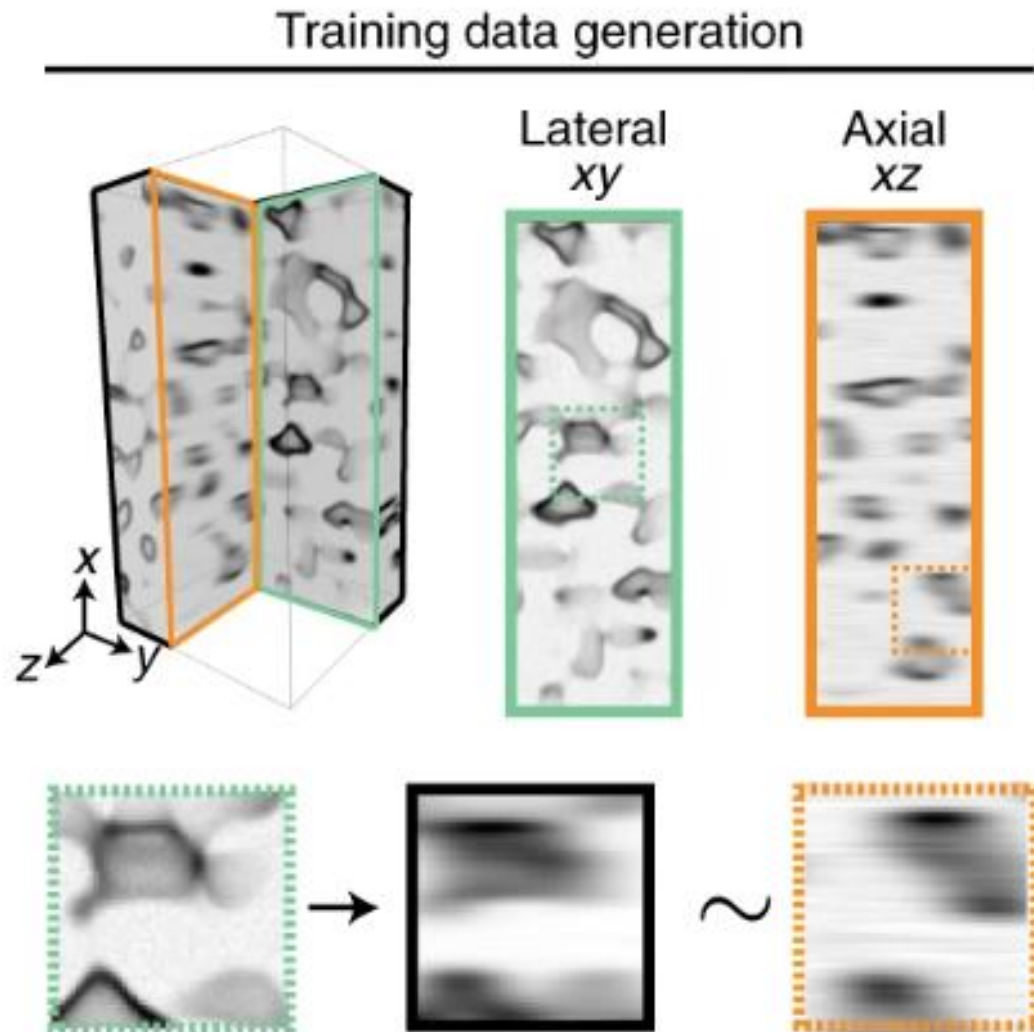
# CARE improves downstream analysis

Cell segmentation and tracking of a projected time-lapse



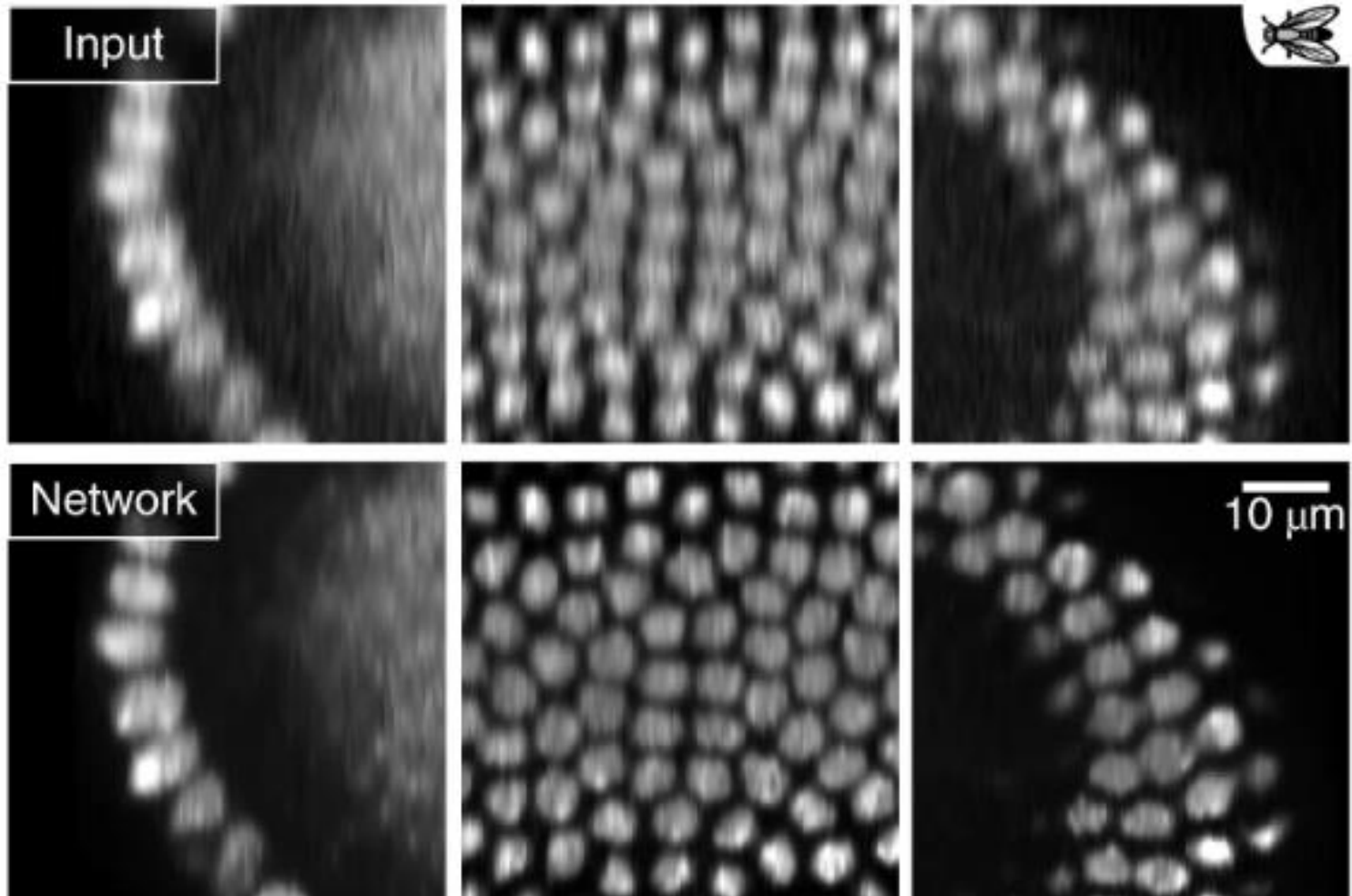


# Image restoration of unseen axial slices with semi-synthetic training data



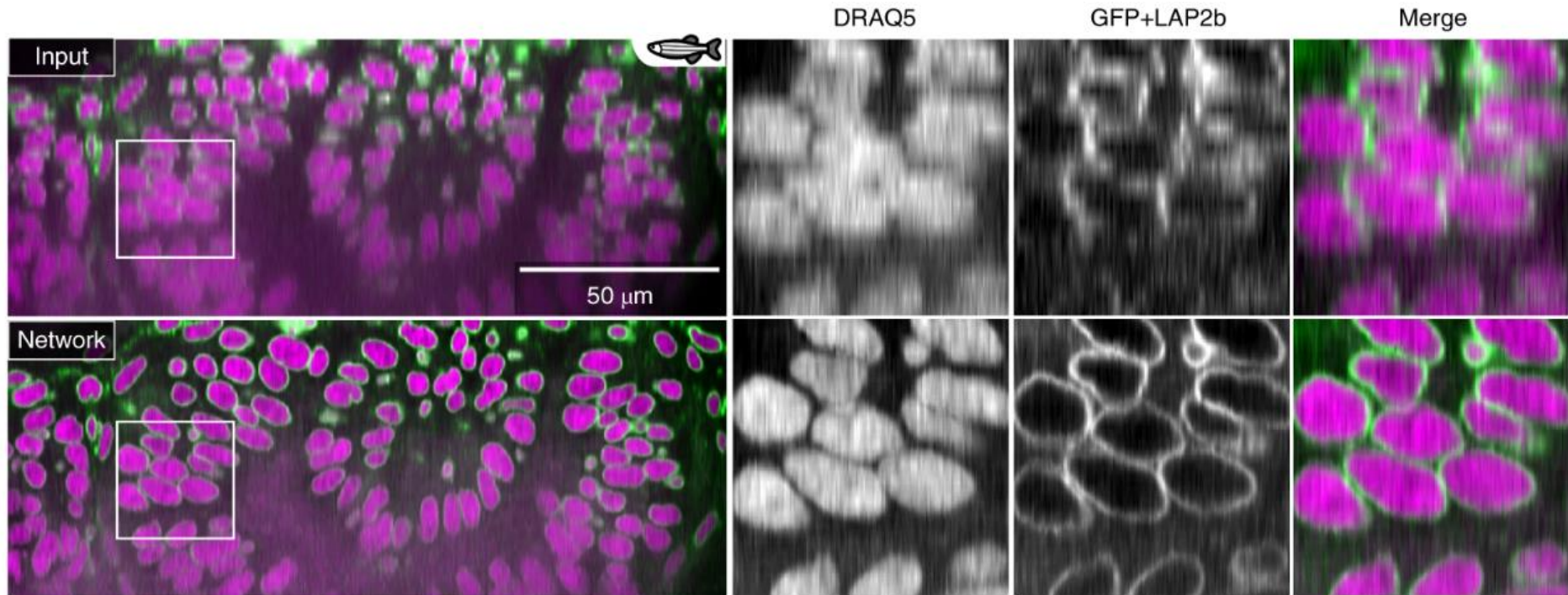


# CARE increase axial resolution of acquired volumes



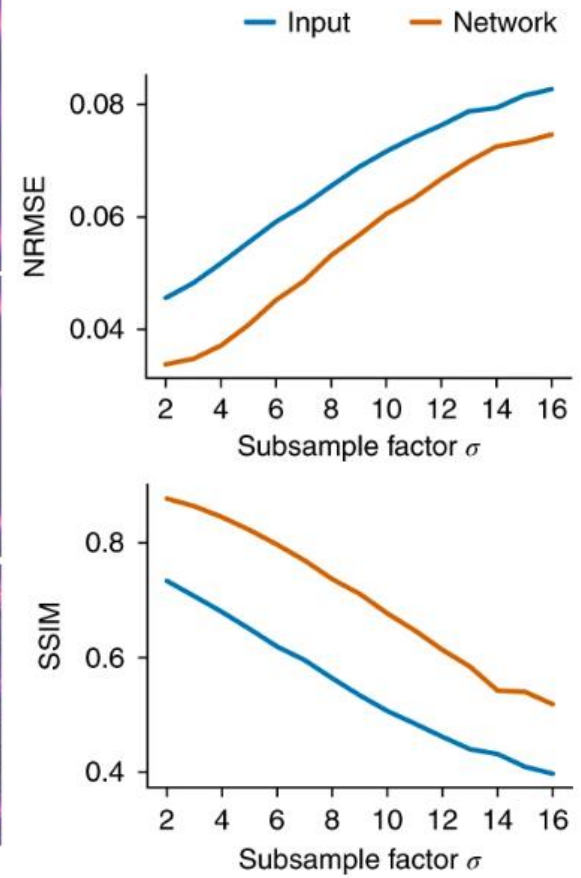
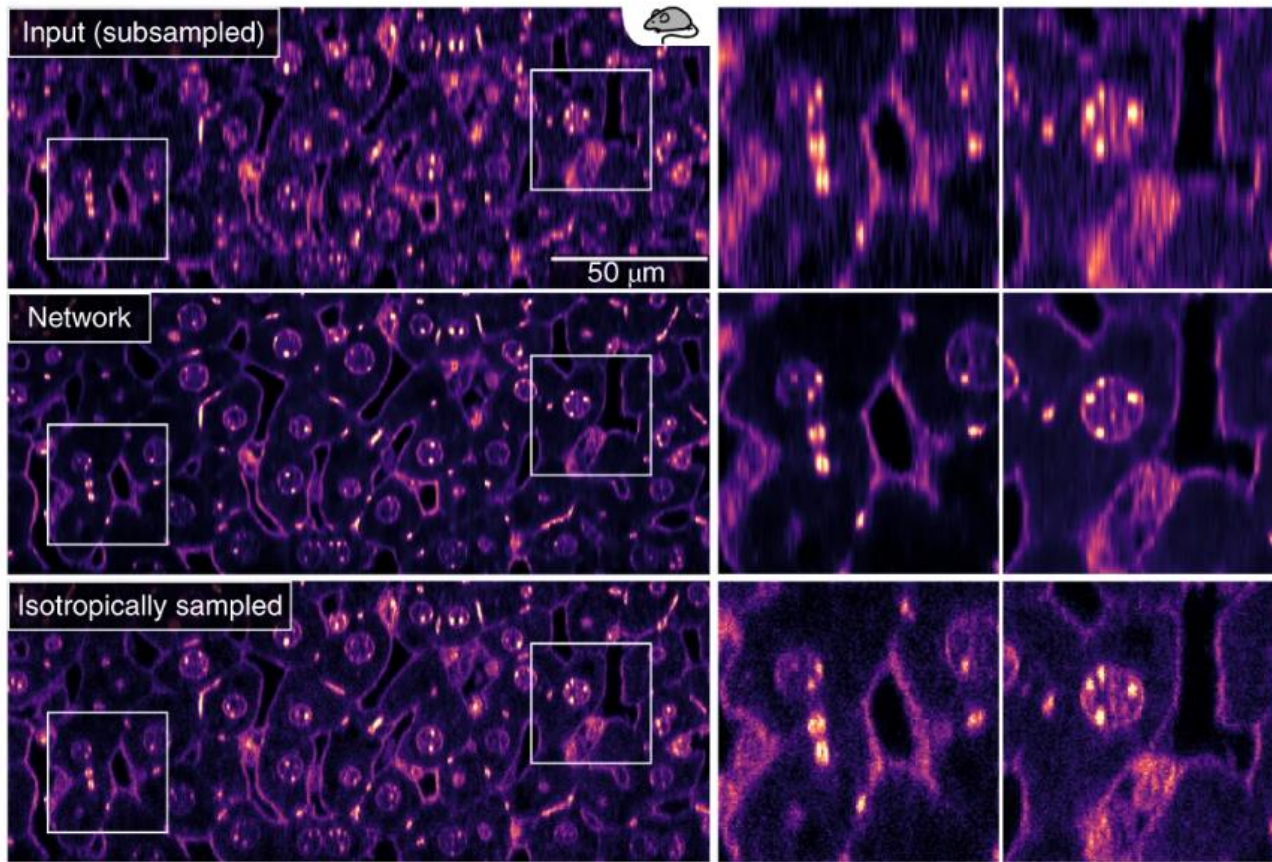


# CARE increase axial resolution of acquired volumes



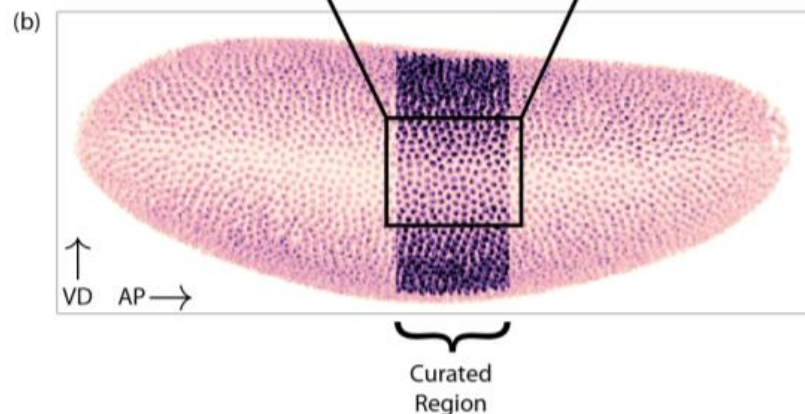
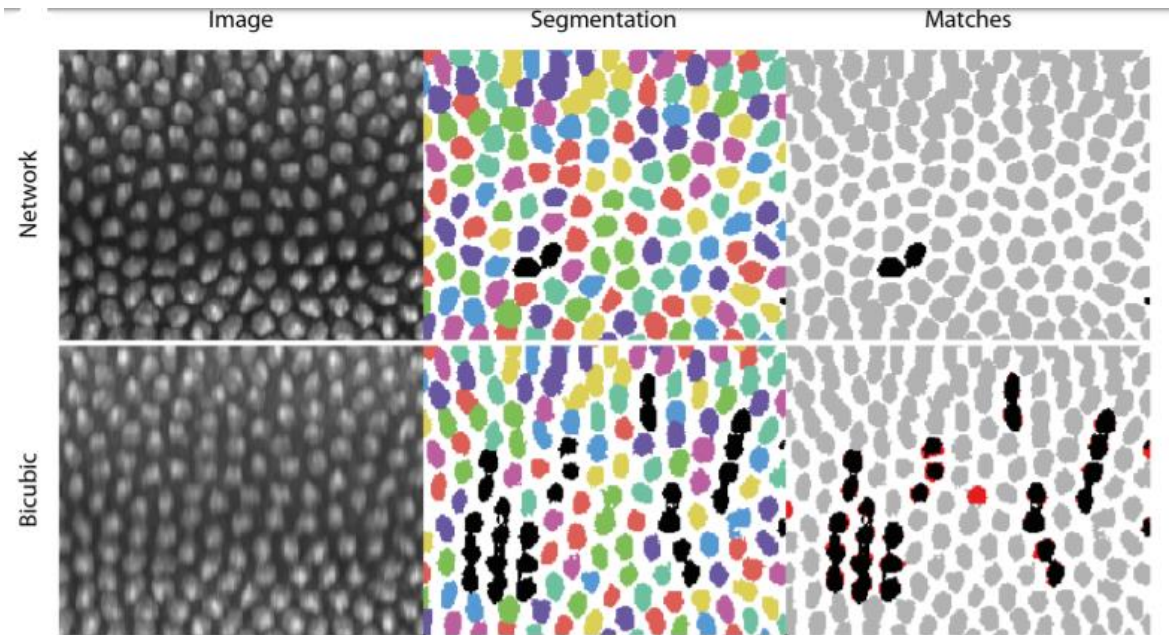


# CARE increase axial resolution of acquired volumes



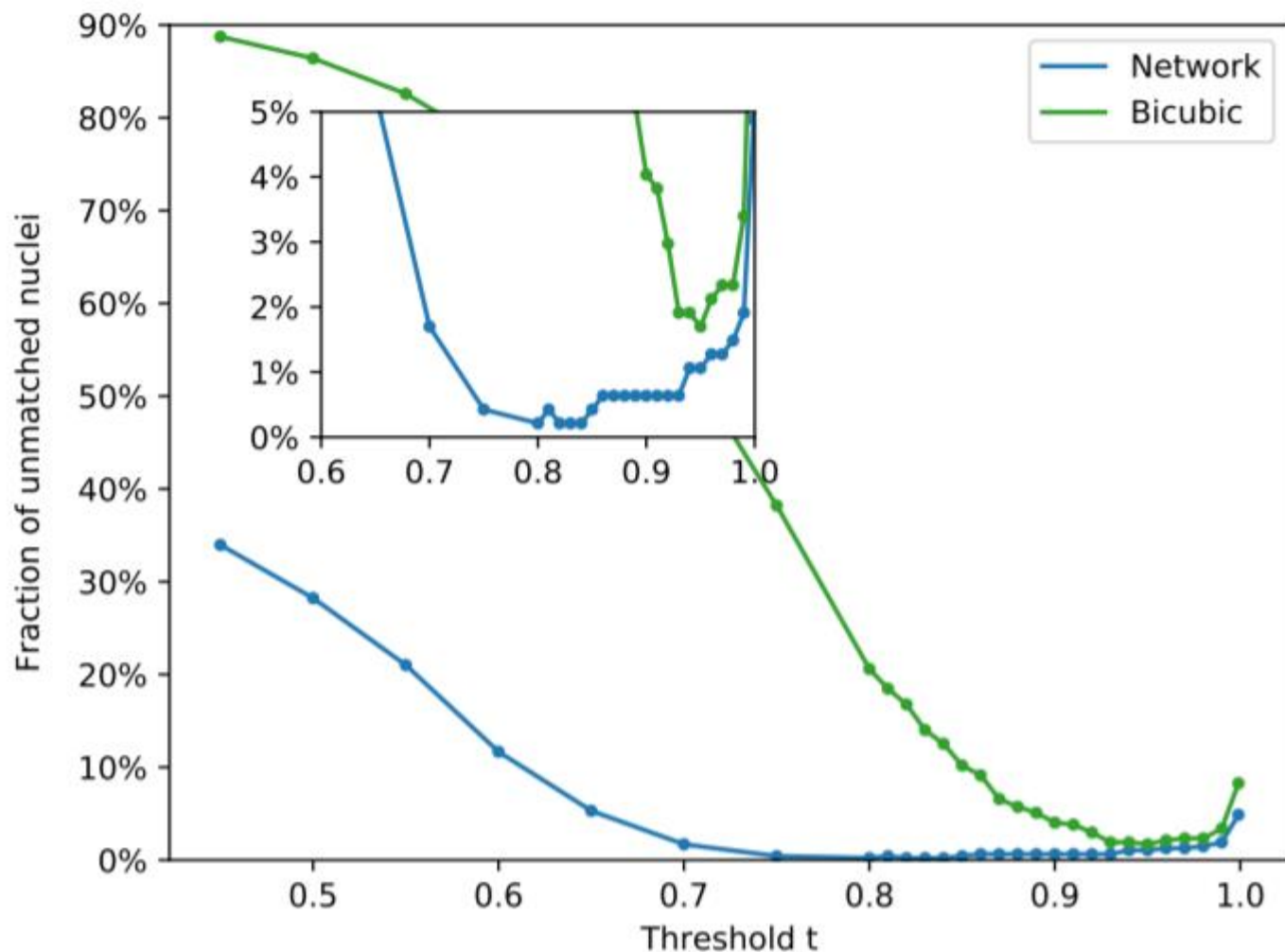


# Anisotropic-to-isotropic restoration leads to improved segmentation





# Anisotropic-to-isotropic restoration leads to improved segmentation



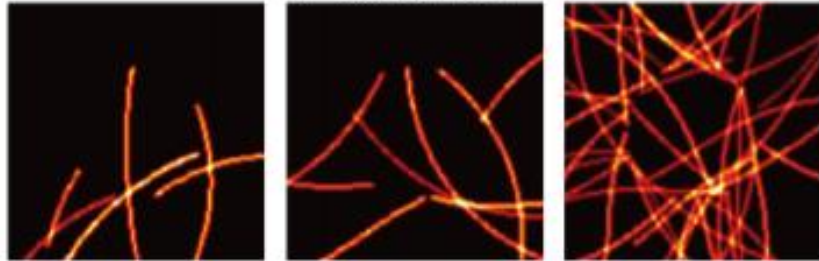


# Synthetically generated training data

PSF, camera noise, and background auto-fluorescence

Microtubules

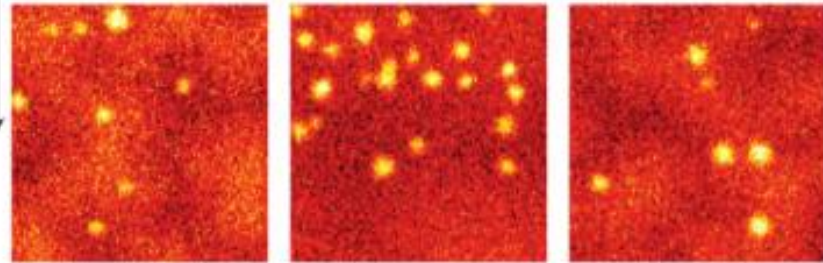
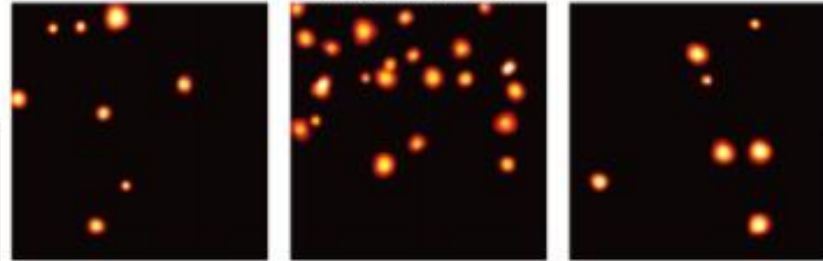
Y Ground Truth



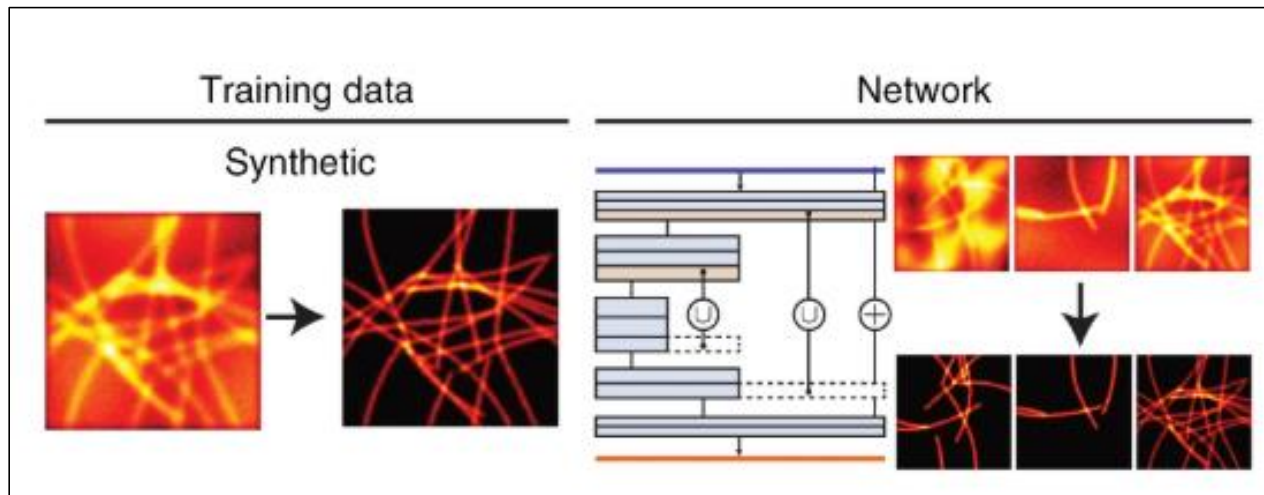
X Synthetic image

Granules

Y Ground Truth



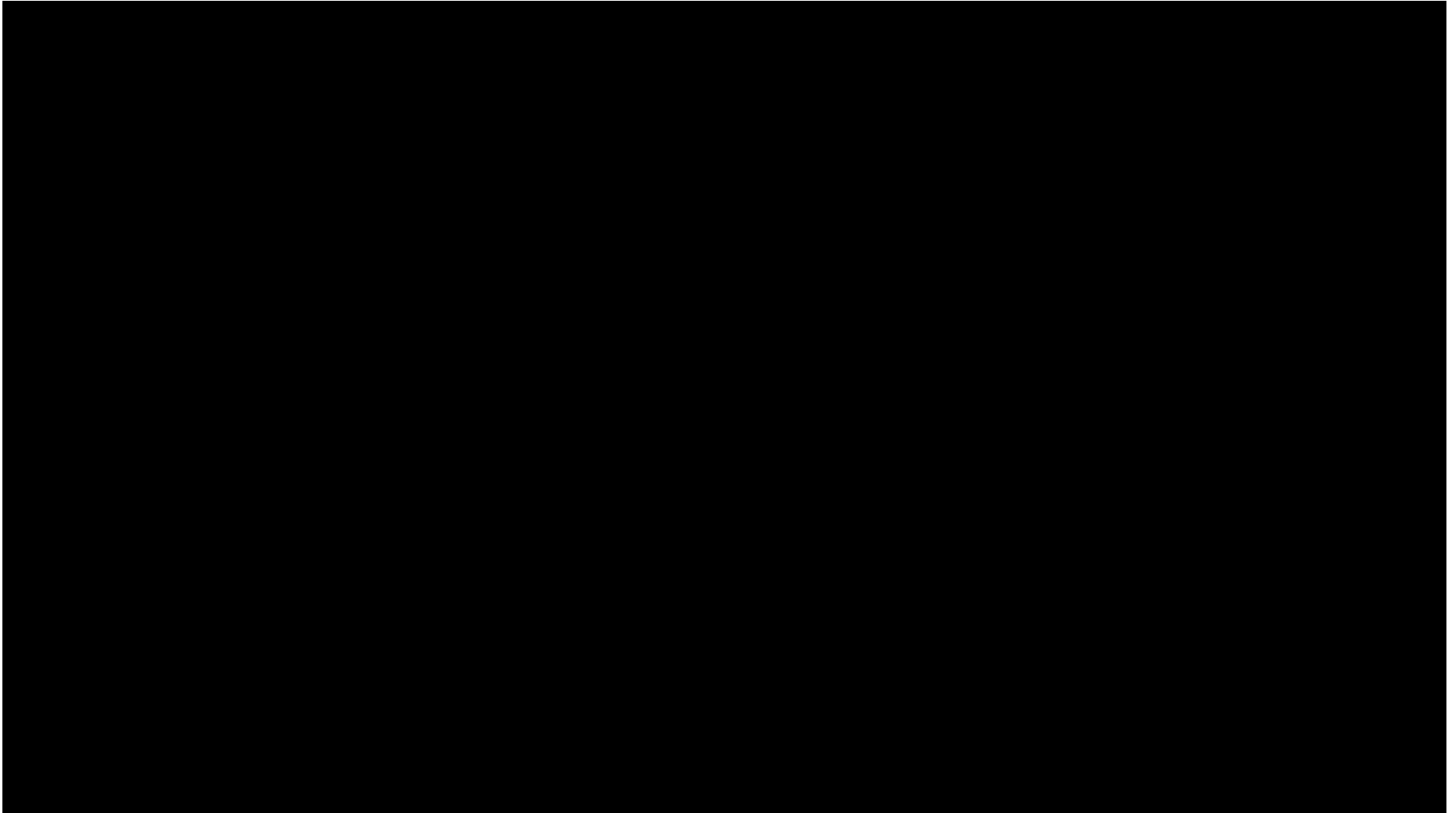
X Synthetic image





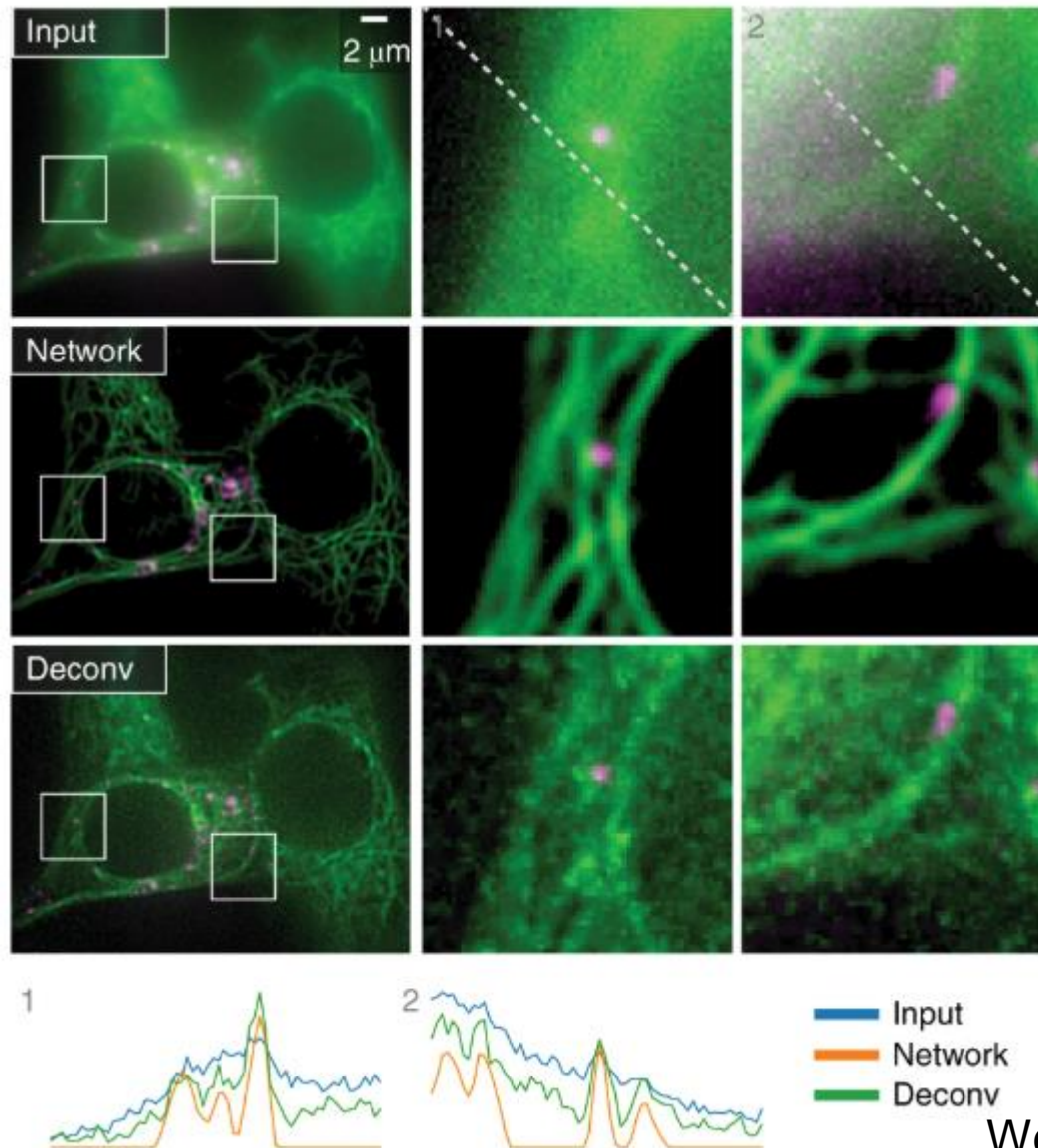
# Image restoration with synthetic training data

Restoring sub-diffraction structures using only widefield images





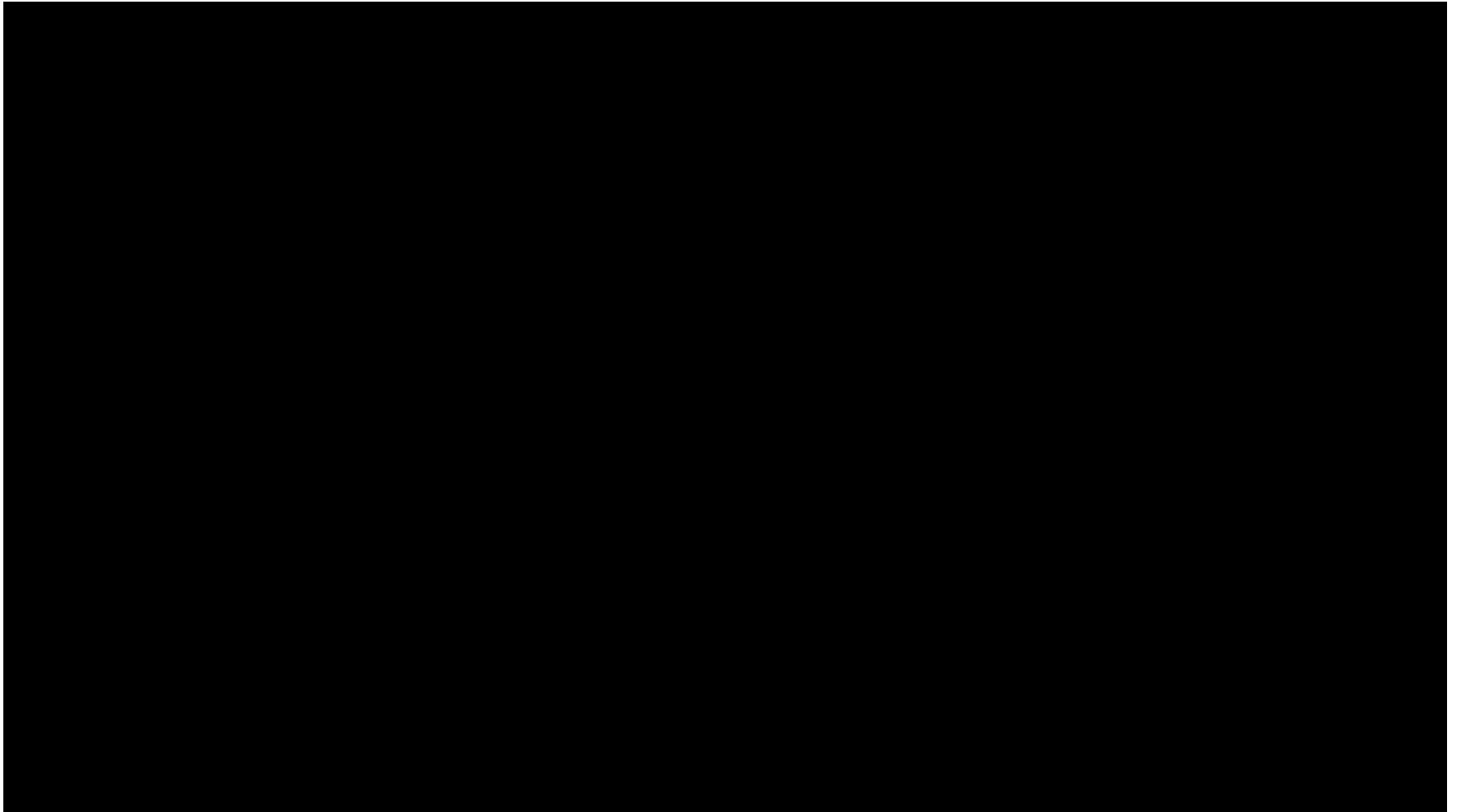
# Improved performance of the CARE network over deconvolution





# CARE vs. super-resolution radial fluctuations (SRRF)

Enabling 20-fold faster imaging





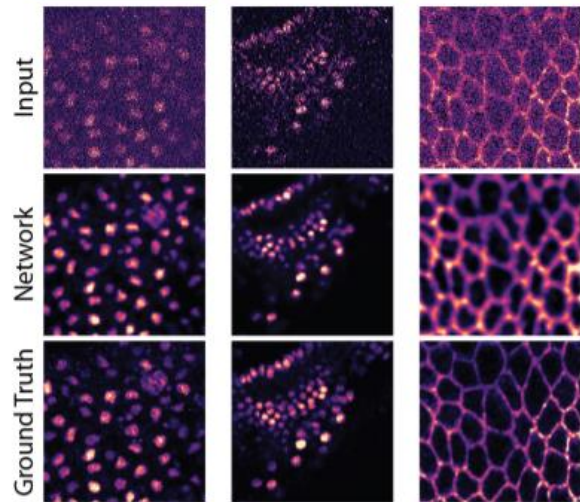
# Cross-application of trained care networks

3D Denoising Network

Trained on Planaria

Applied to:

Planaria Tribolium Flywing

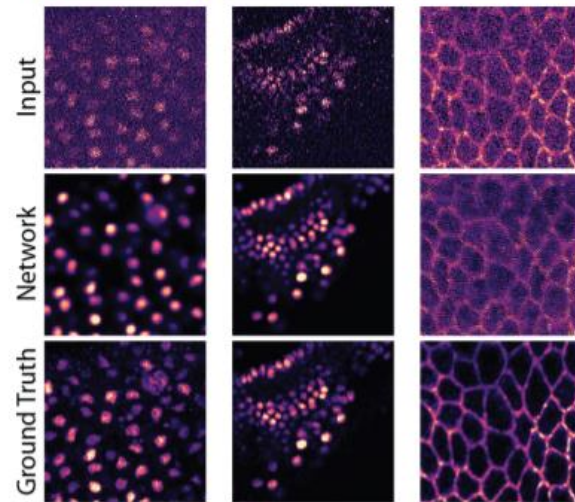


3D Denoising Network

Trained on Tribolium

Applied to:

Planaria Tribolium Flywing

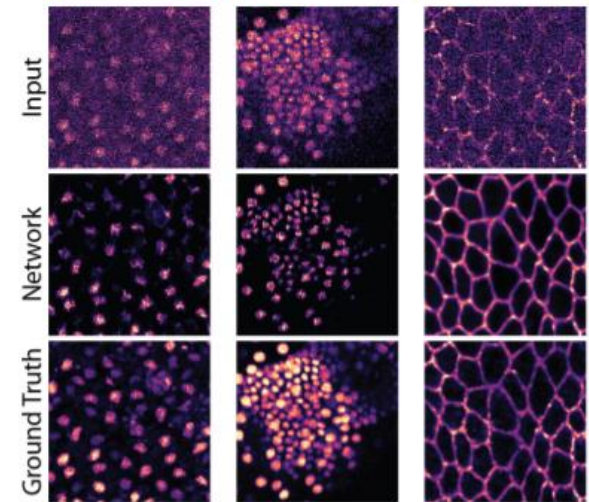


3D Surface Projection Network

Trained on Flywing

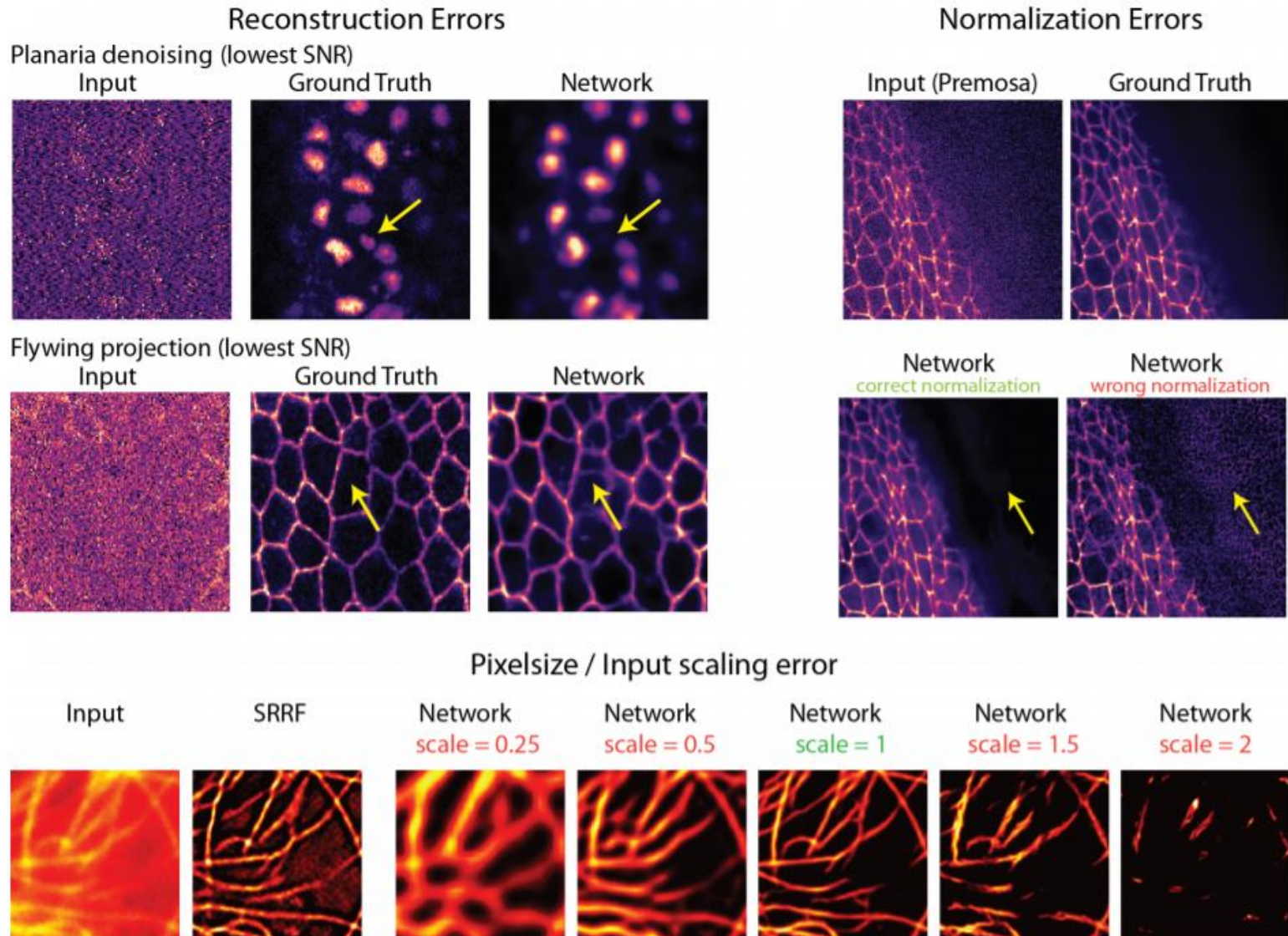
Applied to:

Planaria Tribolium Flywing





# Minimal 'hallucination' effects

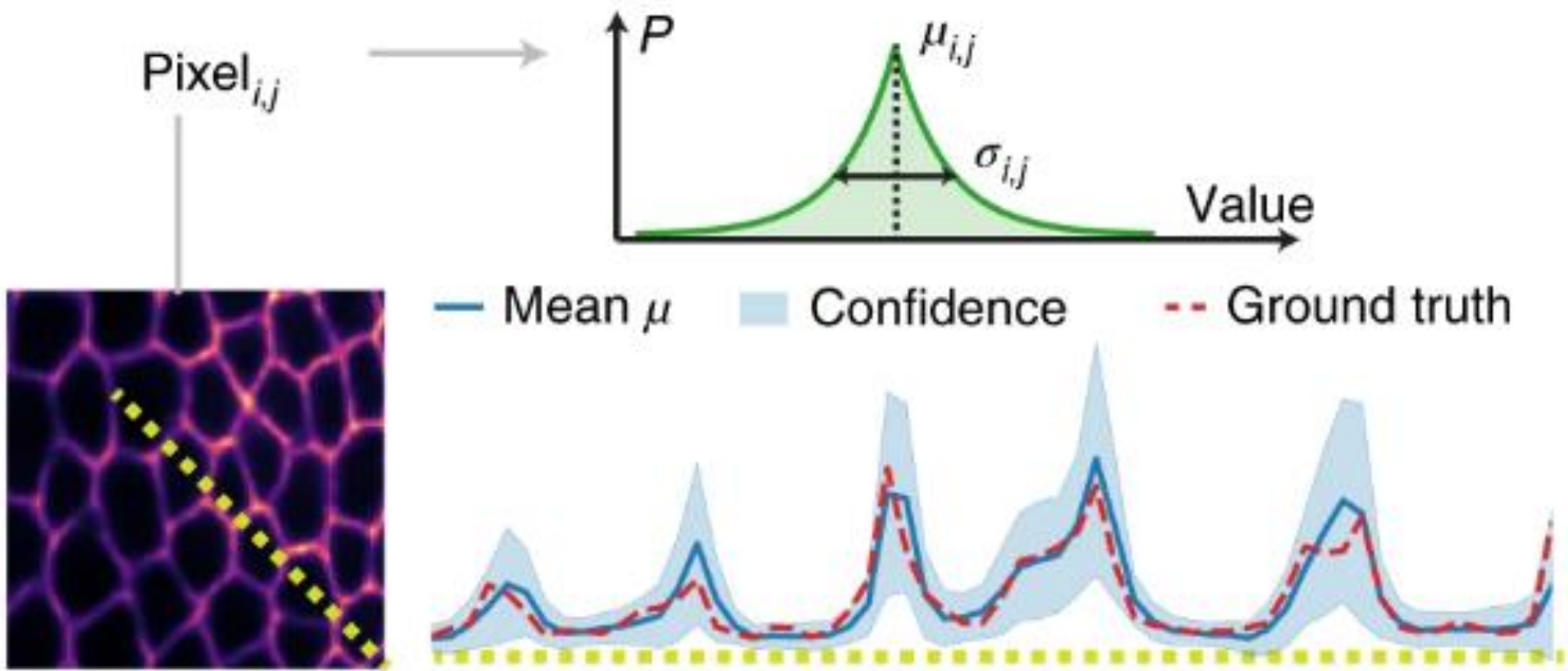




# Reliability of image restoration

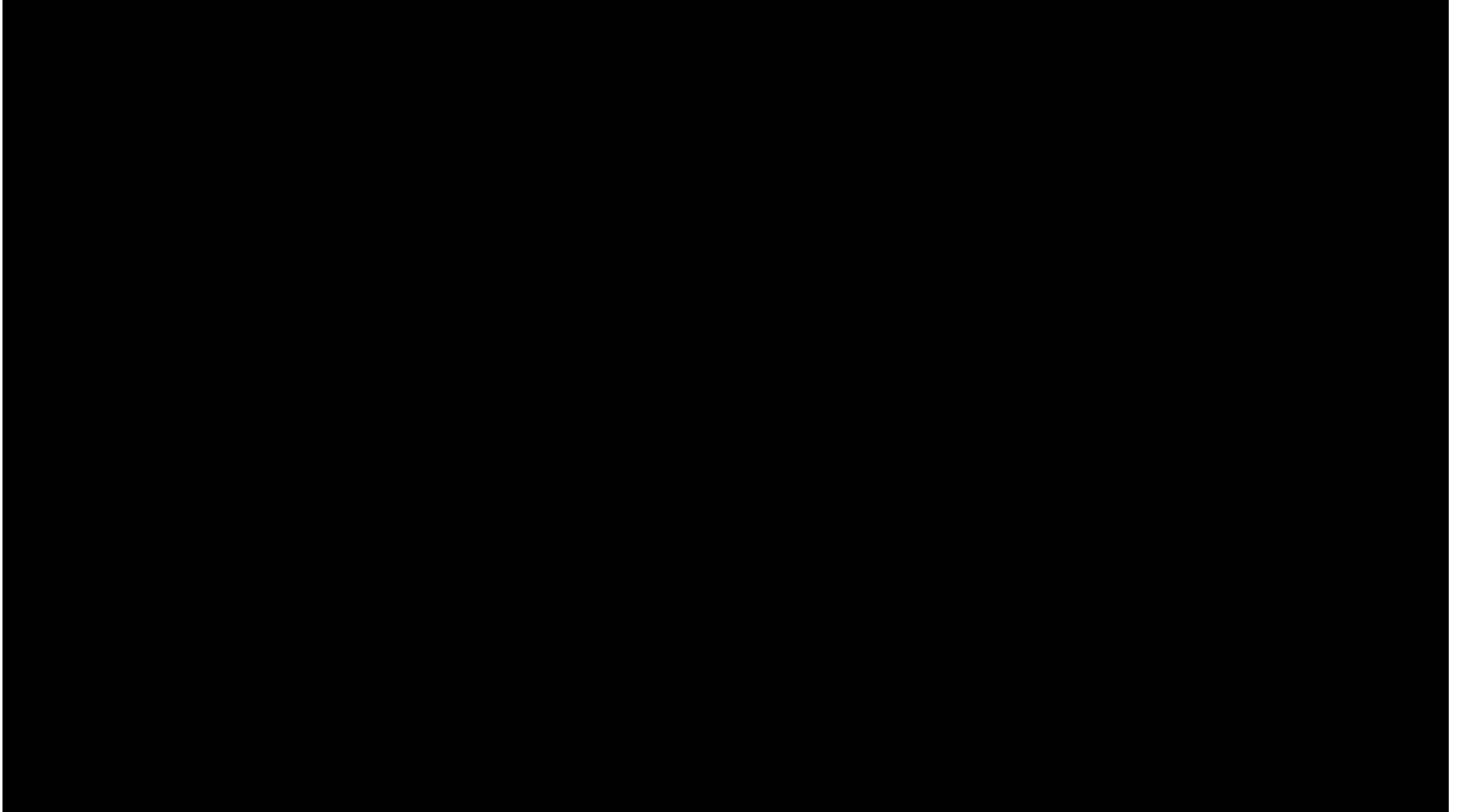
Calculating pixel-wise confidence intervals

Predicting pixel-wise distributions



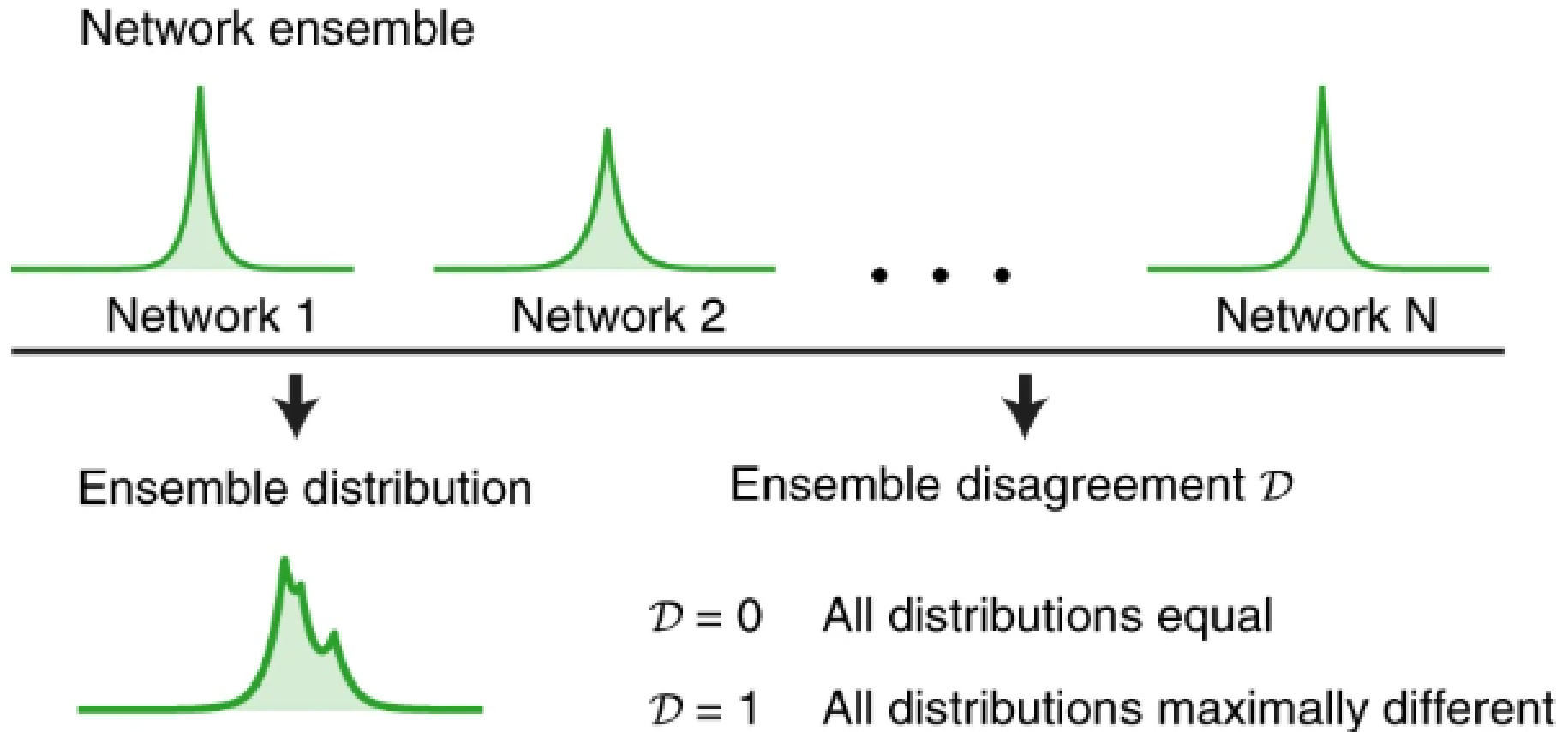


Visualizing the uncertainty by random  
sampling pixel intensities from their  
respective distributions



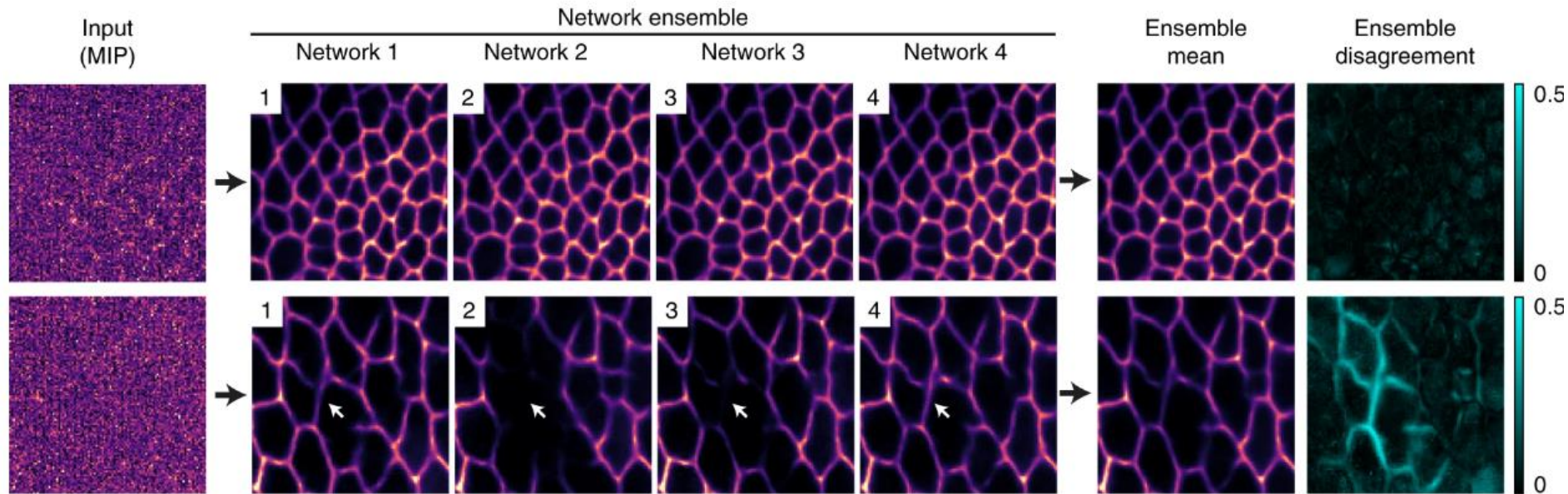


# Assessing reliability via consistency of several trained models

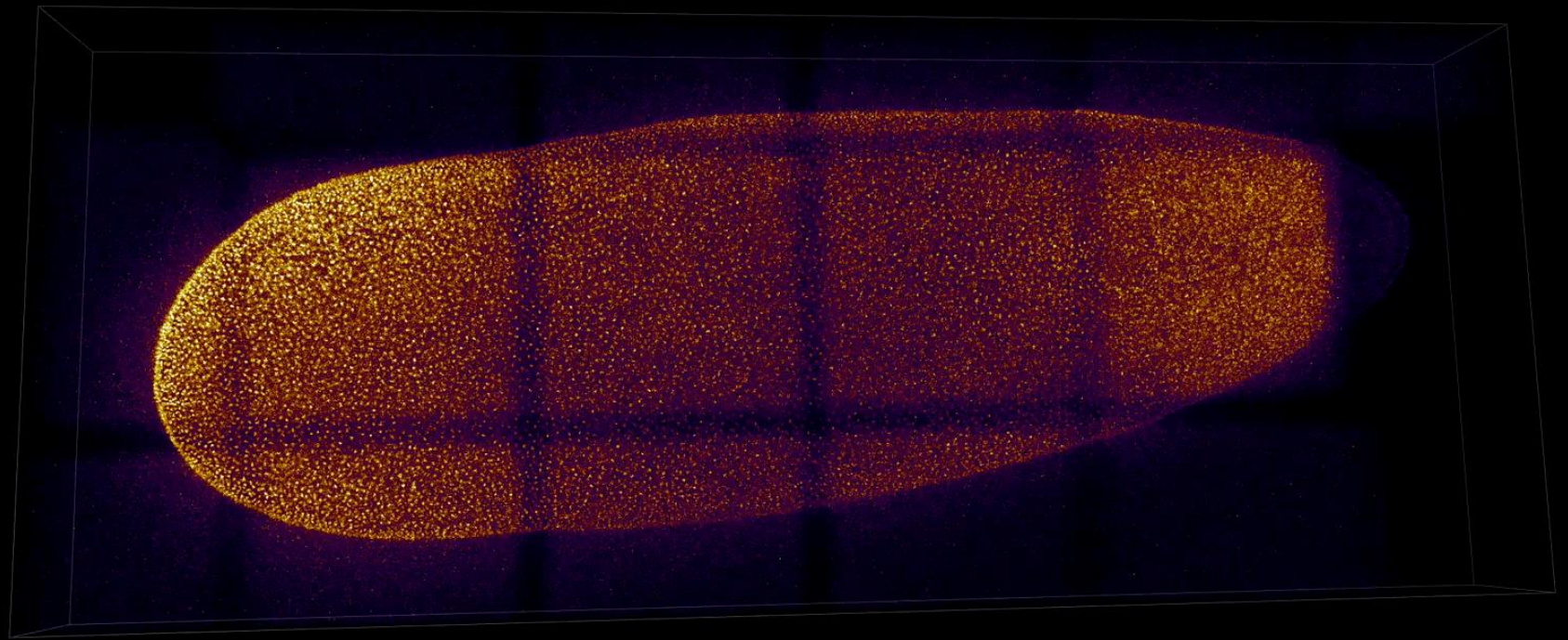




# Assessing reliability via consistency of several trained models









# Usability is important!



## CSBDeep, a toolbox for Content-aware Image Restoration (CARE)

Fluorescence microscopy is a key driver of discoveries in the life-sciences, with observable phenomena being limited by the optics of the microscope, the chemistry of the fluorophores, and the maximum photon exposure tolerated by the sample. These limits necessitate trade-offs between imaging speed, spatial resolution, light exposure, and imaging depth. In this work we show how deep learning enables biological observations beyond the physical limitations of microscopes. On seven concrete examples we illustrate how microscopy images can be restored even if 60-fold fewer photons are used during acquisition, how isotropic resolution can be achieved even with a 10-fold under-sampling along the axial direction, and how diffraction-limited structures can be resolved at 20-times higher frame-rates compared to state-of-the-art methods. All described restoration networks are freely available as open source software in Fiji and KNIME.

## How to start

- To use CSBDeep in Python, follow [this guide](#).
- To use CSBDeep in Fiji, follow [this guide](#).
- To use CSBDeep in KNIME, follow [this guide](#).
- (If you have been given access, try CSBDeep on our [Paperspace server](#).)



<https://csbdeep.bioimagecomputing.com/>  
<https://github.com/csbdeep/csbdeep>



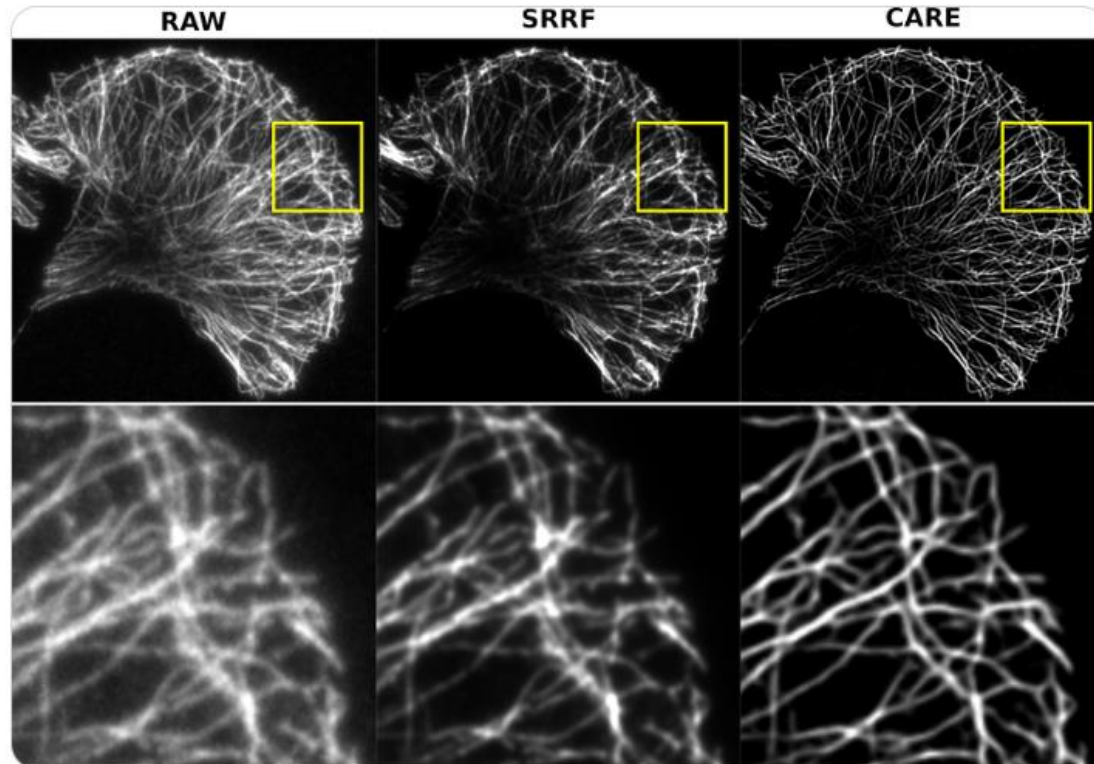
# (Sometimes) even works out of the box!



Guillaume Jacquemet  
@guijacquemet



#CARE vs #SRRF on my microtubule images (TIRF 100x).  
#CARE work beautifully. I look forward to train it to  
work on filopodia ! I already have images ready !  
[biorxiv.org/content/early/...](https://doi.org/10.1101/248859)



12:09 PM · Jan 4, 2018 · [Twitter Web Client](#)

Guillaume Jacquemet, <https://twitter.com/guijacquemet/status/948859038795780101>



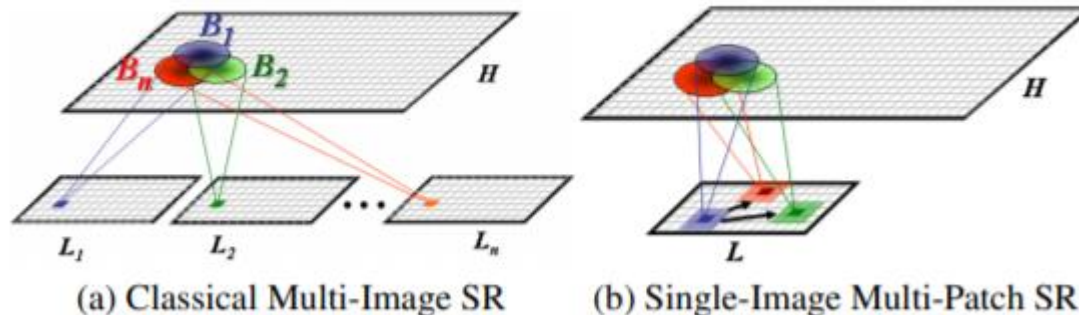
# Summary

- CARE leverages experiment-specific information to imaging tradeoffs
- Matched images, semi-synthetic and synthetic training data
- High quality denoising models can be trained **without the availability of clean ground truth data**
- Great for downstream analysis **not for intensity-based measurements!**
- Could work well “out of the box”
- Usable by others!



# “Super resolution” in the context of computer vision

- Recover a high resolution image from one or more low resolution input images
- Classical multi-image super resolution: combining images obtained at subpixel misalignments (Irani & Peleg, 1991)
- Example-Based super-resolution: learning correspondence between matching low and high resolution image patches (Freeman, Pasztor & Carmichael, 2000)



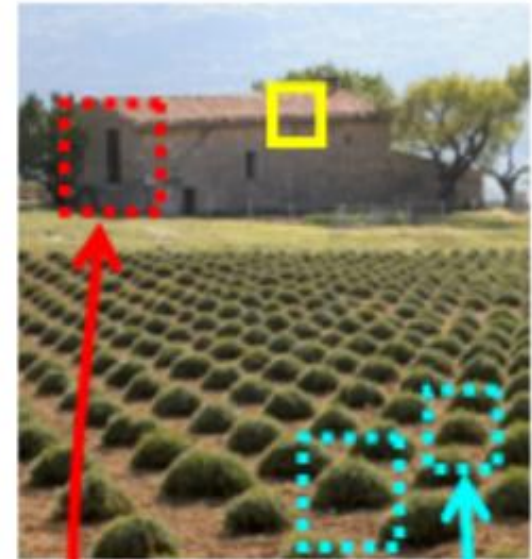
Glasner, Bagon and Irani (2009)



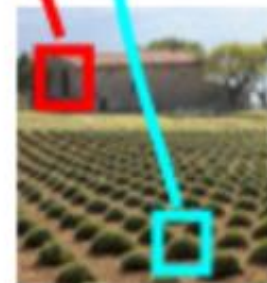
Input image  $I$



Various scales of  $I$



(computer vision) super  
resolution from a single  
image





# (One) result



(a) Input image (scaled for display).

(b) Bicubic interpolation ( $\times 2$ ).

(c) Within image repetitions ( $\times 2$ ).

(d) Unified single-image SR ( $\times 2$ ).



2009 vs. 2018

A conceptual lag of (almost) a **decade**  
between computer vision and  
microscopy!

There might be other opportunities out  
there..



# Handling noisy images: new extensions from the Jug lab

(could be picked as a student presentation)

- Noise2noise: a mapping between pairs of independently degraded versions of the same training image, requires availability of pairs of noisy images (Lehtinen et. al., 2018)
- Noise2Void: self-supervised training method with "blind-spot" networks – excluding a pixel and learning a mapping from the noisy image to the missing pixel (Krull, Buchholz and Jug, 2019)
- Probabilistic Noise2Void: predict per-pixel intensity distributions (Krull, Viřcar and Jug, 2019)



# Examples of mapping matched image pairs for image enhancement

- Super resolution: ANNA-PALM, deep STORM, “cross modality” (resolution), PSSR
- 2D → 3D refocusing, Deep-Z

Could be picked as a student presentation



# ANNA-PALM: DL accelerates super-resolution localization microscopy

- ANNA-PALM (slightly) preceded CARE
- exploits the structural redundancy of most biological images to reconstruct high-quality images from under-sampled localization microscopy data
- Reducing total number of frames and independent localizations without trading off spatial resolution

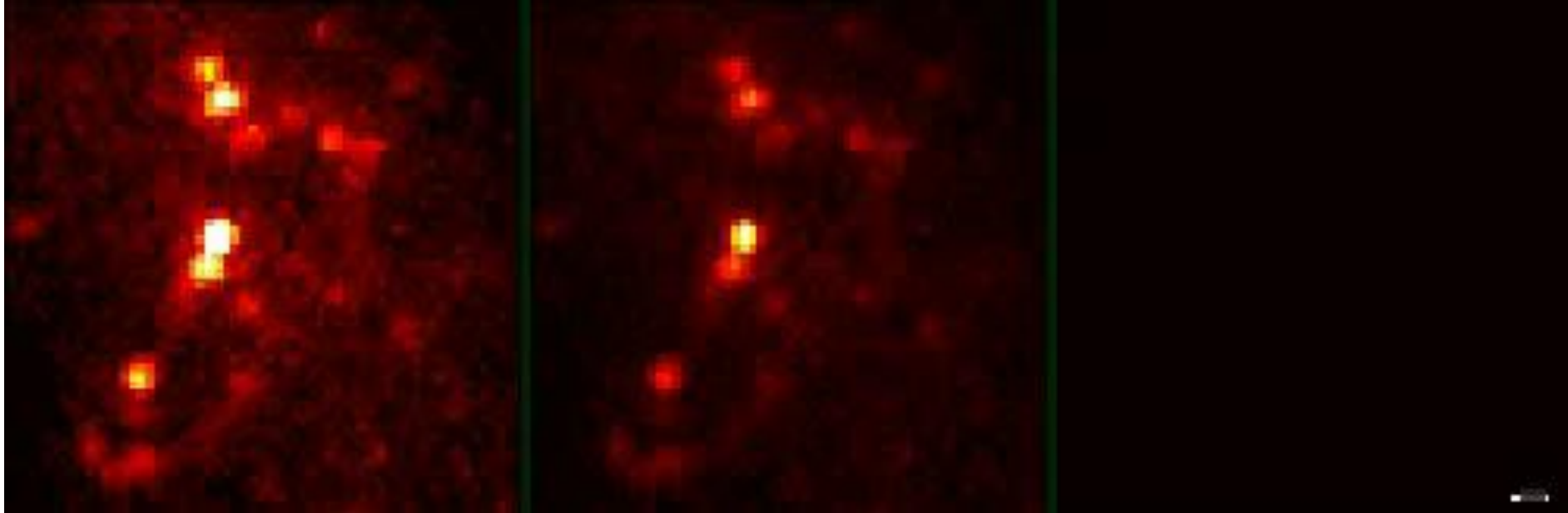


# Photoactivated localization microscopy (PALM)

diffraction-limited  
image sequences

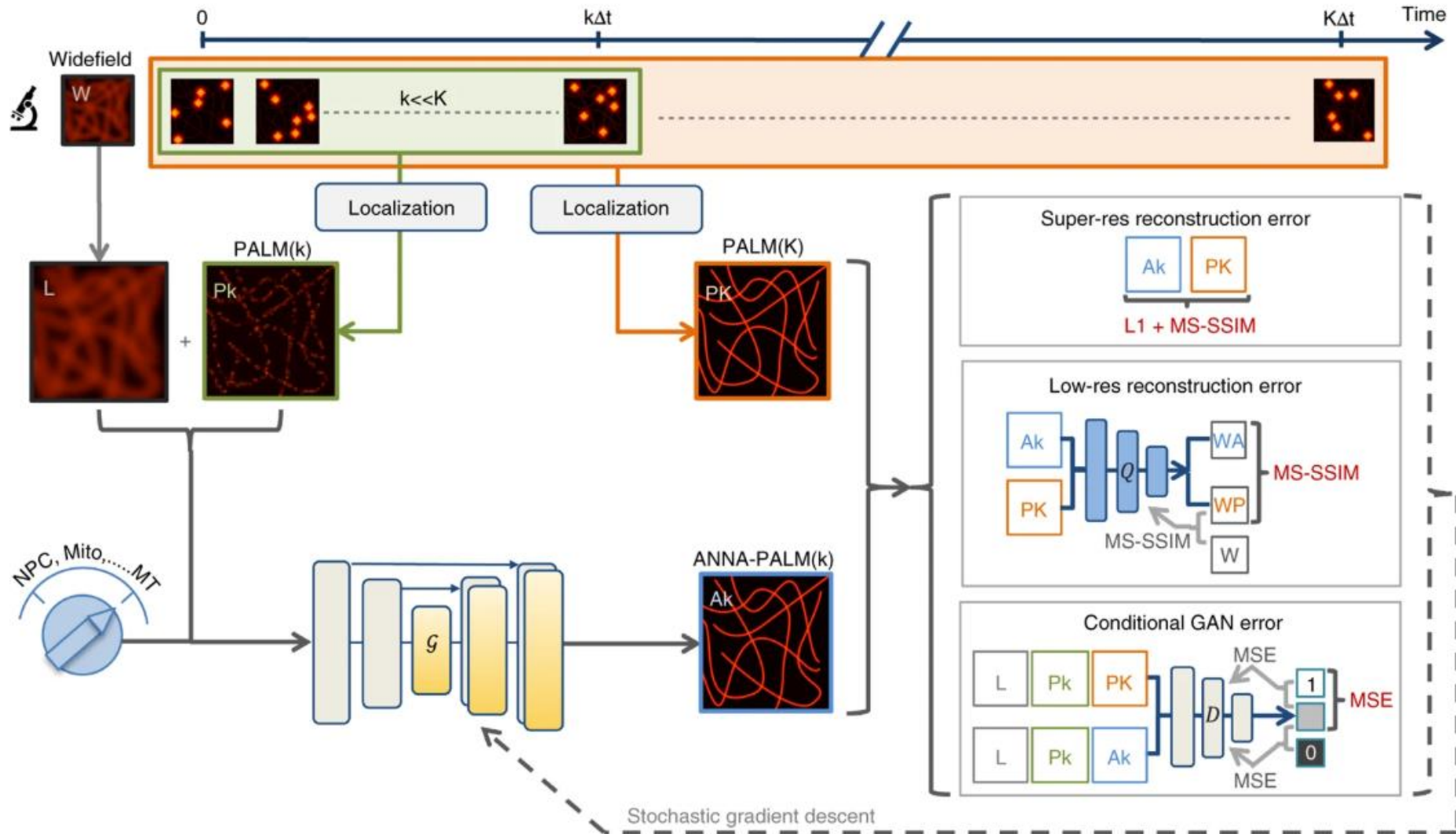
Widefield

Localization



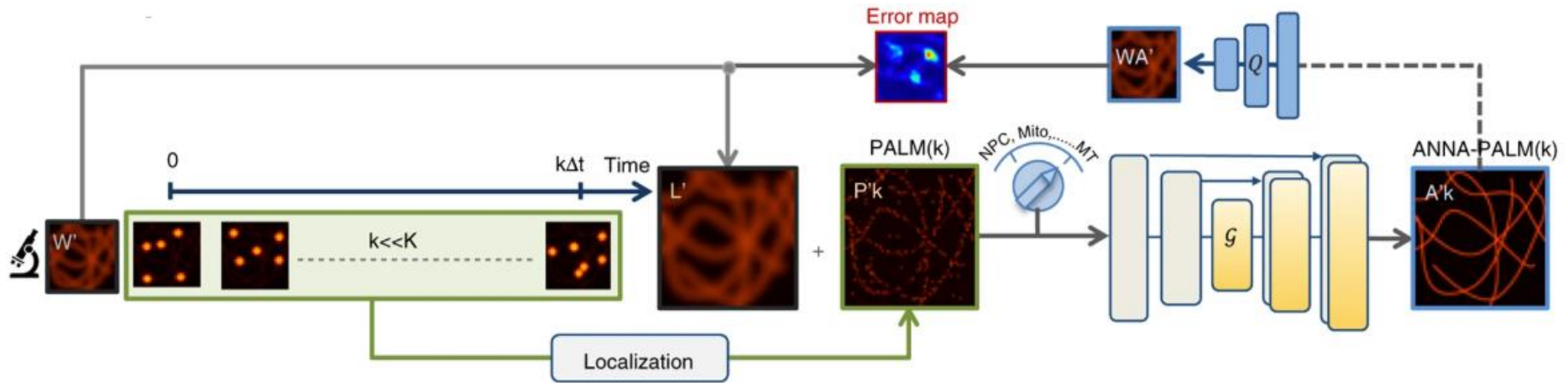


# ANNA-PALM network training



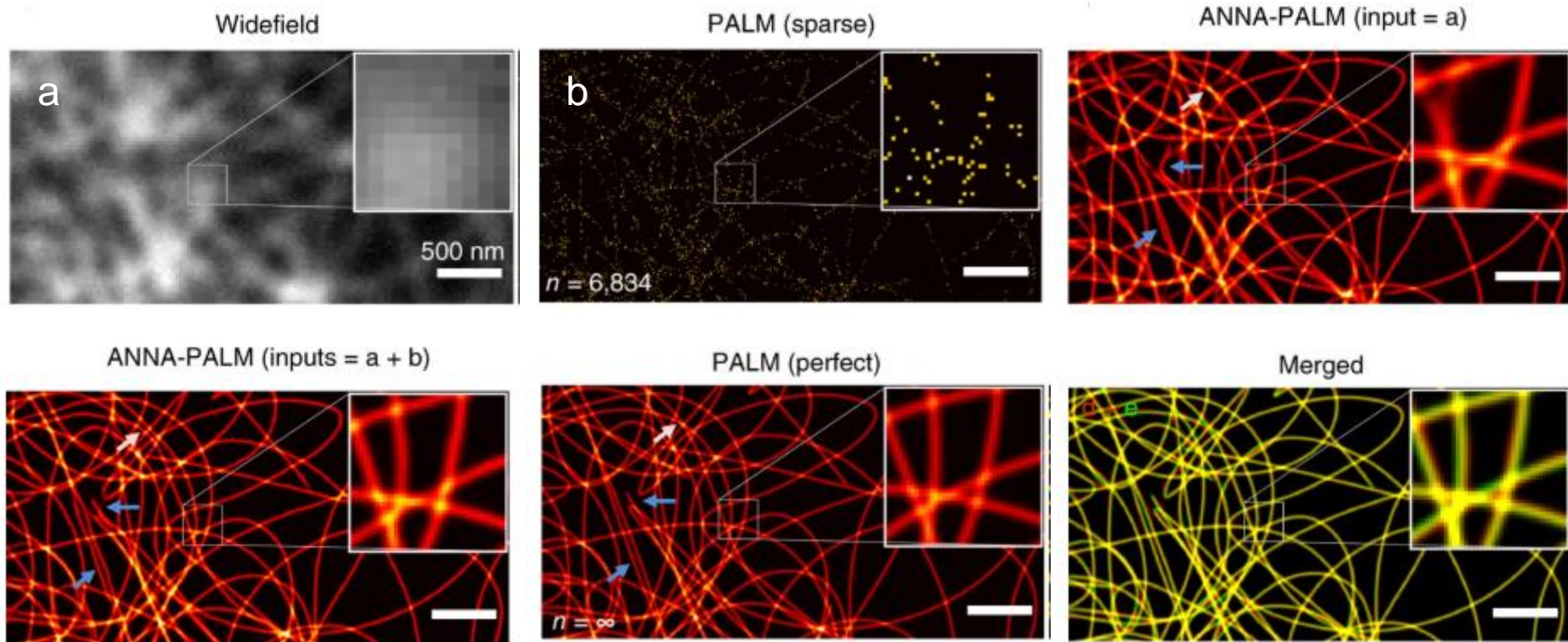


# ANNA-PALM inference



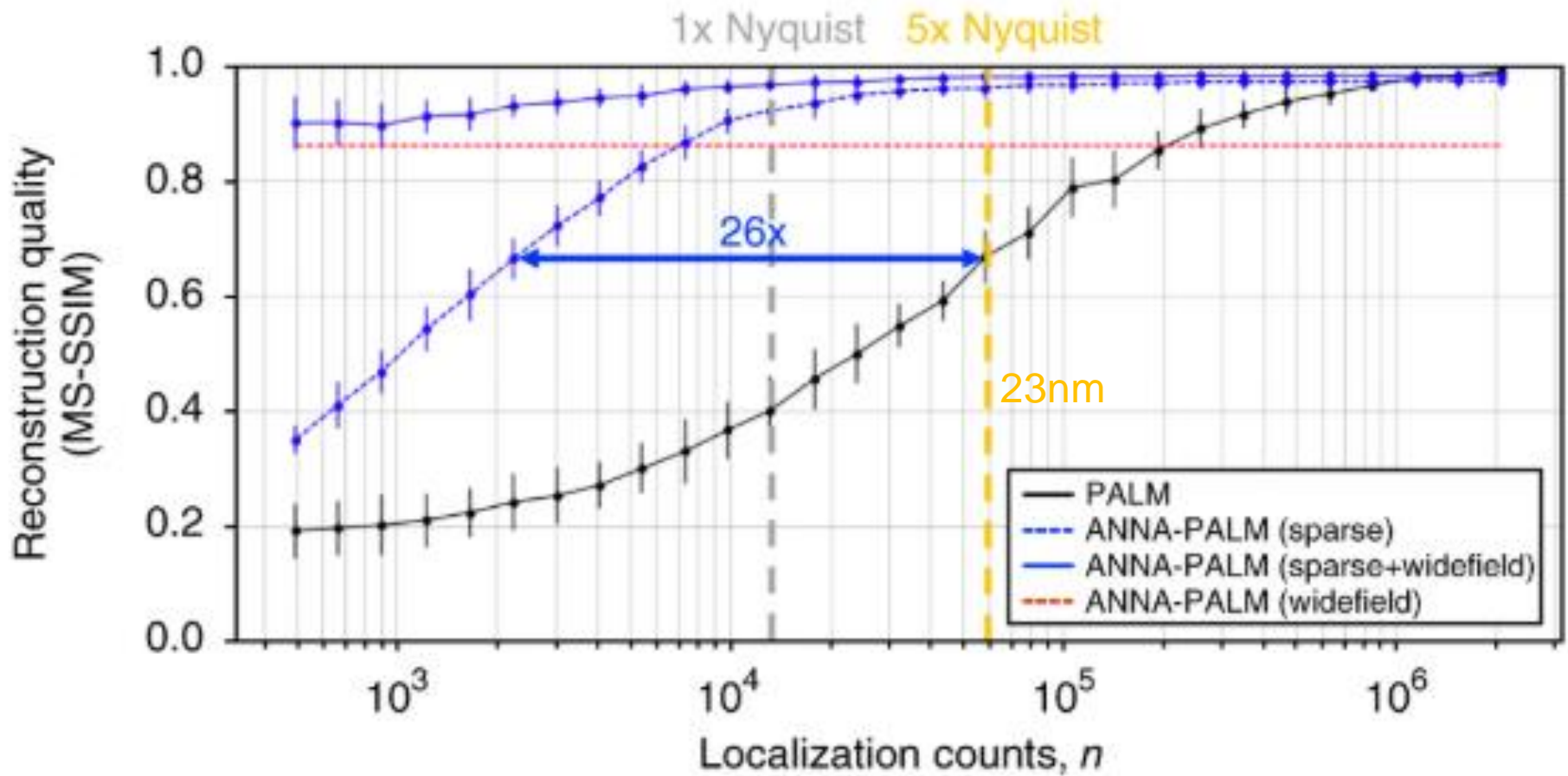


# Validation of ANNA-PALM on simulated images



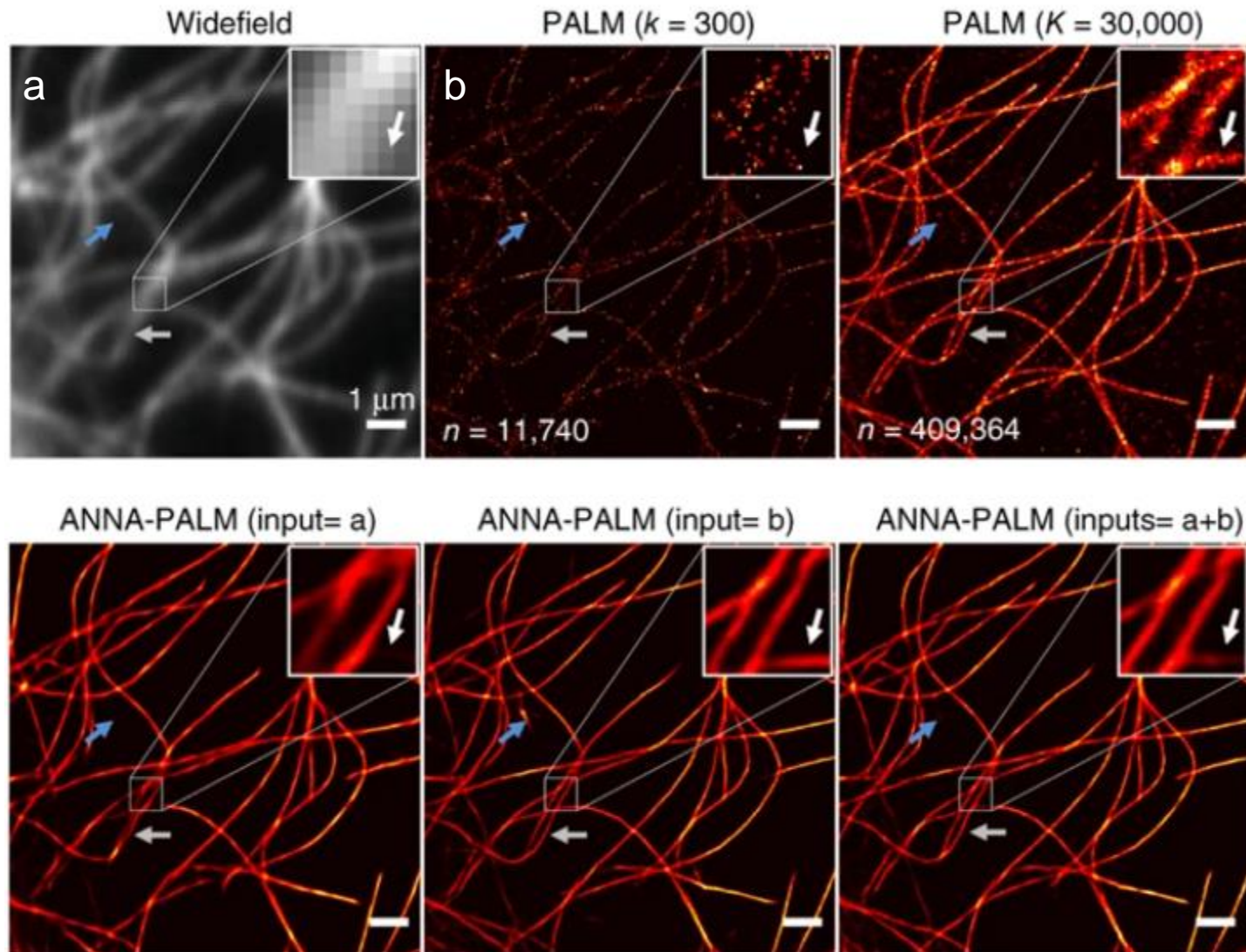


# Validation of ANNA-PALM on simulated images



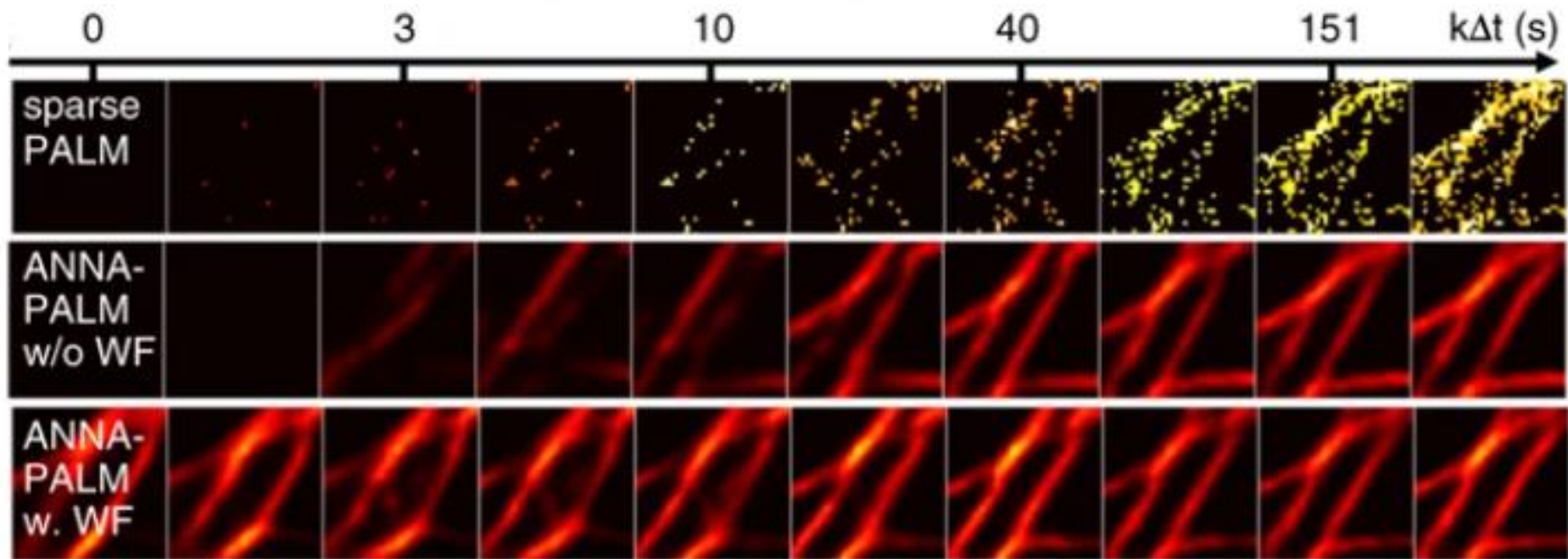


# ANNA-PALM imaging of microtubules



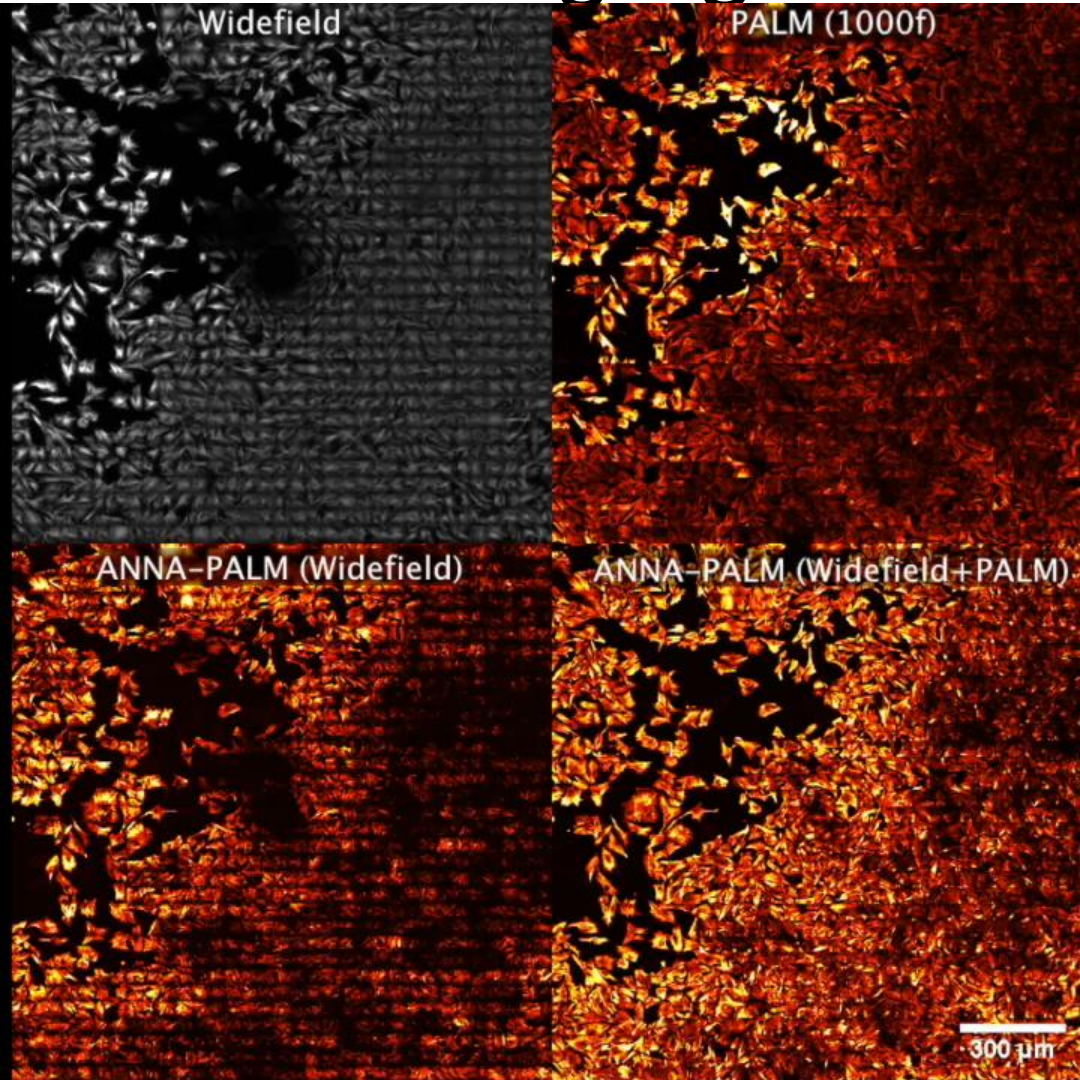


# ANNA-PALM imaging of microtubules



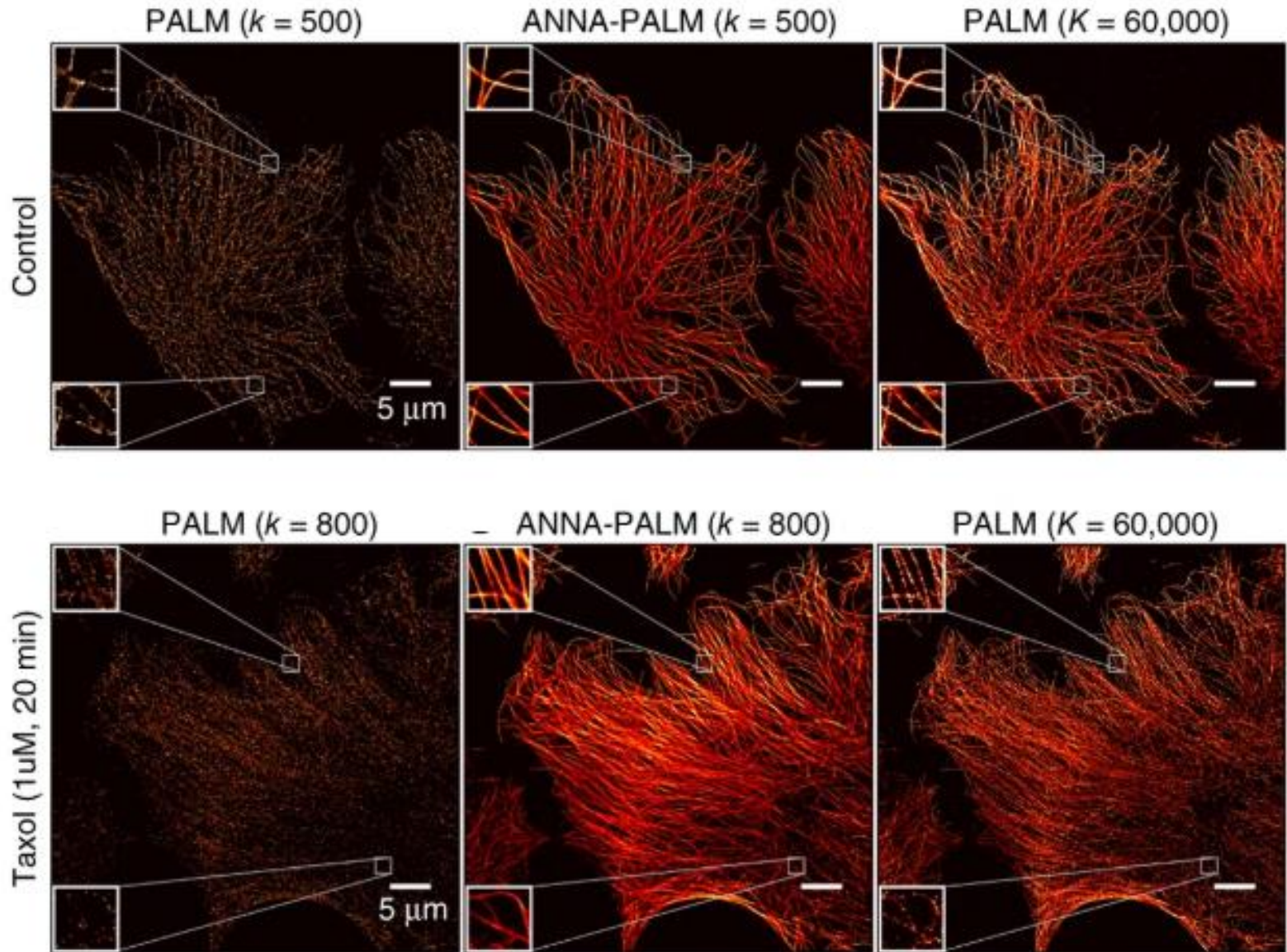


# High-throughput super-resolution imaging





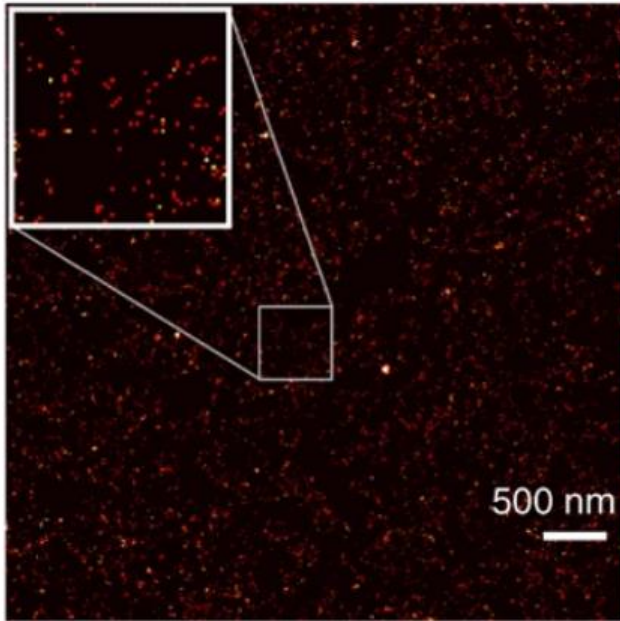
# Robustness to perturbations



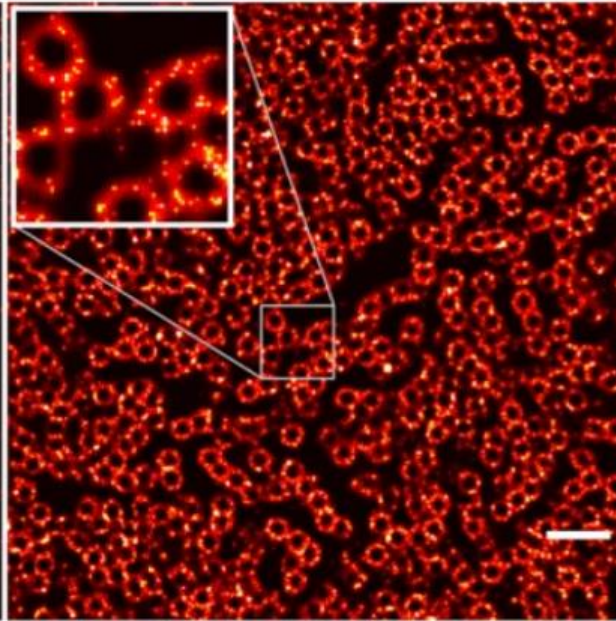


# Training with microtubules and (one) nuclear pores (imaged here)

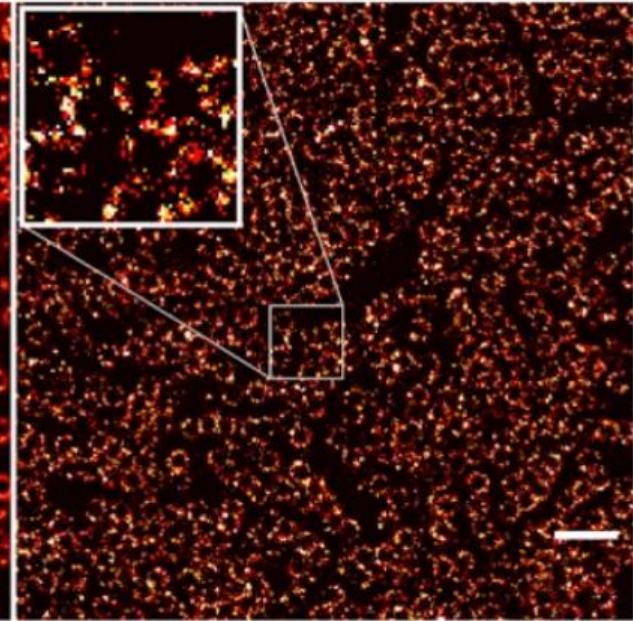
PALM ( $k = 3,000$ )



ANNA-PALM ( $k = 3,000$ )



PALM ( $K = 30,000$ )





# Deep STORM

(the Israeli connection!)

Research Article

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The logo for the journal Optica, featuring the word "optica" in a white serif font on a blue background with abstract light patterns.

## Deep-STORM: super-resolution single-molecule microscopy by deep learning

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# Pick a paper/s and schedule yourself for class presentation

- Preferably from the list I distributed
- Include key + new (ones that I did not read yet) papers
- One (or more – from a common “topic”) paper
- Pair > single
- These paper are LONG! (compared to CS papers)
- Focus on the important stuff, in the context of our course: idea & methodology
- Reading tip: you can (mostly) ignore many biological/experimental details.
  - Example: specific molecule names. Very important, but less in the context of our course.



# Course projects